Mouse IL-12/IL-23p40 ELISA KIT

Catalog Number OKAA00038_96W OKAA00038_48W Size 96 Tests 48 Tests



Mouse IL-12/IL-23p40 ELISA KIT

For the quantitative determination of mouse interleukin 12/23p40 (IL-12/IL-23p40) concentrations in cell culture supernatants, serum, and plasma. This package insert must be read in its entirety before using this product. If you have questions or experience problems with this product, please contact our Technical Support staff. Our scientists commit themselves to providing rapid and effective help.

INTRODUCTION

IL-12, also known as natural killer cell stimulatory factor (NKSF) or cytotoxic lymphocyte maturation factor (CLMF), is a pleiotropic cytokine produced primarily by antigen-presenting cells (monocytes/macrophages, dendritic cells and B lymphocytes). IL-12 has multiple effects on T lymphocytes and natural killer (NK) cells, including the ability to stimulate cytotoxicity, proliferation, cytokine production and Th1 subset development (1, 2). IL-12 is a disulfide-linked, 70 kDa (p70) heterodimeric glycoprotein composed of a unique 35 kDa (p35) subunit and a common 40 kDa (p40) subunit that is also present in IL-23. Monomers of the p40 and p35 subunits by themselves do not have IL-12 activity, but the homodimer of p40 has been shown to bind the IL-12 receptor and is an IL-12 antagonist (3, 4). In cells expressing both p35 and p40 mRNAs, p40 mRNA is expressed to a higher level and free p40 subunits not associated with p35 subunits are secreted together with heterodimeric IL-12 p70 (5). Most of the free p40 subunits secreted by the various human cell lines examined have been found to exist as monomers (1). In the culture supernatants of various activated human monocytes where free p40 is present in vast excess over p70, the levels of p70 measured by bioassays are consistent with those measured using a p70-specific immunoassay, suggesting that p40 monomers are not efficient IL-12 antagonists (1,6). In the mouse system, p40 homodimers are produced in vivo and function as IL-12 antagonists (7). Polymorphisms exist in the mouse IL-12/IL-23 p40 sequence.

PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. A monoclonal antibody specific for IL-12/IL-23p40 has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any IL-12/IL-23p40 present is bound by the immobilized antibody. Following incubation unbound samples are removed during a wash step, and then a detection antibody specific for IL-12/IL-23p40 is added to the wells and binds to the combination of capture antibody- IL-12/IL-23p40 in sample. Following a wash to remove any unbound combination, and enzyme conjugate is added to the wells. Following incubation and wash steps a substrate is added. A colored product is formed in proportion to the amount of IL-12/IL-23p40 present in the sample. The reaction is terminated by addition of acid and absorbance is measured at 450nm. A standard curve is prepared from seven IL-12/IL-23p40 standard dilutions and FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURE.

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IL-12/IL-23p40 sample concentration determined.

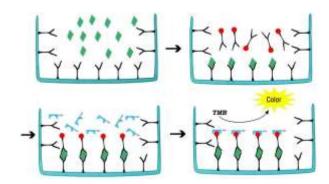


Figure 1: Schematic diagram of the assay

REAGENTS

- Aluminum pouches with a Microwell Plate coated with antibody to mouse IL-12/IL-23p40 (8×12)
- 2. 2 vials mouse IL-12/IL-23p40 Standard lyophilized, 1000 pg/ml upon reconstitution
- 3. 2 vials concentrated Biotin-Conjugate anti-mouse IL-12/IL-23p40 antibody
- 4. 2 vials Streptavidin-HRP solution
- 5. 1 bottle Sample Diluent
- 6. 1 bottle Biotin-Conjugate antibody Diluent
- 7. 1 bottle Streptavidin-HRP Diluent
- 8. 1 bottle Wash Buffer Concentrate 20x (PBS with 1% Tween-20)
- 9. 1 vial Substrate Solution
- 10. 1 vial Stop Solution
- 11. Microwell Plate adhesive films
- 12. Package insert

NOTE: [96 Tests]

