

DYRK1A ELISA Kit (Human) (OKCD01050) Lot# KD5058 Instructions for use

For the quantitative measurement of DYRK1A in tissue homogenates, cell lysates and other biological fluids.

Lot to lot variations can occur. Refer to the manual provided with the kit.

This product is intended for research use only.



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1. Background

Principle

Aviva Systems Biology DYRK1A ELISA Kit (Human) (OKCD01050) is based on standard sandwich enzymelinked immuno-sorbent assay technology. An antibody specific for DYRK1A has been pre-coated onto a 96-wellplate (12 x 8 Well Strips). Standards or test samples are added to the wells, incubated and removed. A biotinylated detector antibody specific for DYRK1A is added, incubated and followed by washing. Avidin-Peroxidase Conjugate is then added, incubated and unbound conjugate is washed away. An enzymatic reaction is produced through the addition of TMB substrate which is catalyzed by HRP generating a blue color product that changes to yellow after adding acidic stop solution. The density of yellow coloration is read by absorbance at 450 nm and is quantitatively proportional to the amount of sample DYRK1A captured in well.

Background

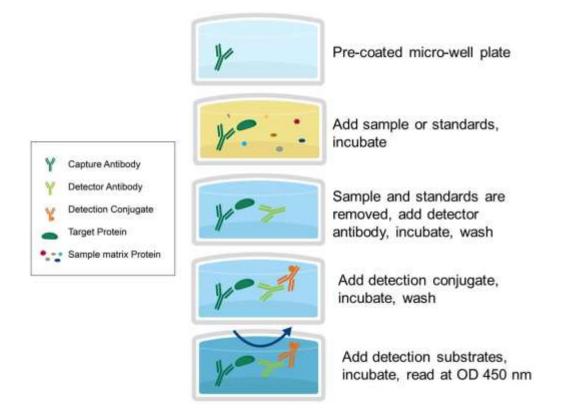
May play a role in a signaling pathway regulating nuclear functions of cell proliferation. Modulates alternative splicing by phosphorylating the splice factor SRSF6 .. Phosphorylates serine, threonine and tyrosine residues in its sequence and in exogenous substrates such as CRY2, FOXO1, SRSF6 and SIRT1. Exhibits a sugstrate preference for proline at position P+1 and arginine at position P-3.

General Specifications

General Specifications					
Range	0.312-20 ng/mL				
LOD	< 0.116 ng/mL (Derived by linear regression of OD ₄₅₀ of the Mean Blank + 2xSD)				
	Human Dual specificity tyrosine-phosphorylation-regulated kinase 1A				
	UniProt ID: Q13627				
Specificity	<u>GeneID</u> : 1859				
	<u>Target Alias</u> : Dual specificity tyrosine-phosphorylation-regulated kinase 1A, Dual specificity YAK1-related kinase, DYRK, DYRK1, hMNB, HP86, MNB, MNBH, Protein kinase minibrain homolog				
Cross-Reactivity	No detectable cross-reactivity with other relevant proteins				



2. Assay Summary



3. Storage and Stability

• Open kit immediately upon receipt. Store components at -20°C (NOTE: exceptions below) for 6 months or until expiration date. Avoid any freeze/thaw cycles.

4. Kit Components

•The following reagents are the provided contents of the kit.

Description	Quantity	Storage Conditions
Anti-DYRK1A Microplate	96 Wells (12 x 8 Well strips)	
DYRK1A Lyophilized Standard	2 x 20 ng	
100X Biotinylated DYRK1A Detector Antibody	1 x 120 μL	-20°C for 6 months
100X Avidin-HRP Conjugate	1 x 120 μL	
Standard Diluent	1 x 20 mL	
Detector Antibody Diluent	1 x 12 mL	
Conjugate Diluent	1 x 12 mL	Store at 4°C for 6
30X Wash Buffer	1 x 20 mL	months
Stop Solution	1 x 6 mL	
TMB Substrate	1 x 9 mL	



5. Precautions

- Read instructions fully prior to beginning use of the assay kit.
- Any deviations or modifications from the described method or use of other reagents could result in a reduction of performance.
- Reduce exposure to potentially harmful substances by wearing personal protective lab equipment including lab coats, gloves and glasses.
- For information on hazardous substances included in the kit please refer to the Material Safety Data Sheet (MSDS).
- Kit cannot be used beyond the expiration date on the label.

