



Nori® Canine MMP-3 ELISA Kit-DataSheet

MMP-3 is a member of the matrix metalloproteinase (MMP) family are involved in the breakdown of extracellular matrix and during tissue remodeling in normal physiological processes, such as embryonic development and reproduction, as well as in disease processes, such as arthritis, and tumour metastasis. Most MMPs are secreted as inactive proproteins which are activated when cleaved by extracellular proteinases. The MMP-3 enzyme degrades collagen types II, III, IV, IX, and X, proteoglycans, fibronectin, laminin, and elastin. In addition, MMP-3 can also activate other MMPs such as MMP-1, MMP-7, and MMP-9, rendering MMP-3 crucial in connective tissue remodeling.^[1] The enzyme is thought to be involved in wound repair, progression of atherosclerosis, and tumor initiation. Expression of MMP3 is primarily regulated at the level of transcription, where the promoter of the gene responds to various stimuli, including growth factors, cytokines, tumor promoters, and oncogene products.^[2] Recently, the MMP-3 gene was shown to be down-regulated in individuals with cleft lip and palate when compared to controls,^[3] reinforcing the nature of cleft lip/palate as a condition resulting from insufficient or defective embryonic tissue remodeling.

References

1. Ye S, et al. (1996). *J. Biol. Chem.* **271** (22): 13055–60.
2. Matrisian LM (1990). *Trends Genet.* **6** (4): 121–5.
3. Bueno DF, et al. (2010). *Stem Cell Rev* **7** (2): 446–457.

PRINCIPLE OF THE ASSAY

This ELISA kit is for quantification of MMP-3 in canine samples. This is a shorter ELISA assay that reduces time to 50% compared to the conventional method, and the entire assay only takes 3 hours. This assay employs the quantitative sandwich enzyme immunoassay technique and uses biotin-streptavidin chemistry to improve the performance and the sensitivity of the assays. An antibody specific for canine MMP-3 has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any MMP-3 present is bound by the immobilized antibody. After washing away any unbound substances, a detection antibody specific for canine MMP-3 is added to the wells. Following wash to remove any unbound antibody reagent, a detection reagent is added. After intensive wash a substrate solution is added to the wells and color develops in proportion to the amount of MMP-3 bound in the initial step. The color development is stopped and the intensity of the color is measured.

This package insert must be read in its entirety before using this product.

Storage

Store the kit at 4°C. The kit can be used in 3 months.



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MATERIALS PROVIDED

Description	Quantity	Description	Quantity	Description	Quantity
Antibody Precoated Plate	1	20 x PBS	1	Substrate Solution	1
Detection Antibody	1	20 x Assay Buffer	1	Stop Solution	1
Conjugate	1	Reagent Diluent	1	DataSheet/Manual	1
Standard	3	MSDS/CoA	1	96-well plate sheet	1

Bring all reagents to room temperature before use.

Reagent Preparations

Canine MMP-3 Detection Antibody– The lyophilized Detection Antibody should be stored at 4°C or -20°C in a manual defrost freezer for up to 3 months, if not used immediately. Centrifuge for 1 min at 6000 x g to bring down the material prior to open the vial. The vial contains sufficient Detection Antibody for a 96-well plate. Add 200 µL of sterile 1 x PBS to the antibody vial, vortex 20 sec and allow it to sit for 5 min. Take 200 µL of detection antibody to 10.5 mL of Reagent Diluent to make **working dilution of detection antibody** if the entire 96-well plate is used. If the partial antibody is used store the rest at -20°C until use.

Canine MMP-3 Standard (3 vials) – The lyophilized canine MMP-3 Standard has 3 vials. Each vial contains the standard sufficient for generating a standard curve. The non-reconstituted standard can be stored at 4°C or -20°C for up to 3 months if not used immediately. Centrifuge for 1 min at 6000 x g to bring down the material prior to open the vial. Add 500 µL of 1 x Assay Buffer to a standard vial to make the high standard concentration of 1800 pg/mL. Vortex 20 sec and allow it to sit for 5 min prior to use. A seven-point standard curve is generated using 2-fold serial dilutions in the Assay Buffer, vortex 20 sec for each of dilution steps.

