



**TRU** **Rabbit Anti-Free Testosterone Antibody Coated Microwell Plate-Break Apart Wells - Ready To Use.**

Contents: One 96 well (12x8) polyclonal antibody-coated microwell plate in a resealable pouch with desiccant.

Storage: Refrigerate at 2-8°C

Stability: 12 months or as indicated on label.

AG	HRP	CONC
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**Free Testosterone-Horse Radish Peroxidase (HRP)  
Conjugate Concentrate – X50**

Contents: Free Testosterone-HRP conjugate in a protein-based buffer with a non-mercury preservative.

Volume: 300 µl/vial

Storage: Refrigerate at 2-8°C

Stability: 12 months or as indicated on label.

Preparation: Dilute 1:50 in assay buffer before use (eg. 40 µl of HRP in 2 ml of assay buffer). If the whole plate is to be used dilute 240 µl of HRP in 12 ml of assay buffer. Discard any that is left over.

**CAL**   **N**   **Free Testosterone Calibrators - Ready To Use. N = 0 to 5**

Contents: six vials containing testosterone in a human serum-based buffer with a non-mercury preservative. Prepared by spiking serum with a precise quantity of testosterone equivalent to approximately 0, 0.25, 1.02, 5.5, 25 and 125 pg/ml of free testosterone.

\*Listed below are approximate concentrations, please refer to vial labels for exact concentrations.

Calibrator	Concentration	Volume
Calibrator 0	0 pg/ml	0.5 ml
Calibrator 1	0.25 pg/ml	0.5 ml
Calibrator 2	1.02 pg/ml	0.5 ml
Calibrator 3	5.5 pg/ml	0.5 ml
Calibrator 4	25 pg/ml	0.5 ml
Calibrator 5	125 pg/ml	0.5 ml

**Storage:** Refrigerate at 2-8°C  
**Stability:** 12 months in unopened vials or as indicated on label. Once opened, the calibrators should be used within 14 days or aliquoted and stored frozen. Avoid multiple freezing and thawing cycles.

**CONTROL**      **Control** - Ready To Use.

Contents: One vial containing testosterone in a human serum-based buffer with a non-mercury preservative. Prepared by spiking serum with a precise quantity of testosterone. Refer to vial label for expected value and acceptable range.

Volume: 0.5 ml/vial

Storage: Refrigerate at 2-8 °C

Stability: 12 months in unopened vial or as indicated on label. Once opened, the control should be used within 14 days or aliquoted and stored frozen. Avoid multiple freezing and thawing cycles.

WASH	SOLN	CONC	Wash Buffer Concentrate - <b>X10</b>
<p>Contents: One bottle containing buffer with a non-ionic detergent and a non-mercury preservative.            Volume: 50 ml/bottle            Storage: Refrigerate at 2-8°C            Stability: 12 months or as indicated on label.</p> <p>Preparation: Dilute 1:10 in distilled or deionized water before use. If the whole plate is to be used dilute 50 ml of the wash buffer concentrate in 450 ml of water.</p>			

ASS	BUF
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**Assay Buffer - Ready To Use.**

Contents: One bottle containing a protein-based buffer with a non-mercury preservative.

Volume: 15 ml/vial

Storage: Refrigerate at 2-8°C

Stability: 12 months or as indicated on label.

<b>CHROM</b>	<b>TMB</b>	<b>TMB Substrate</b> - Ready To Use.
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Contents: One bottle containing tetramethylbenzidine and hydrogen peroxide in a non-DMF or DMSO containing buffer.  
Volume: 16 ml/bottle  
Storage: Refrigerate at 2-8°C  
Stability: 12 months or as indicated on label.

<b>STOP</b>	<b>SOLN</b>	<b>Stopping Solution - Ready To Use.</b>
Contents: One vial containing 1M sulfuric acid.		
Volume: 6 ml/vial		
Storage: Refrigerate at 2-8°C		
Stability: 12 months or as indicated on label.		

**ASSAY PROCEDURE:**  
**Specimen Pretreatment:**  
*None.*

All reagents must reach room temperature before use. Calibrators, controls and specimen samples should be assayed in duplicate. Once the procedure has been started, all steps should be completed without interruption.