6. Required Materials Not Supplied

- Microplate reader capable of reading absorbance at 450 nm.
- Automated plate washer (optional).
- Pipettes capable of precisely dispensing 0.5 µL through 1 mL volumes of aqueous solutions.
- Pipettes or volumetric glassware capable of precisely measuring 1 mL through 100 mL of aqueous solutions.
- New, clean tubes and/or micro-centrifuge tubes for the preparation of standards or samples.
- · Absorbent paper or paper toweling.
- Distilled or deionized ultrapure water.
- 37°C Incubator (optional)

7. Technical Application Tips

- Do not mix or substitute components from other kits.
- To ensure the validity of experimental operation, it is recommended that pilot experiments using standards and a small selection of sample dilutions to ensure optimal dilution range for quantitation.
- Samples exhibiting OD measurements higher than the highest standard should be diluted further in the appropriate sample dilution buffers.
- Prior to using the kit, briefly spin component tubes to collect all reagents at the bottom.
- Replicate wells are recommended for standards and samples.
- Cover microplate while incubating to prevent evaporation.
- Do not allow the microplate wells dry at any point during the assay procedure.
- Do not reuse tips or tube to prevent cross contamination.
- Avoid causing bubbles or foaming when pipetting, mixing or reconstituting.
- Completely remove of all liquids when washing to prevent cross contamination.
- Prepare reagents immediately prior to use and do not store, with the exception of the top standard.
- Equilibrate all materials to ambient room temperature prior to use (standards exception).
- For optimal results for inter- and intra-assay consistency, equilibrate all materials to room temperature prior to performing assay (standards exception) and perform all incubations at 37°C.
- Pipetting less than 1 µL is not recommended for optimal assay accuracy.
- Once the procedure has been started, all steps should be completed without interruption. Ensure that all reagents, materials and devices are ready at the appropriate time.
- Incubation times will affect results. All wells should be handled in the same sequential order and time intervals for optimal results.
- Samples containing precipitates, fibrin strands or bilirubin, or are hemolytic or lipemic might cause inaccurate results due to interfering factors.
- TMB Substrate is easily contaminated and should be colorless or light blue until added to plate. Handle carefully and protect from light.



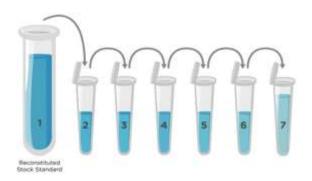
8. Reagent Preparation

Equilibrate all materials to room temperature prior to use and use immediately.

8.1 <u>Human DYRK1A Assay Standards</u>

- 8.1.1 Prepare the DYRK1A standards no greater than 2 hours prior to performing experiment. Standards should be held on ice until use in the experiment.
- 8.1.2 Reconstitute one vial of the provided 20 ng Lyophilized DYRK1A Standard for each experiment. Prepare the stock 20 ng/mL Standard by reconstituting one tube of Lyophilized DYRK1A Standard as follows:
 - 8.1.2.1 Gently spin or tap the vial at 6,000 10,000 rpm for 30 seconds to collect all material at the bottom.
 - 8.1.2.2 Add 1 mL of Standard Diluent to the vial.
 - 8.1.2.3 Seal the vial then mix gently and thoroughly.
 - 8.1.2.4 Leave the vial at ambient temperature for 15 minutes.
- 8.1.3 Prepare a set of seven serially diluted standards as follows:
 - 8.1.3.1 Label tubes with numbers 2 8.
 - 8.1.3.2 Use the undiluted 20 ng/mL DYRK1A Standard as the high standard point (Tube #1).
 - 8.1.3.3 Add 300 μ L of **Standard Diluent** to Tube #'s 2 8.
 - 8.1.3.4 Prepare **Standard #2** by adding 300 μ L of 20 ng/mL **DYRK1A** (Tube #1) to Tube #2. Mix gently and thoroughly.
 - 8.1.3.5 Prepare **Standard #3** by adding 300 μ L of **Standard #2** from Tube #2 to Tube #3. Mix gently and thoroughly.
 - 8.1.3.6 Prepare further serial dilutions through Tube #7. Reference the table below as a guide for serial dilution scheme.
 - 8.1.3.7 Tube #8 is a blank standard (only **Standard Diluent**), which should be included with every experiment.

Standard Number (Tube)	Standard To Dilute	Volume Standard to Dilute (µL)	Volume Standard Diluent (μL)	Total Volume (μL)	Final Concentration
1	20 ng/mL Reconstituted DYRK1A Standard	NA	1,000	1,000	20 ng/mL
2	20 ng/mL	300	300	600	10 ng/mL
3	10 ng/mL	300	300	600	5.0 ng/mL
4	5.0 ng/mL	300	300	600	2.5 ng/mL
5	2.5 ng/mL	300	300	600	1.25 ng/mL
6	1.25 ng/mL	300	300	600	0.625 ng/mL
7	0.625 ng/mL	300	300	600	0.312 ng/mL
8	NA	0	300	300	0.0 (Blank)





8.2 1X Biotinylated DYRK1A Detector Antibody

- Prepare the **1X Biotinylated DYRK1A Detector Antibody** immediately prior to use by diluting the **100X Biotinylated DYRK1A Detector Antibody** 1:100 with **Detector Antibody Diluent**.
- 8.2.2 For each well strip to be used in the experiment (8-wells) prepare 1,000 μL by adding 10 μL of **100X Biotinylated DYRK1A Detector Antibody** to 990 μL **Detector Antibody Diluent**.
- 8.2.3 Mix thoroughly and gently. Hold no longer than 2 hours prior to using in procedure. Do not store at 1X concentration for future use.