Conjugate (53 µL) – Centrifuge for 1 min at 6000 x g to bring down the material prior to open the vial. The vial contains 53 µL Conjugate sufficient for a 96-well plate. If the volume is less than 50 µL, add sterile 1 x PBS to reach 53 µL and vortex 10 sec. Make 1:200 dilutions in Reagent Diluent. If the entire 96-well plate is used, add 53 µL of Conjugate to 10.5 mL of Reagent Diluent to make **working dilution of Conjugate** prior to the assay. The rest of undiluted Conjugate can be stored at 4°C for up to 3 months. DO NOT FREEZE.

20 x PBS, pH 7.3, 30 mL- Dilute to 1 x PBS with deionized distilled water and mix well prior to use.

20 x Assay Buffer, 20 mL- Dilute to 1 x Assay Buffer with 1 x PBS prior to use.

Reagent Diluent, 21 mL.

Substrate Solution, 10.5 mL.

Stop Solution, 5.5 mL.



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Assay Procedure

1. Lift the plate cover from the top left corner and cover the wells that are not used. Vortex briefly the samples and the standards prior to the assay. Add 100 μ L of **sample** (such as plasma or serum) or **standard** to each well, use duplicate wells for each of the standards and samples, cover the 96-well plate and incubate 1 hour at room temperature.
2. Aspirate each well and wash with **1 x Assay Buffer**, repeating the process two times for a total of three washes. Wash by filling each well with 1 x Assay Buffer (300 μ L) using a multi-channel pipette, manifold dispenser or auto-washer. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining Assay Buffer by aspirating or by inverting the plate and blotting it against clean paper towels.
3. Add 100 μ L of the working dilution of the **Detection Antibody** to each well. Cover the plate and incubate 1 hour at room temperature.
4. Repeat the aspiration/wash as in step 2.
5. Add 100 μ L of the working dilution of **Conjugate** to each well. Cover the plate and incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.
6. Repeat the aspiration/wash as in step 2.
7. Add 100 μ L of **Substrate Solution** to each well. Incubate for 5 minutes at room temperature. Avoid placing the plate in direct light.
8. Add 50 μ L of **Stop Solution** to each well. Gently tap the plate to ensure thorough mixing.
9. Determine the optical density of each well immediately, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

Precaution and Technical Notes

1. It is critical to follow the procedure step by step otherwise appropriate color development may not occur as expected.
2. A standard curve should be generated for each set of samples assayed. Thorough mixing of the standard at each of the dilution steps is critical to ensure a normal calibration curve.
3. Plasma or serum sample should be diluted with equal volume of 1 x Assay Buffer and vortex for 1 min prior to assay. If the OD value still exceeds the upper limit of the standard curve, further dilution is recommended till it falls in the detection range and the dilution factor must be used for calculation of the concentration.
4. Conjugate contains enzyme, DO NOT mass up with Detection Antibody.
5. The Stop Solution is an acid solution, handle with caution.
6. This kit should not be used beyond the expiration date on the label.
7. A thorough and consistent wash technique is essential for proper assay performance. Assay Buffer should be dispensed forcefully and removed completely from the wells by aspiration or decanting. Remove any remaining Assay Buffer by aspiration or by inverting the plate and blotting it against clean paper towels.
8. Use a fresh reagent reservoir and pipette tips for each step.
9. It is recommended that all standards and samples be assayed in duplicate.
10. Avoid microbial contamination of reagents and buffers. This may interfere with the sensitivity of the assay.

