

Human Laminin (LN) ELISA Kit

Catalog No.: abx050139

Size: 96T

Range: 156 pg/ml - 10000 pg/ml

Sensitivity: < 10 pg/ml

Storage: Store at 2-8°C for 6 months, or at -20°C for 12 months.

Application: For quantitative detection of LN in Human Serum, Plasma, Tissue Homogenates and other biological fluids.

Introduction: Laminins are major proteins in the basal lamina, a protein network foundation for most cells and organs. Fifteen laminin trimers have been identified. The laminins are combinations of different alpha-, beta-, and gamma-chains. The laminins are an important and biologically active part of the basal lamina, influencing cell differentiation, migration, adhesion as well as phenotype and survival. Recently, several publications have demonstrated that laminins can be used to culture cells, such as pluripotent stem cells, that are difficult to culture on other substrates. Rodin et al. showed that recombinant laminin 511 can be used to create a totally xeno-free and defined cell culture environment to culture human pluripotent ES cells and human iPS cells.

Principle of the Assay

This kit is based on sandwich enzyme-linked immune-sorbent assay technology. Anti-LN is pre-coated onto a 96-well plate. Biotin conjugated anti-LN antibody is used as a detection antibody. The standards, test samples and biotin conjugated antibody are added to the wells and washed with wash buffer. Avidin-Biotin-Peroxidase Complex is added and unbound conjugates are washed away with wash buffer. TMB substrate is used to visualize HRP enzymatic reaction. TMB is catalyzed by HRP to produce a blue color product that changes into yellow after adding acidic stop solution. The intensity of the color yellow is proportional to the LN amount bound on the plate. The O.D. absorbance is measured spectrophotometrically at 450 nm in a microplate reader, and then the concentration of LN can be calculated.

Kit components

1. One pre-coated 96-well microplate (12 × 8 well strips)
2. Standard: 2 tubes
3. Sample / Standard diluent buffer: 30 ml
4. Biotin conjugated antibody: 130 µl - Dilution: 1:100
5. Antibody diluent buffer: 12 ml
6. Avidin-Biotin-Peroxidase Complex (ABC): 130 µl - Dilution: 1:100
7. ABC diluent buffer: 12 ml
8. TMB substrate: 10 ml
9. Stop solution: 10 ml
10. Wash buffer (25X): 30 ml

Material Required But Not Provided

1. 37°C incubator
2. Microplate reader (wavelength: 450 nm)
3. High-precision pipette and sterile pipette tips
4. Automated plate washer
5. ELISA shaker
6. 1.5 ml tubes to prepare standard/sample dilutions
7. Deionized or distilled water
8. Absorbent filter papers
9. 100 ml and 1 liter graduated cylinders

Protocol

A. Preparation of sample and reagents

1. Sample

Store samples to be assayed within 24 hours at 2-8°C. Alternatively, aliquot and store at -20°C or -80°C for long term. Avoid repeated freeze-thaw cycles.

- **Serum:** Allow the serum to clot in a serum separator tube (about 30 min) at room temperature. Centrifuge at approximately 1000 x g for 10 min. Analyze the serum immediately or aliquot and store frozen at -20°C or -80°C.
- **Plasma:** Collect plasma using EDTA as the anticoagulant. Centrifuge for 15 min at 1000 x g within 30 min of collection. Analyze immediately or aliquot and store frozen at -20°C or -80°C.
- **Tissue homogenates:** The preparation of tissue homogenates will vary depending upon tissue type – this is just an example. Rinse tissues with ice-cold PBS to remove the excess of blood. Weigh before homogenization. Finely mince tissues and homogenize with a tissue homogenizer on ice in PBS and sonicate the cell suspension. Centrifuge the homogenates at 5000 x g for 5 min and collect the supernatant. Assay immediately or aliquot and store at -20°C.
- **Other biological fluids:** Centrifuge at approximately 1000 x g for 20 min to remove precipitant. Analyze immediately or aliquot and store at -20°C or -80°C.