STORAGE

Table 1: Storage of the kit

Unopened Kit	Store at 2 - 8° C. Do not use past kit ov	piration date			
Onopened Kit					
Opened/ Reconstituted	Sample Diluent	May be stored for up to 1 month at 2 - 8° C.**			
	Concentrated Biotin-Conjugate				
	Streptavidin-HRP solution				
	Biotin-Conjugate antibody Diluent				
	Streptavidin-HRP Diluent				
	Wash Buffer Concentrate 20x				
	Substrate Solution				
	Stop Solution				
	Standard	Aliquot and store for up to 1			
		month at ≤-20°C.			
Reagents		Avoid repeated freeze-thaw			
		cycles. Diluted standard shall			
		not be reused.			
	Microplate Wells	Return unused wells to the foil			
		pouch containing the			
		desiccant pack, reseal along			
		entire edge of zip-seal. May			
		be stored for up to 1 month at			
		2 - 8° C.**			

^{**}Provided this is within the expiration date of the kit.

THE REQUIRED ITEMS (not provided, but can help to buy):

- 1. Microplate reader (450nm).
- 2. Micro-pipette and tips: 0.5-10, 2-20, 20-200, 200-1000ul.
- 3. 37°C incubator, double-distilled water or deionized water, coordinate paper, graduated cylinder.

PRECAUTIONS FOR USE

1. Store kit reagents between 2°C and 8°C. After use all reagents should

- be immediately returned to cold storage (2°C to 8°C).
- 2. Please perform simple centrifugation to collect the liquid before use.
- 3. To avoid cross contamination, please use disposable pipette tips.
- 4. The Stop Solution suggested for use with this kit is an acid solution. Wear eye, hand, face, and clothing protection when using this material. Avoid contact of skin or mucous membranes with kit reagents or specimens. In the case of contact with skin or eyes wash immediately with water.
- Use clean, dedicated reagent trays for dispensing the washing liquid, conjugate and substrate reagent. Mix all reagents and samples well before use.
- 6. After washing microtiter plate should be fully pat dried. Do not use absorbent paper directly into the enzyme reaction wells.
- 7. Do not mix or substitute reagents with those from other lots or other sources. Do not use kit reagents beyond expiration date on label.
- 8. Each sample, standard, blank and optional control samples should be assayed in duplicate or triplicate.
- Adequate mixing is very important for good result. Use a mini-vortexer at the lowest frequency or Shake by hand at 10min interval when there is no vortexer.
- 10. Avoid microtiter plates drying during the operation.
- 11. Dilute samples at the appropriate multiple, and make the sample values fall within the standard curve. If samples generate values higher than the highest standard, dilute the samples and repeat the assay.
- 12. Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time and temperature, and kit age can cause variation in binding.
- 13. This method can effectively eliminate the interference of the soluble receptors, binding proteins and other factors in biological samples.

SAMPLE COLLECTION AND STORAGE

- 1. **Cell Culture Supernatants** Remove particulates by centrifugation.
- 2. **Serum** Use a serum separator tube (SST) and allow samples to clot

- for 30 minutes before centrifugation for 15 minutes at approximately 1000 x g. Remove serum, avoid hemolysis and high blood lipid samples.
- Plasma Recommended EDTA as an anticoagulant in plasma.
 Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection.
- 4. Assay immediately or aliquot and store samples at -20°C. Avoid repeated freeze-thaw cycles.
- 5. Dilute samples at the appropriate multiple (recommended to do pre-test to determine the dilution factor).

Note: The normal mouse serum or plasma samples are suggested to make a 1:2 dilution.

REAGENT PREPARATION

- 1. Bring all reagents to room temperature before use.
- 2. Wash Buffer Dilute 10 mL of Wash Buffer Concentrate into deionized or distilled water to prepare 200 mL of Wash Buffer. If crystals have formed in the concentrate Wash Buffer, warm to room temperature and mix gently until the crystals have completely dissolved.
- 3. Standard Reconstitute the Standard with 1.0mL of Standard /sample Diluent. This reconstitution produces a stock solution of 1000 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions.
 - Pipette 500 μ L of Sample Diluent into the 500 pg/mL tube and the remaining tubes. Use the stock solution to produce a 2-fold dilution series (below). Mix each tube thoroughly and change pipette tips between each transfer. The 500 pg/mL standard serves as the high standard. The Sample Diluent serves as the zero standard (0 pg/mL).

