Mouse HDL-Cholesterol Kit Instructions

For the quantitative determination of HDL-cholesterol in mouse serum or plasma

Catalog #79990
96 Assays

For research use only. Not for use in diagnostic procedures.

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TABLE OF CONTENTS

A. Intended Use ................................................................................................. 1
B. Introduction ..................................................................................................... 1
C. Principles of the Assay .................................................................................. 1
D. Kit Storage ...................................................................................................... 1
E. Assay Materials
   E.1. Materials provided .................................................................................. 1
   E.2. Materials to be supplied by user .......................................................... 1
F. Assay Precautions ......................................................................................... 2
G. Maximizing Kit Performance ...................................................................... 2
H. Sample Collection ........................................................................................ 2
I. Assay Procedure
   I.1. Preparation of reagents ......................................................................... 2
   I.2. Preparation of samples, calibrators, and controls ................................. 2
   I.3. Assay procedure .................................................................................... 3
   I.4. Determining the HDL-cholesterol concentration .................................. 3
J. Performance characteristics
   J.1. Assay range ............................................................................................ 4
   J.2. Precision ................................................................................................. 4
Warranty ............................................................................................................. 4
A. Intended Use
The Mouse HDL-Cholesterol kit is for the quantitative determination of HDL-cholesterol in mouse serum or plasma. Please read the complete kit insert before performing this assay. The kit is for RESEARCH USE ONLY. It is not intended for use in diagnostic procedures.

B. Introduction
An inverse relationship between HDL-cholesterol levels in serum and the incidence/prevalence of coronary heart disease has been demonstrated in a number of epidemiological studies. The importance of HDL as a risk factor for coronary heart disease is now recognized.

Accurate measurement of HDL is of vital importance when assessing patient’s risk for coronary heart disease.

C. Principle of the Assay
Crystal Chem’s Mouse HDL-cholesterol assay is based on a modified polyvinyl sulfonic acid (PVS) and polyethylene-glycol methyl ether (PEGME) coupled classic precipitation method with the improvements in using optimized quantities of PVS/PEGME and selected detergents. LDL, VLDL, and chylomicron (CM) react with PVS and PEGME and the reaction results in inaccessibility of LDL, VLDL and CM by cholesterol oxidase (CHOD) and cholesterol esterase (CHER), whereas HDL reacts with the enzymes. The HDL reacts with the enzymes to produce H$_2$O$_2$ which is quantified by the Trinder reaction.

D. Kit Storage
1. Upon receipt of the Mouse HDL-Cholesterol Kit, store it at 2-8°C (do not freeze the kit or hold it at temperatures above 25°C).
2. The kit should not be used after the expiration date.

E. Assay Materials
E.1. Materials provided

<table>
<thead>
<tr>
<th>Mark</th>
<th>Description</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>CC1</td>
<td>Reagent CC1 (liquid)</td>
<td>1 X 25 mL</td>
</tr>
<tr>
<td>CC2</td>
<td>Reagent CC2 (liquid)</td>
<td>1 X 8 mL</td>
</tr>
<tr>
<td>CAL1</td>
<td>Calibrator 1 (lyophilized)</td>
<td>1 X 1 mL</td>
</tr>
</tbody>
</table>

E.2. Materials required but not provided
Microplates
Micropipettes and disposable tips
Clean glass tubes and test tube racks
Volumetric flasks
Incubator (37°C)
Distilled water
Microplate reader or spectrophotometer (should read $A_{600}$ values)
0.9% Saline
F. Assay Precautions
1. Only appropriately-trained personnel should use the kit. Laboratory personnel should wear suitable protective clothing. All chemicals and reagents should be considered potentially hazardous. Avoid ingestion and contact with skin and eyes.
2. Some assay components contain human sourced materials. Accordingly, all assay components should be handled as if potentially infectious using safe laboratory procedures.
4. Do not use the reagents after the expiration date.