Note:

- » Samples must be diluted so that the expected concentration falls within the kit's range.
- » Fresh samples or recently obtained samples are recommended to prevent protein degradation and denaturalization that may lead to erroneous results.
- » Please bring sample slowly to room temperature. Sample hemolysis will influence the result. Hemolyzed specimen should not be used.
- » NaN₃ cannot be used as test sample preservative, since it inhibits HRP.
- » Always use non-pyrogenic, endotoxin-free tubes for blood collection.

General Sample guideline:

Estimate the concentration of the target in the sample and select the correct dilution factor to make the diluted target concentration fall near the middle of the kit's range. Generally, for high concentration (100000 pg/ml - 1000000 pg/ml), dilute 1:100, for medium concentration (10000 pg/ml - 100000 pg/ml), dilute 1:10 and for low concentration (156 pg/ml - 10000 pg/ml), dilute 1:2. Very low concentrations (≤ 156 pg/ml) do not need dilution. Dilute the sample with the provided Sample Diluent Buffer and mix thoroughly. Several trials may be necessary to determine the optimal dilution factor.

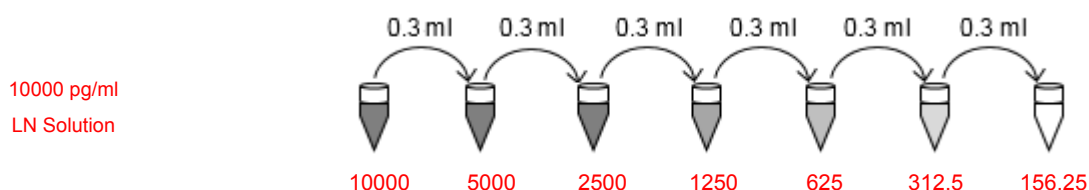
2. Wash buffer

Dilute the concentrated Wash buffer 25-fold (1/25) with distilled water (i.e. add 30 ml of concentrated wash buffer into 720 ml of distilled water).

3. Standard

The standard solution should be prepared no more than 2 hours prior to the experiment. Note: Do not dilute the standard directly in the plate. Two tubes of standard are included in each kit. Use one tube for each experiment.

- 10,000 pg/ml of standard solution: Add 1 ml of Sample/Standard diluent buffer into one Standard tube to make the 10,000 pg/ml solution. Keep the tube at room temperature for 10 min, mix thoroughly and avoid foaming or bubbles. Further dilute the above solution by a factor of 1 to give the highest standard (10000 pg/ml).
- 10000 pg/ml $\rightarrow$ 156.25 pg/ml standard solutions: Label 6 tubes with 5000 pg/ml, 2500 pg/ml, 1250 pg/ml, 625 pg/ml, 312.5 pg/ml and 156.25 pg/ml. Aliquot 0.3 ml of the Sample / Standard diluent buffer into each tube. Add 0.3 ml of the above 10000 pg/ml standard solution into the 1st tube and mix thoroughly. Transfer 0.3 ml from the 1st tube to the 2nd tube and mix thoroughly. Transfer 0.3 ml from the 2nd tube to the 3rd tube and mix thoroughly, and so on.



Note: Do not vortex the standard during reconstitution, as this will destabilize the protein. Once your standard has been reconstituted, it should be used right away. We do not recommend reusing the reconstituted standard.

4. Preparation of Biotin conjugated antibody working solution: prepare no more than 2 hour before the experiment.

- Calculate the total volume of the working solution: $0.1 \text{ ml} / \text{well} \times \text{quantity of wells}$. (Allow 0.1-0.2 ml more than the total volume).
- Dilute the Biotin conjugated antibody with Antibody diluent buffer at 1/100 and mix thoroughly. i.e. Add 1 μl of Biotin conjugated antibody into 99 μl of Antibody diluent buffer.

5. Preparation of Avidin-Biotin-Peroxidase Complex (ABC) working solution: prepare no more than 1 hour before the experiment.

- Calculate the total volume of the working solution: $0.1 \text{ ml} / \text{well} \times \text{quantity of wells}$. (Allow 0.1-0.2 ml more than the total volume).
- Dilute the ABC with ABC diluent buffer at 1/100 and mix thoroughly. i.e. Add 1 μl of ABC into 99 μl of ABC diluent buffer.

