**Product Manual** 

# CytoSelect™ 24-Well Cell Invasion Assay (Laminin, Colorimetric Format)

**Catalog Number** 

CBA-110-LN 12 assays

FOR RESEARCH USE ONLY Not for use in diagnostic procedures



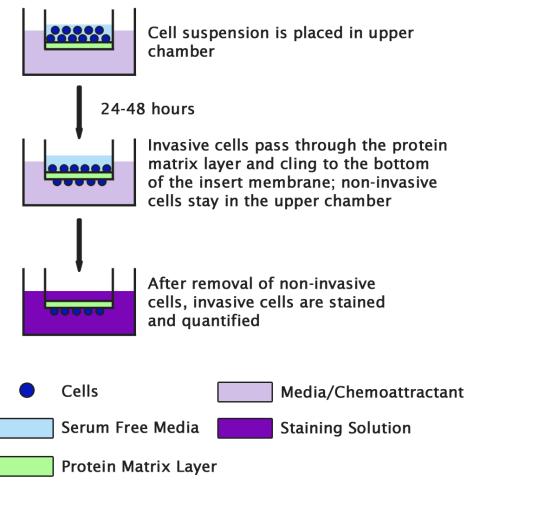
# **Introduction**

The ability of malignant tumor cells to invade normal surrounding tissue contributes in large part to the significant morbidity and mortality of cancers. Invasiveness requires several distinct cellular functions including adhesion, motility, detachment, and extracellular matrix proteolysis. Metastatic cells produce many proteolytic enzymes (e.g. lysosomal hydrolysates, collagenases, plasminogen activators) while the expression of certain cell surface protease receptors is also increased.

Cell Biolabs CytoSelect<sup>™</sup> Laminin Cell Invasion Assay Kit utilizes Murine Laminin I-coated inserts to assay the invasive properties of tumor cells. It contains sufficient reagents for the evaluation of 12 samples.

# Assay Principle

The CytoSelect<sup>TM</sup> Laminin Cell Invasion Assay Kit contains polycarbonate membrane inserts (8 µm pore size) in a 24-well plate. The upper surface of the insert membrane is coated with a uniform layer of dried Murine Laminin I matrix. This laminin matrix layer serves as a barrier to discriminate invasive cells from non-invasive cells. Invasive cells are able to degrade the laminin matrix layer, and ultimately pass through the pores of the polycarbonate membrane. Finally, the cells are removed from the top of the membrane and the invaded cells are stained and quantified.





# **Related Products**

- 1. CBA-100: CytoSelect<sup>TM</sup> 24-Well Cell Migration Assay (8µm, Colorimetric)
- 2. CBA-100-C: CytoSelect<sup>™</sup> 24-Well Cell Migration and Invasion Assay (8µm, Colorimetric)
- 3. CBA-110: CytoSelect<sup>TM</sup> 24-Well Cell Invasion Assay (Basement Membrane, Colorimetric)
- 4. CBA-110-COL: CytoSelect<sup>TM</sup> 24-Well Cell Invasion Assay (Collagen I, Colorimetric)
- 5. CBA-111: CytoSelect<sup>TM</sup> 24-Well Cell Invasion Assay (Basement Membrane, Fluorometric)
- 6. CBA-111-COL: CytoSelect<sup>™</sup> 24-Well Cell Invasion Assay (Collagen I, Fluorometric)
- 7. CBA-111-LN: CytoSelect<sup>TM</sup> 24-Well Cell Invasion Assay (Laminin I, Fluorometric)
- 8. CBA-112: CytoSelect<sup>TM</sup> 96-Well Cell Invasion Assay (Basement Membrane, Fluorometric)
- 9. CBA-112-COL: CytoSelect<sup>™</sup> 96-Well Cell Invasion Assay (Collagen I, Fluorometric)
- 10. CBA-112-LN: CytoSelect<sup>™</sup> 96-Well Cell Invasion Assay (Laminin, Fluorometric)
- 11. CBA-130: CytoSelect<sup>™</sup> 96-Well Cell Transformation Assay (Soft Agar Colony Formation)

## **Kit Components**

- 1. <u>Laminin Invasion Chamber Plate</u> (Part No. 111001-LN): One 24-well plate containing 12 laminincoated cell culture inserts
- 2. Cell Stain Solution (Part No. 11002): One 10 mL bottle
- 3. Extraction Solution (Part No. 11003): One 10 mL bottle
- 4. Cotton Swabs (Part No. 11004): 40 each
- 5. Forceps: (Part No. 11005): One each

# **Materials Not Supplied**

- 1. Invasive cell lines
- 2. Cell culture medium
- 3. Serum free medium, such as DMEM containing 0.5% BSA, 2 mM CaCl<sub>2</sub> and 2 mM MgCl<sub>2</sub>
- 4. Cell culture incubator (37°C, 5% CO<sub>2</sub> atmosphere)
- 5. Light microscope
- 6. 96-well microtiter plate
- 7. Microtiter plate reader

## **Storage**

Store all components at 4°C.