1. Prepare working solutions of the Free Testosterone-HRP conjugate and wash buffer.
2. Remove the required number of microwell strips. Reseal the bag and return any unused strips to the refrigerator.
3. Pipette 25 µl of each calibrator, control and specimen sample into correspondingly labelled wells in duplicate.
4. Pipette 100 µl of the conjugate working solution into each well (We recommend using a multichannel pipette).
5. Gently shake the plate for 10 seconds.
6. Incubate the plate at 37°C for 1 hour.
7. Wash the wells 3 times with 300 µl of diluted wash buffer per well and tap the plate firmly against absorbent paper to ensure that it is dry (the use of a washer is recommended).
8. Pipette 150 µl of TMB substrate into each well at timed intervals.
9. Incubate the plate at 37°C for 10-15 minutes (or until calibrator 0 attains dark blue colour for desired OD).
10. Pipette 50 µl of stopping solution into each well at the same timed intervals as in step 8.
11. Read the plate on a microwell plate reader at 450 nm within 20 minutes after addition of the stopping solution.

\* If the OD exceeds the upper limit of detection or if a 450 nm filter is unavailable, a 405 or 415 nm filter may be substituted. The optical densities will be lower, however, this will not affect the results of [ ] / control samples.

## CALCULATIONS

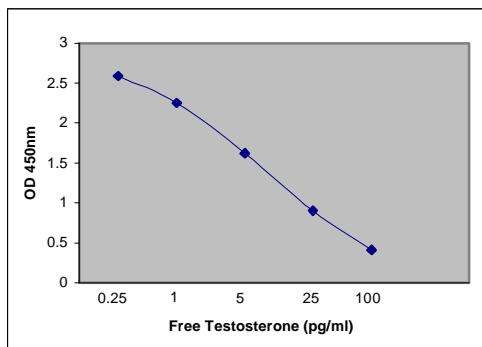
1. Calculate the mean optical density of each calibrator duplicate.
2. Draw a calibration curve on semi-log paper with the mean optical densities on the Y-axis and the calibrator concentrations on the X-axis. If immunoassay software is being used, a 4-parameter curve is recommended.
3. Calculate the mean optical density of each unknown duplicate.
4. Read the values of the unknowns directly off the calibration curve.

### TYPICAL TABULATED DATA

Calibrator	OD 1	OD 2	Mean OD	Value (pg/ml)
0	2.100	2.013	2.057	0
1	1.463	1.506	1.485	0.25
2	0.908	0.922	0.915	1.02
3	0.472	0.462	0.467	5.5
4	0.277	0.254	0.266	25
5	0.153	0.146	0.150	125
Unknown	0.464	0.458	0.461	5.7

## TYPICAL CALIBRATION CURVE

Sample curve only. **Do not** use to calculate results.



## PERFORMANCE CHARACTERISTICS

### SENSITIVITY

The lower detection limit is calculated from the calibration curve by determining the resulting concentration of the mean OD of Calibrator 0 (based on 10 replicate analyses) minus 2 SD. Therefore, the sensitivity of the  $\alpha$ -Direct Free Testosterone ELISA kit is **0.17 pg/ml**.

### SPECIFICITY (CROSS REACTIVITY)

The following compounds were tested for cross-reactivity with the  $\alpha$ -Direct Free Testosterone ELISA kit with testosterone cross-reacting at 100%.

Steroid	%Cross Reactivity
Testosterone	100
5 $\alpha$ -DHT	5.2
Androstenedione	1.4
Androstanediol	0.8
Progesterone	0.5
Androsterone	0.1

The following steroids were tested but cross-reacted at less than 0.1%: Aldosterone, Andrenosterone, Cholesterol, Corticosterone, Dehydroepiandrosterone, Dehydroepiandrosterone Sulfate, Epiandrosterone, 17 $\beta$ -Estradiol, Estriol and Pregnenolone.