B. Assay Procedure

The ABC working solution and TMB substrate must be kept warm at 37 °C for 30 min before use. It is recommend to plot a standard curve for each test.

- Set standard, test sample and control (zero) wells on the pre-coated plate and record their positions. It is recommend to measure each standard and sample in duplicate or triplicate.
- Aliquot 0.1 ml of the diluted standards into the standard wells.
- Add 0.1 ml of Sample / Standard diluent buffer into the control (zero) well.
- Add 0.1 ml of appropriately diluted sample into test sample wells.
- Seal the plate with a cover and incubate at 37°C for 90 min.
- Remove the cover and discard the plate content, tap the plate on absorbent filter papers. **Do not let the wells completely dry at any time. Do not wash plate.**
- Add 0.1 ml of Biotin conjugated antibody working solution into the wells (standard, test sample & zero wells). Add the solution at the bottom of each well without touching the side wall.
- Seal the plate with a cover and incubate at 37°C for 60 min.
- Remove the cover and wash the plate 3 times with Wash buffer using one of the following methods:

Manual Washing: Discard the solution without touching the side walls. Fill each well completely (approximately 400 μl) with Wash buffer and incubate on an ELISA shaker for 2 min. Invert the plate each time and decant the contents; tap it 4-5 times on absorbent paper towel to completely remove the liquid. Repeat this procedure for a total of three times.

Automated Washing: Discard the solution and wash the plate three times overfilling the wells with Wash buffer. After the final wash invert the plate, decant the contents; tap it 4-5 times on absorbent paper towel to completely remove the liquid. It is recommended that the washer be set for a soaking time of 1-2 min.

10. Add 0.1 ml of ABC working solution into each well, cover the plate and incubate at 37°C for 30 min.
11. Remove the cover and wash the plate 5 times with Wash buffer, and each time let the wash buffer stay in the wells for 1-2 min. (See Step 9 for plate wash method).
12. Add 0.1 ml of TMB substrate into each well, cover the plate and incubate at 37°C in dark conditions for 15-30 min. (Note: This incubation time is for reference only, the optimal time should be determined by the end user.)
13. Add 0.1 ml of Stop solution into each well and mix thoroughly. The color changes into yellow immediately.
14. Ensure that there are no fingerprints or water on the bottom of the plate, and that the fluid in the wells is free of bubbles. Measure the absorbance at 450 nm immediately.

For calculation: (the relative O.D.450) = (the O.D.450 of each well) – (the O.D.450 of Zero well). The standard curve can be plotted as the relative O.D.450 of each standard solution (Y) vs. the respective concentration of the standard solution (X). Log-log curve fitting is recommended for data analysis. The Human LN concentration of the samples can be interpolated from the standard curve.

Note: If the samples measured were diluted, multiply the dilution factor by the concentrations from interpolation to obtain the concentration before dilution. Equilibrate the ABC working solution for at least 30 minutes to room temperature prior to use. It is recommended to plot a standard curve for each test.

C. Precautions

1. Before the experiment, centrifuge the tubes briefly to bring down any contents trapped in the lid.
2. For each step in the procedure, total dispensing time for addition of reagents to the assay plate should not exceed 10 minutes. It is recommended to assay all standards, controls and samples in duplicate or triplicate.
3. Avoid foaming or bubbles when mixing or reconstituting components. Prepare the Standard solutions within 15 min of starting the experiment. Please use the diluted Standard for a single assay procedure and discard after use.
4. Do not let the plate dry out completely during the assay as this will inactivate the biological material on the plate. Wash buffer may crystallize and separate. If this happens, please warm the tube and mix gently to dissolve.
5. Ensure plates are properly sealed or covered during incubation steps.
6. Complete removal of all solutions and buffers during wash steps is necessary for accurate measurement readings.
7. Do not reuse pipette tips and tubes to avoid cross contamination.
8. Do not use expired components or components from different batches.
9. The TMB Substrate solution is easily contaminated; work under sterile conditions when handling the TMB substrate solution. The TMB Substrate solution should also be protected from light. Unreacted substrate should be colorless or very light yellow in appearance. Aspirate the dosage needed with sterilized tips and do not dump the residual solution back into the vial.