8.3 1X HRP-Avidin Conjugate

- 8.3.1 Prepare the **1X Avidin-HRP Conjugate** immediately prior to use by diluting the **100X Avidin-HRP Conjugate** 1:100 with **Conjugate Diluent**.
- 8.3.2 For each well strip to be used in the experiment (8-wells) prepare 1,000 μL by adding 10 μL of **100X Avidin-HRP Conjugate** to 990 μL **Conjugate Diluent**.
- 8.3.3 Mix thoroughly and gently. Hold no longer than 2 hours prior to using in procedure. Do not store at 1X concentration for future use.

8.4 1X Wash Buffer

- 8.4.1 If crystals have formed in the **30X Wash Buffer** concentrate, equilibrate to room temperature and mix gently until crystals have completely dissolved.
- 8.4.2 Add the entire 20 mL contents of the **30X Wash Buffer** bottle to 580 mL of ultra-pure water to a clean > 1,000 mL bottle or other vessel.
- 8.4.3 Seal and mix gently by inversion. Avoid foaming or bubbles.
- 8.4.4 Store the **1X Wash Buffer** at room temperature until ready to use in the procedure. Store the prepared **1X Wash Buffer** at 4°C for no longer than 1 week. Do not freeze.

8.5 Microplate Preparation

- Micro-plates are provided ready to use and do not require rinsing or blocking.
- Unused well strips should be returned to the original packaging, sealed and stored at 4°C.
- Equilibrate microplates to ambient temperatures prior to opening to reduce potential condensation.



9. Sample Preparation

9.1 Sample Preparation and Storage

- Store samples to be assayed at 2-8°C for 24 hours prior being assayed.
- For long term storage, aliquot and freeze samples at -20°C. Avoid repeated freeze-thaw cycles.
- Samples not indicated in the manual must be tested to determine if the kit is valid.
- · Prepare samples as follows:
 - Tissue Homogenates Rinse 100 mg tissue with 1X PBS then homogenize in 1 mL of 1X PBS and store overnight at -20°C. Perform two freeze-thaw cycles to break the cell membranes then centrifuge for 5 minutes at 5,000 x g, 2-8°C. Remove the supernatant and assay immediately. Alternatively, aliquot and store samples at -20°C or -80°C. Centrifuge the sample again after thawing before the assay. Avoid repeated freeze-thaw cycles.
 - Cell Lysates Adherent cells should be detached with trypsin and then collected by centrifugation (suspension cells can be collected by centrifugation directly). Wash cells three times in cold PBS. Resuspend cells in PBS (1×) and ultrasonicate the cells 4 times (or freeze cells at ≤ -20°C. Thaw cells with gentle mixing. Repeat the freeze/thaw cycle 3 times.) Centrifuge at 1,500 x g for 10 minutes at 2 8°C to remove cellular debris.
 - Other biological fluids Centrifuge samples for 20 minutes at 1000 x g. Remove particulates and assay immediately or store samples in aliquot at -20°C or -80°C. Avoid repeated freeze/thaw cycles.

9.2 Sample Dilution

Target protein concentration must be estimated and appropriate sample dilution selected such that the final target protein concentration falls near the middle of the assay linear dynamic range. Samples exhibiting saturation should be further diluted.

- Dilute samples using Standard Diluent.
- Mix diluted samples gently and thoroughly.
- Pipetting less than 2 µL is not recommended for optimal assay accuracy.