### INTRA-ASSAY PRECISION

Three samples were assayed ten times each on the same calibration curve. The results (in pg/ml) are tabulated below:

Sample	Mean	SD	CV%
1	1.17	0.20	17.0
2	15.96	0.79	4.9
3	62.46	2.95	4.7

### INTER-ASSAY PRECISION

Three samples were assayed ten times over a period of two weeks. The results (in pg/ml) are tabulated below:

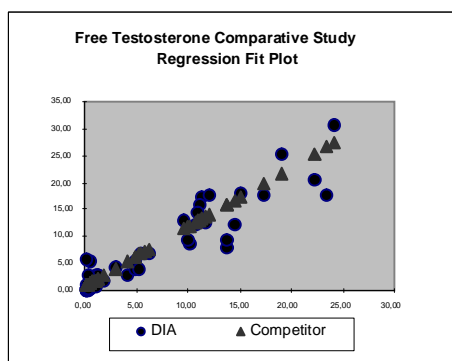
Sample	Mean	SD	CV%
1	0.97	0.12	12.4
2	25.81	1.36	5.3
3	75.81	6.66	8.8

## COMPARATIVE STUDIES

The  $\alpha$ -Direct Free Testosterone ELISA Kit (y) was compared with a competitor's Free Testosterone Coated Tube RIA Kit (x). The comparison of 61 serum samples yielded the following linear regression results:

$$y (\alpha) = 1.0137x (\text{competitor}) + 0.6404$$

$$r = 0.89$$



## EXPECTED NORMAL VALUES

As for all assays each laboratory should collect data and establish their own range of expected normal values. The results of an expected range study with apparently normal healthy subjects yielded the following results (all values are reported in pg/ml):

Group	N	Median	Central 95% Range	Absolute Range
Males	71	12.3	4.25-30.37	3.84-34.17
Females	60	1.03	0.04-4.18	0.01-7.01

## EFFECT OF SEX HORMONE BINDING GLOBULIN (SHBG)

The purpose of this study was to investigate a possible interference caused by the binding of SHBG to the free testosterone-horse radish peroxidase conjugate. A charcoal-stripped human serum pool was spiked precisely with SHBG at concentrations ranging from 6-200  $\mu$ g/ml and was assayed with the  $\alpha$ -Direct Free Testosterone ELISA Kit. Results tabulated below (in pg/ml):

SHBG Added	OD 450nm	Percent B/B <sub>0</sub> (%)
0 $\mu$ g/ml	2.34	100.0
6.25 $\mu$ g/ml	2.33	99.7
12.5 $\mu$ g/ml	2.27	97.2
50 $\mu$ g/ml	2.14	91.6
200 $\mu$ g/ml	2.10	89.7

The results showed bound values between 90-100% of B/B<sub>0</sub> (B<sub>0</sub>=unspiked serum) even at higher than normal (0.5-5  $\mu$ g/ml) SHBG levels. In conclusion, the results showed that there was no significant influence by SHBG in the  $\alpha$ -Direct Free Testosterone Direct ELISA kit.

## EFFECT OF HUMAN SERUM ALBUMIN (HSA)

The purpose of this study was to investigate a possible interference of human serum albumin (HSA) on the assay procedure. HSA was added to three samples at concentrations of 1.25, 2.5 and 5.0 g/dl. All samples were assayed with the  $\alpha$ -Direct Free Testosterone ELISA Kit and yielded the following results (in pg/ml):

Sample	Added HSA g/dl			
	0	1.25	2.5	5.0
1	0.52	0.34	0.54	0.53
2	15.8	14.2	12.5	10.9
3	26.2	23.0	21.0	18.6

The results demonstrate no significant influence of added HSA on the three serum samples.