### Assay Protocol

- 1. Under sterile conditions, allow the laminin invasion chamber plate to warm up at room temperature for 10 minutes.
- 2. Rehydrate the laminin layer of the cell culture inserts by adding  $300 \,\mu\text{L}$  of warm, serum-free media to the inner compartment. Incubate at room temperature for 30 minutes.
- 3. Prepare a cell suspension containing  $0.5-1.0 \ge 10^6$  cells/ml in serum free media. Agents that inhibit or stimulate cell invasion can be added directly to the cell suspension.

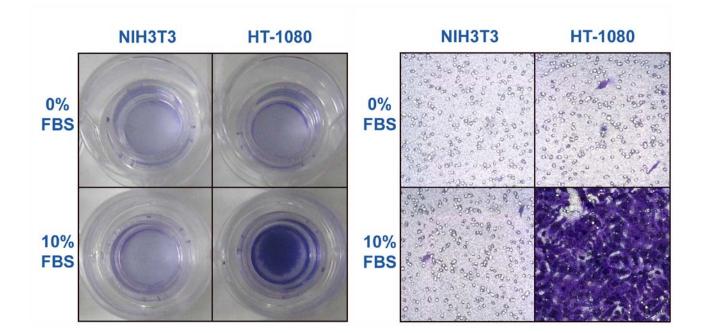
Note: Overnight starvation may be performed prior to running the assay.

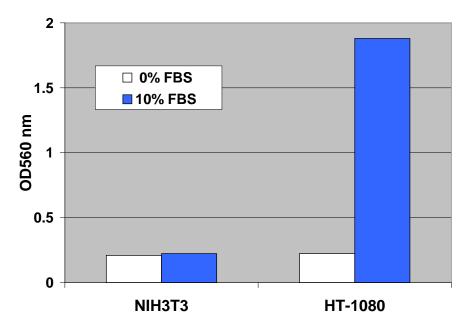
- 4. Carefully remove 250  $\mu$ L of rehydration medium (step 2) from the inserts without disturbing the laminin layer (leaving 50  $\mu$ L inside).
- 5. Add  $250 \,\mu$ L of the cell suspension solution to the inside of each insert.
- 6. Add 500  $\mu$ L of media containing 10% fetal bovine serum or desired chemoattractant(s) to the lower well of the invasion plate.
- 7. Incubate for 12-24 hours in a cell culture incubator.
- 8. Carefully aspirate the media from the inside of the insert. Wet the ends of 2-3 cotton-tipped swabs with water, flatten the ends of the swabs by pressing them against a clean hard surface, and gently swab the interior of the inserts to remove non-invasive cells. Take care not to puncture the polycarbonate membrane. Be sure to remove cells on the inside perimeter of the insert.
- 9. Transfer the insert to a clean well containing 400  $\mu$ L of Cell Stain Solution and incubate for 10 minutes at room temperature.
- 10. Gently wash the stained inserts several times in a beaker of water. Allow the inserts to air dry.
- 11. (optional) Count invasive cells with a light microscope under high magnification objective, with at least three individual fields per insert.
- 12. Transfer each insert to an empty well, adding 200  $\mu$ L of Extraction Solution per well, then incubating 10 minutes on an orbital shaker.
- 13. Transfer 100  $\mu$ L from each sample to a 96-well microtiter plate and measure the OD 560nm in a plate reader.



### **Example of Results**

The following figures demonstrate typical with the CytoSelect<sup>™</sup> Laminin Cell Invasion Assay Kit. One should use the data below for reference only. This data should not be used to interpret actual results.





**Figure 1. Human Fibrosarcoma HT-1080 Laminin Cell Invasion.** HT-1080 and NIH3T3 (negative control) were seeded at 200,000 cells/well and allowed to invade toward FBS for 24 hrs. Invasive cells on the bottom of the invasion membrane were stained (top panel picture) and quantified at OD 560nm after extraction (bottom panel figure).



## **References**

- 1. Erkell, L. J., Schirrmacher, V. (1988) Cancer Res 48, 6933-6937.
- 2. Montgomery, A. M. P., De Clerck, Y. A., Langley, K. E., Reisfeld, R. A., Mueller, B. M. (1993) *Cancer Res* **53**,693-700.
- 3. Monsky, W. L., Lin, C. Y., Aoyama, A., Kelly, T., Akiyama, S. K., Mueller, S. C., Chen, W. T. (1994) *Cancer Res* **54**,5702-5710.

### **Warranty**

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