10. Assay Procedure

- Equilibrate all reagents and materials to ambient room temperature prior to use in the procedure.
- Optimal results for intra- and inter-assay reproducibility will be obtained when performing incubation steps at 37°C as indicated below.
- **10.1** Determine the required number of wells and return any remaining unused wells and desiccant to the pouch.
- 10.2 Add 100 μL of serially titrated standards, diluted samples or blank into wells of the Anti-DYRK1A Microplate. At least two replicates of each standard, sample or blank is recommended.
- **10.3** Cover the plate with the plate sealer and incubate at 37°C for 60 minutes.
- **10.4** Remove the plate sealer and discard the liquid in the wells by rigorously flicking into an acceptable waste receptacle or aspiration.
- **10.5** Gently blot any remaining liquid from the wells by tapping inverted on the benchtop onto paper toweling. Do not allow the wells to completely dry at any time.
- 10.6 Add 100 µL of prepared 1X Biotinylated DYRK1A Detector Antibody to each well.
- **10.7** Cover with the plate sealer and incubate at 37°C for 60 minutes.
- 10.8 Discard the liquid in the wells by rigorously flicking into an acceptable waste receptacle or aspiration.
- **10.9** Gently blot any remaining liquid from the wells by tapping inverted on the benchtop onto paper toweling. Do not allow the wells to completely dry at any time.
- 10.10 Wash plate 3 times with 1X Wash Buffer as follows:
 - 10.10.1 Add 300 µL of 1X Wash Buffer to each assay well.
 - 10.10.2 Incubate for 1 minute.
 - 10.10.3 Discard the liquid in the wells by rigorously flicking into an acceptable waste receptacle.
 - 10.10.4 Gently blot any remaining liquid from the wells by tapping inverted on the benchtop onto paper toweling. Do not allow the wells to completely dry at any time.
 - 10.10.5 Repeat steps 10.10.1 through 10.10.4 **two** more times.
- **10.11** Add 100 μL of prepared **1X Avidin-HRP Conjugate** into each well, cover with plate sealer and incubate at 37°C for 30 minutes.
- 10.12 Discard the liquid in the wells by rigorously flicking into an acceptable waste receptacle or aspiration.
- **10.13** Gently blot any remaining liquid from the wells by tapping inverted on the benchtop onto paper toweling. Do not allow the wells to completely dry at any time.
- 10.14 Wash plate 5 times with 1X Wash Buffer as in Step 10.10.
- 10.15 Add 90 μL of TMB Substrate to each well, cover with plate sealer and incubate at 37°C in the dark for 10-20 minutes. Wells should change to gradations of blue. If the color is too deep, reduce the incubation time.
 - (NOTE: optimal incubation time must be determined by the user. Optimal development can be visualized by blue shading in the top four standard wells, while the remaining standards are still clear.)
- **10.16** Add 50 μL of **Stop Solution** to each well. Well color should change to yellow immediately. Add the **Stop Solution** in the same well order as done for the **TMB Substrate**.
- **10.17** Read the O.D. absorbance at 450 nm with a standard microplate reader within 5 minutes of stopping the reaction in step 10.16. If wavelength correction is available, set to 540 nm or 570 nm.



11. Calculation of Results

For analysis of the assay results, calculate the **Relative OD**₄₅₀ for each test or standard well as follows:

(Relative
$$OD_{450}$$
) = (Well OD_{450}) – (Mean Blank Well OD_{450})

The standard curve is generated by plotting the mean replicate **Relative OD**₄₅₀ of each standard serial dilution point vs. the respective standard concentration. The **DYRK1A** concentration contained in the samples can be interpolated by using linear regression of each mean sample **Relative OD**₄₅₀ against the standard curve. This is best achieved using curve fitting software.

Note: If wavelength correction readings were available, subtract the readings at 540 nm or 570 nm from the readings at 450 nm. This may provide greater reading accuracy.

Note: If the samples measured were diluted, multiply the derived mean sample concentration by the dilution factor for a final sample concentration.

12. Typical Expected Data

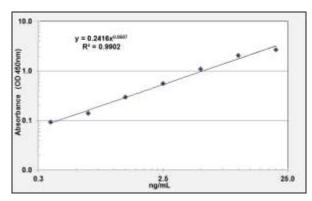
12.1 Reproducibility

Intra-assay Precision: 3 samples with known low, middle and high levels DYRK1A were tested with 20 replicates on one plate, respectively. Inter-assay Precision: 3 samples with known low, middle and high level DYRK1A were tested on 3 different plates, 8 replicates in each plate.

Sample	Intra-Assay			Inter-Assay		
Sample	1	2	3	1	2	3
Sample	1	2	3	1	2	3
n	20	20	20	24	24	24
Mean (ng/ml)	0.98	4.71	9.43	0.87	4.86	9.58
SD	0.068	0.301	0.500	0.057	0.282	0.498
CV (%)	6.9	6.4	5.3	6.5	5.8	5.2

12.2 Typical standard curve

This standard curve is for demonstration purposes only. An assay specific standard curve should be performed with each assay.



ng/mL	Absorbance		Mean	Blank	
Hg/IIIL	Rep 1	Rep 2	Absorbance	Subtracted	
20	2.622	2.614	2.618	2.581	
10	2.017	2.003	2.010	1.973	
5.0	1.084	1.072	1.078	1.041	
2.5	0.554	0.566	0.560	0.523	
1.25	0.297	0.305	0.301	0.264	
0.625	0.138	0.142	0.140	0.103	
0.312	0.092	0.086	0.089	0.052	
Blank	0.036	0.038	0.037	NA	



13. Technical Resources

Technical Support:

For optimal service please be prepared to supply the lot number of the kit used.

<u>USA</u>

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