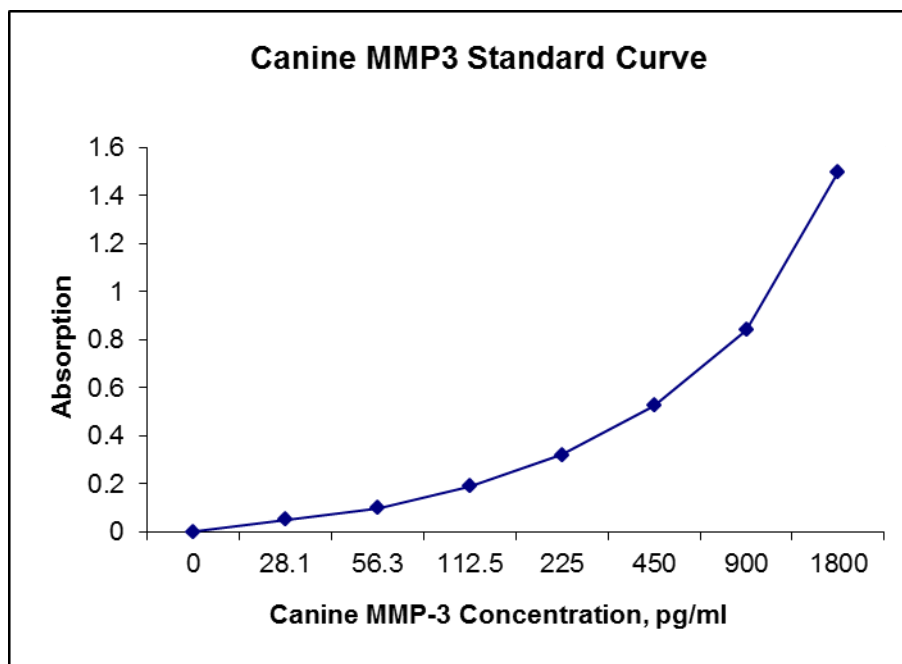
Calculation of Results

Average the duplicate readings for each standard, control, and sample and subtract the average zero (blank) standard optical density.

Create a standard curve by reducing the data using computer software capable of generating a four-parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the MMP-3 concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

The Standard Curve

The graph below represents typical data generated when using this Canine MMP-3 ELISA Kit. The standard curve was calculated using a computer generated 4-PL curve-fit. For this case, a Bio-Rad iMark™ Microplate Reader and a Microplate Manager 6 Software were used to generate this curve. The correlation coefficient (r^2) is 0.999-1.000.





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Specificity

The following recombinant canine proteins prepared at 10 ng/ml were tested and exhibited no cross-reactivity or interference.

ApoAI, BMP1, BMP2, BMP3, BMP4, BMP7, CRP, HGF, HSP27, IL-1 α , IL-1 RI, IL-1 β , IL-1RA, IL-2, IL-4, IL-5, IL-6, IL-8, IL-10, IL-12, IL-13, IL-15, IL-17C, IL-21, IL-23, IFN γ , MMP-2, sIL-2R, sIL-6R, PDGF, PLA2G7, prolactin, TGF β 1, TGF β 2, TGF β 3, TLR1, TLR2, TLR3, TNF- α , TNF RI, TNF RII, VEGF.

Calibration

This kit is calibrated against a highly-purified CHO cell-expressed recombinant canine MMP-3.

Detection Range

28-1800 pg/ml

Assay Sensitivity

6 pg/ml

Assay Precision

Intra-Assay %CV: 6; Inter-Assay %CV: 9

For Research Use Only

Related products

20 x Sample Diluent, GR103058

20 x PBS, Cat. GR103004-20

10 x ELISA Assay Buffer, Cat. GR103028

10 x ELISA Reagent Diluent, Cat. GR103055

Universal Blocking Buffer, Cat. GR103005

2 x Recombinant Protein Stabilizer, Cat. GR03014-2

5 x Recombinant Protein Stabilizer, Cat. GR103014-5

ELISA G-Blue Substrate Solution, Cat. GR103021

Canine MMP-3 Standard

Canine MMP-3 detection antibody

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Troubleshooting Guide

Problem	Possible causes	Solution
Poor standard curve	• Inaccurate pipetting • Improper standard curve	• Check pipettes • Check and use the correct dilution buffer • Vortex 30 sec for each of standard dilution steps
Low signal	• Improper preparation of standard, samples, detection antibody, and/or conjugate • Too brief incubation times • Inadequate reagent volume or improper dilution	• Briefly spin down vials before opening. Reconstitute the powder thoroughly. • Ensure sufficient incubation time. • Check pipettes and ensure correct preparation.
Large CV	• Inaccurate pipetting and mixing • Improper standard/sample dilutions. • Air bubbles in wells.	• Check pipettes and ensure thorough mixing. • Use the correct dilution buffers • Remove bubbles in wells.
High background	• Plate is insufficiently washed. • Contaminated Assay Buffer	• Review the datasheet for proper wash. If using a plate washer, ensure that all ports are unobstructed. • Make fresh Assay Buffer
No signal detected	• The procedure was misconducted.	• Ensure the step-by-step protocol was correctly followed and no misstep was conducted.
Low sensitivity	• Improper storage of the ELISA kit • Stop solution	• Store standards and detection antibody at -20°C after reconstitution, others at 4°C. Keep substrate protected from light. • Adding stop solution to each well before reading plate