If you do not run out of re-melting standard, store it at -20 $^{\circ}$ C. Diluted standard shall not be reused.

4. Working solution of Biotin-Conjugate anti-mouse IL-12/IL-23p40 antibody: Make a 1:100 dilution of the concentrated Biotin-Conjugate solution with the Biotin-Conjugate antibody Diluent in a clean plastic tube.

The working solution should be used within one day after dilution.

5. Working solution of Streptavidin-HRP: Make a 1:100 dilution of the

concentrated Streptavidin-HRP solution with the Streptavidin-HRP Diluent in a clean plastic tube.

The working solution should be used within one day after dilution.

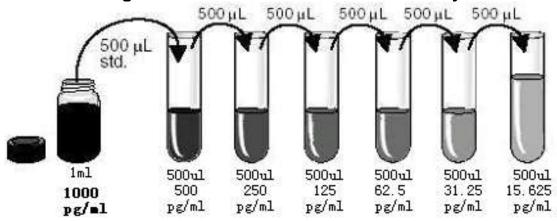


Figure 2: Preparation of IL-12/IL-23p40 standard dilutions

GENERAL ELISA PROTOCOL

- 1. Prepare all reagents and working standards as directed in the previous sections.
- Determine the number of microwell strips required to test the desired number of samples plus appropriate number of wells needed for running blanks and standards. Remove extra microwell strips from holder and store in foil bag with the desiccant provided at 2-8° C sealed tightly.
- 3. Add 100μL of Standard, control, or sample, per well. Cover with the adhesive strip provided. Incubate for 1.5 hours at 37° C.
- 4. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with Wash Buffer (350μL) using a squirt bottle, manifold dispenser or auto-washer. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
- 5. Add 100 μ L of the working solution of Biotin-Conjugate to each well. Cover with a new adhesive strip and incubate 1 hour at 37° C.
- 6. Repeat the aspiration/wash as in step 4. FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURE.

- 7. Add 100 μ L of the working solution of Streptavidin-HRP to each well. Cover with a new adhesive strip and incubate for 30 minutes at 37° C Avoid placing the plate in direct light.
- 8. Repeat the aspiration/wash as in step 4.
- 9. Add 100 μ L of Substrate Solution to each well. Incubate for 10-20 minutes at 37° C. Avoid placing the plate in direct light.
- 10. Add 100 μ L of Stop Solution to each well. Gently tap the plate to ensure thorough mixing.
- 11. Determine the optical density of each well immediately, using a microplate reader set to 450 nm.(optionally 630nm as the reference wave length;610-650nm is acceptable)

ASSAY PROCEDURE SUMMARY

Prepare all reagents and standards as directed Add 100µl standard or samples to each well, incubate 90 minutes, 37° C Aspirate and wash 4 times Add 100µl working solution of Biotin-Conjugate anti-mouse IL-12/IL-23p40 antibody to each well, incubate 60 minutes, 37° C \prod Aspirate and wash 4 times Add 100µl working solution of Streptavidin-HRP to each well, incubate 30 minutes, 37° C Add 100µl Substrate solution to each well, incubate 10-20 minutes,37° C.Protect from light. \int Add 100µl Stop solution to each well. Read at 450nm

Figure 3: Assay procedure summary

within 30 minutes.

TECHNICAL HINTS

- 1. When mixing or reconstituting protein solutions, always avoid foaming.
- 2. To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.

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- 3. To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- 4. Substrate Solution should remain colorless until added to the plate. Stop Solution should be added to the plate in the same order as the Substrate Solution. Keep Substrate Solution protected from light. Substrate Solution should change from colorless to gradations of blue.
- 5. A standard curve should be generated for each set of samples assayed. According to the content of tested factors in the sample, appropriate diluted or concentrated samples, it is best to do pre-experiment.