## REFERENCES

- Winter, S.J., et al., The Analog Free Testosterone Assay: *Anal. Clin. Chem.* 44 (10):2178-2182, 1998.
- Ooi, D.S., et al., Establishing Reference Interval for DPC's Free Testosterone Radioimmunoassay. *Clin. Biochem.* 31(1):15-21, 1998.
- Marcus, G.J., et al., A Simple Linked Immunoassay for Testosterone. *Steroids* 46:975, 1985.
- Joshi, U.M., et al., A Sensitive Specific Enzyme Immunoassay for Serum Testosterone. *Steroids* 34:35, 1979.
- Swinkels, L.M. J. et al., Salivary and Plasma Free Testosterone and Androstenedione Levels in Women. *Am. Clin. Biochem.* 25:354, 1988.
- Swinkels, L.M.J., et al., A Symmetric Dialysis Method for the Determination of Free Testosterone in Human Serum. *Clin. Chem. Acta* 165:341, 1987.
- Ekins, R. Hirsutism: Free and Bound Testosterone. *Ann. Clin. Biochem* 27:91, 1990.
- Manni, A., et al., Bioavailability of Albumin-Bound Testosterone. *J. Clin. Endo. Metab.* 61:705, 1985.
- Ekins, R. The Science of Free Testosterone Measurement. *Proc. UK NEQAS Meeting.* 3:35-39, 1998.
- Longcope, C. et al., Free Estradiol, Free Testosterone and Sex Hormone Binding Globulin in Perimenopausal Women. *J. Clin. Endo. Metab.* 64:513, 1987.
- Vermeulen, A., et al., The Apparent Free Testosterone Concentration: An Index of Androgenicity. *J. Clin. Endo. Metab.* 33:759, 1971.
- Paulson, J.D., et al., Free Testosterone Concentration in Serum. Elevation is the Hallmark of Hirsutism. *Am. J. Obs. Gynecol.* 128:851, 1977.
- Cumming D.C., et al., Non Sex Hormone Binding Globulin Bound Testosterone as a Marker for Hyperandrogenism. *J. Clin. Endo. Metab.* 61:873, 1985.
- Baxendale, P.M., et al., Salivary Testosterone Relationship to Unbound Plasma Testosterone in Normal and Hyperandrogenic Women. *Clin. Endocrinol.* 16:595, 1982.
- Biffignandi, P., et al., Female Hirsutism: Pathophysiological Considerations and Therapeutic Implications. *Endocrinol Rev.* 5:488, 1984.
- Wu, C.H., Plasma Free and Protein-Bound Testosterone in Hirsutism. *Obstet. Gynecol.* 60:188, 1982.

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	<u>Used symbols</u>	<u>Symboles utilisés</u>
	Consult instructions for use	Consulter les instructions d'utilisation
	Storage temperature	Température de conservation
	Use by	Utiliser jusque
	Batch code	Numéro de lot
	Catalogue number	Référence de catalogue
	Control	Contrôle
	In vitro diagnostic medical device	Dispositif médical de diagnostic in vitro
	Manufacturer	Fabricant
	Contains sufficient for <n> tests	Contenu suffisant pour <n> tests
	Wash solution concentrated	Solution de lavage concentrée
	Zero calibrator	Calibrateur zéro
	Calibrator #	Calibrateur #
	Control #	Contrôle #
	Tracer	Traceur
	Tracer	Traceur
	Tracer concentrated	Traceur concentré
	Tracer concentrated	Traceur concentré
	Tubes	Tubes
	Incubation buffer	Tampon d'incubation
	Acetonitrile	Acétonitrile
	Serum	Sérum
	Specimen diluent	Diluant du spécimen
	Dilution buffer	Tampon de dilution
	Antiserum	Antisérum
	Immunoabsorbent	Immunoabsorbant
	Calibrator diluent	Diluant de calibrateur
	Reconstitution solution	Solution de reconstitution
	Polyethylene glycol	Glycol Polyéthylène
	Extraction solution	Solution d'extraction
	Elution solution	Solution d'elution
	Bond Elut Silica cartridges	Cartouches Bond Elut Silica
	Pre-treatment solution	Solution de pré-traitement
	Neutralization solution	Solution de neutralisation
	Tracer buffer	Tampon traceur
	Microtiterplate	Microplaque de titration
	HRP Conjugate	HRP Conjugué
	HRP Conjugate	HRP Conjugué
	HRP Conjugate concentrate	HRP Conjugué concentré
	HRP Conjugate concentrate	HRP Conjugué concentré
	Conjugate buffer	Tampon conjugué
	Chromogenic TMB concentrate	Chromogène TMB concentré
	Chromogenic TMB solution	Solution chromogène TMB
	Substrate buffer	Tampon substrat
	Stop solution	Solution d'arrêt
	Incubation serum	Sérum d'incubation
	Buffer	Tampon
	AP Conjugate	AP Conjugué
	Substrate PNPP	Tampon PNPP
	Biotin conjugate concentrate	Biotine conjugué concentré
	Avidine HRP concentrate	Avidine HRP concentré
	Assay buffer	Tampon de test
	Biotin conjugate	Biotine conjugué
	Specific Antibody	Anticorps spécifique
	Streptavidin HRP concentrate	Concentré streptavidine HRP
	Non-specific binding	Liant non spécifique
	2nd Antibody	Second anticorps
	Acidification Buffer	Tampon d'acidification