

## Instructions for Use

### Super G Plus Preservation Buffer

#### Use:

Super G Plus Preservation Buffer is a non-protein based reagent designed to both block non-specific protein binding on porous nitrocellulose substrates and preserve protein immuno-reactivity over prolonged storage of printed microarrays. It is supplied as a 1X solution, ready to use out of the bottle, for microarray applications with fluorescence endpoint detection. Super G Plus may be used as a blocking agent only, or as a blocking agent and preservative.

### Storage:

Super G Plus Preservation Buffer should be stored at 4°C once received.

### **Blocking and Preservation Procedure:**

After spotting content on the slides, allow spots to completely dry before proceeding with the use of Super G Plus. It is important to cover the slide surface coating completely with Super G Plus. For best results, submerge the slide completely. Incubation time should be optimized for your specific application.

- I. For using Super G Plus as a blocking agent only, and performing assays immediately after blocking step:
  - 1. Soak slides in Super G Plus for 1 hour, wash with 1X PBS (or buffer similar in composition to your assay buffer) for 5 minutes with agitation prior to proceeding with the assay.
  - 2. Proceed with assay as per protocol.
- II. For using Super G Plus as a blocking agent and a preservative and storing printed microarrays prior to assay:
  - 1. Soak slides in Super G Plus for 1 hour, then agitate on an orbital shaker (15 minutes).
  - 2. Transfer slides to fresh Super G Plus and agitate further on an orbital shaker (15 min.).
  - 3. Remove microarray slides from preservation buffer and place individual slides in 50ml conical screw-cap tubes (1 slide per tube or 2 slides back-to-back).
  - 4. Spin slides dry in centrifuge by spinning at 200 rpm for 1 minute.

Caution: DO NOT WASH arrays after incubation with Super G Plus.

- 5. Remove slides from centrifuge tubes and allow to air dry.
- 6. Place slides in an airtight slide box with desiccant.
- 7. For long-term storage, place desiccated slide at 4 to -20°C.

# GRACE BIO-LABS

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### Notes:

- 1. Centrifugation of slides allows for even drying of Super G Plus over the slide surface while removing excess reagent from the surface. Slides will not be completely dry after centrifugation. After air drying, the nitrocellulose slide surface will appear slightly darker and the surface will be sticky.
- 2. If slides are allowed to air dry without centrifugation to remove surface solution, drying may be uneven and the nitrocellulose surface may appear irregularly patterned with some translucent spots. This irregular appearance does not alter the preservation properties of Super G Plus. The slide surface will appear evenly white again once the preservative is removed during downstream processing.
- 3. For optimal results, slides should not be pre-treated with other reagents before using Super G Plus.

### **Post-Storage Rehydration Procedure:**

Remove slides from refrigerator/freezer and allow slides to come to room temperature while desiccated to avoid condensation on the slide surface. Rehydrate slides by washing with 1X PBS (or with a buffer similar in composition to your assay buffer) for 30 minutes with agitation. Proceed with the assay.

Caution: DO NOT allow slides to dry out until assay is completed and you are ready to scan).