CALCULATION OF RESULTS

- 1. Average the duplicate readings for each standard, control, and sample and subtract the average zero standard optical density.
- 2. Create a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph.
- 3. The data may be linearized by plotting the log of the IL-12/IL-23p40 concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.
- 4. This standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

Table 2: Typical data using the IL-12/IL-23p40 ELISA (Measuring wavelength:450nm, Reference wavelength:630nm)

Standard (pg/ml)	OD.	OD.	Average	Corrected
0	0.033	0.029	0.031	
7.8	0.108	0.105	0.106	0.110
15.625	0.177	0.176	0.176	0.181
31.25	0.304	0.300	0.302	0.308
62.5	0.510	0.505	0.508	0.524
125	0.926	0.926	0.926	0.878
250	1.381	1.379	1.380	1.412
500	2.136	2.134	2.135	2.128

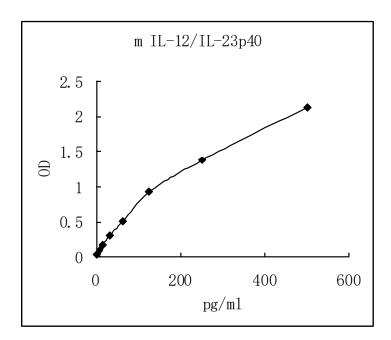


Figure 4: Representative standard curve for IL-12/IL-23p40 ELISA. IL-12/IL-23p40 was diluted in serial two-fold steps in Sample Diluent.

Do not use this standard curve to derive test results. A standard curve must be run for each group of microwell strips assayed.

SENSITIVITY, SPECIFICITY AND REPEATABILITY

- 1. **REPEATABILITY**: The coefficient of variation of both intra-assay and inter-assay were less than 10%.
- 2. **SENSITIVITY**: The minimum detectable dose was 4pg/mL.
- 3. **SPECIFICITY:** This assay recognizes both natural and recombinant mouse IL-12/IL-23p40. The factors listed below were prepared at 50ng/ml in Standard /sample Diluent and assayed for cross-reactivity and no significant cross-reactivity or interference was observed.

Table 3: Factors assayed for cross-reactivity

Recombinant human	Recombinant mouse	
IL-12/IL-23p40 (dimer)	IL-1α	
IL-12p70	IL-1β	
	IL-3	
	IL-4	
	IL-5	
	IL-6	

REFERENCES

- 1. Wolf, S.F. et al. (1994) Stem Cells . 12:154.
- 2. Trinchieri, G. and F. Gerosa (1996) J. Leuk. Biol. 59:505.
- 3. Gillessen, S. et al. (1995) Eur. J. Immunol. 25:200.
- 4. Ling, P. et al. (1995) J. Immunol. 154:116.
- 5. D'Andrea, A. et al. (1992) J. Exp. Med. 176:1387.
- 6. Tsang, M.L-S. and J.A. Weatherbee (1996) J. of NIH Research 8:68.
- 7. Heinzel, F.P. et al. (1997) J. Immunol. 158:4381.

RELATED PRODUCTS

Table 3: Related products

Product name	Catalog Number	Size
Mouse IL-2 ELISA Kit	OKAA00031	48T/96T
Mouse GM-CSF ELISA kit	OKAA00032	48T/96T
Mouse IL-1a ELISA kit	OKAA00033	48T/96T
Mouse IL-1β ELISA Kit	OKAA00034	48T/96T
Mouse IL-4 ELISA Kit	OKAA00035	48T/96T
Mouse IL-5 ELISA kit	OKAA00036	48T/96T
Mouse IL-10 ELISA Kit	OKAA00037	48T/96T
Mouse IL-12 IL-23p40 ELISA kit	OKAA00038	48T/96T
Mouse IL-12p70 ELISA Kit	OKAA00039	48T/96T
Mouse IL-25 IL-17E ELISA kit	OKAA00040	48T/96T
Mouse IL-6 ELISA Kit	OKAA00043	48T/96T