G. Maximizing Kit Performance
1. Given the small sample volumes required (3 µL), pipetting should be done as carefully as possible. A high quality 10 µL or better precision pipette should be used for such volumes. Drops of liquid adhering to the outside of the pipette tips should be removed by wiping to ensure the highest degree of accuracy.
2. In order to prevent the wells from drying out and to get the best results, samples and reagents should be dispensed quickly into the wells.
3. Each calibrator and sample should be assayed in duplicate.
4. The same sequence of pipetting and other operations should be maintained in all procedures.
5. Do not mix reagents that have different lot numbers.

H. Sample Collection
Use fresh mouse serum or plasma samples (EDTA, Citrate, Li Heparin). Fasting and non-fasting samples can be used.

I. Assay Procedure
I.1. Preparation of reagents
All reagents are provided ready-to-use and should be brought to room temperature for at least 30 minutes prior to use. Reagents should be stored at 2-8°C immediately after use. Before use, mix the reagents thoroughly by gentle agitation or swirling.

I.2. Preparation of samples, calibrators, and controls
1. Reconstitute the calibrators as directed on the label with 1 mL of distilled water. To ensure complete reconstitution, equilibrate vials at room temperature for 30 minutes before first use.
   Note: Reconstituted calibrators are stable for 7 days when capped tightly and stored at 2-8°C. In addition to running the calibrator provided, the assay requires running a blank calibrator. 0.9% saline should be used for running the blank calibrator. Optional controls are sold separately (Cat# 79993). Controls should be reconstituted per the directions provided on the label.
2. Bring all samples, calibrators, and controls to room temperature.
I.3. Assay procedure

The procedure below reflects a manual procedure performed using a microplate reader. The procedure can be adopted to be run in a glass tube using a spectrophotometer. The assay can also be adopted to work on various automated analyzers. Please contact Crystal Chem for more information.

1. Add 225 µL of Reagent CC1 and 3 µL of sample, calibrator, or control into each well (as needed) of a microplate and mix well by repeated pipetting.
2. Place microplate in incubator (37ºC) and allow microplate to equilibrate to 37ºC over 5 minutes.
3. Measure absorbance using a plate reader (measure \(A_{600}\) values).

**Note:** The Mouse HDL-Cholesterol assay is an end-point assay and the first reading point A1 is right before the addition of reagent CC2.

4. Pipette 75 µL of Reagent CC2 and mix well by repeated pipetting.
5. Measure the increase in absorbance after 5 minutes at 37ºC using a plate reader (measure \(A_{600}\) values).

**Figure 1. Summary of assay procedure**

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I.4. Determining the mouse HDL-cholesterol concentration

1. Calculate the change in absorbance \(\Delta A\) (5 mins ~ 0 mins)

\[
\Delta A = (OD_{600nm, 5 mins}) - (OD_{600nm, 0 mins})
\]

2. Using linear graph paper, construct the HDL-cholesterol calibration curve by plotting the mean change in absorbance value for the calibrator (incl. blank) on the Y axis versus the corresponding HDL-cholesterol concentration on the X axis.

**Note:** Calibrator value varies per lot and should be obtained from the calibrator label.

3. HDL-cholesterol concentrations in the samples are interpolated using the calibration curve and mean change in absorbance values for each sample. This interpolation can be simplified using Equation 1 below. The HDL-cholesterol concentration is expressed as mg/dL. This unit of measure can be converted in mmol/L by multiplying the obtained concentration in mg/dL by 0.02586.

**Note:** Samples with a high mouse HDL-cholesterol concentration (180.0 mg/dL or higher) should be diluted with 0.9% saline and rerun.

**Equation 1. Calculation of HDL-cholesterol concentration**

\[
\text{HDL-cholesterol concentration} = \left[ \frac{(\text{sample } \Delta A_{600} - \text{blank } \Delta A_{600})}{(\text{cal } \Delta A_{600} - \text{blank } \Delta A_{600})} \right] \times \text{cal conc.}
\]
J. Performance characteristics

J.1. Assay range
The Mouse HDL-cholesterol assay has a linear range from 1.5 – 180.0 mg/dL.

J.2. Precision
The assay has a within-run and total precision of CV < 10%.

Warranty

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