

# Mouse C-Telopeptide Of Type I Collagen CTX-1 ELISA Kit

**Catalog #: BG-MUS10728 (96 wells)**

## User Manual

(please be notified that the manual is updated, and different from the online data, to improve the assay)

*This kit is designed to quantitatively detect the levels of Mouse CTX-1 in cell lysates, serum/ plasma and other suitable sample solution.*

## **Manufactured and Distributed by:**

**Novatein Biosciences**  
310 W Cummings Park, Woburn, MA, 01801, USA  
Phone: (617) 238-1396, Fax: (617) 380-0053  
Email: [Info@novateinbio.com](mailto:Info@novateinbio.com)

## Important notes

Before using this product, please read this manual carefully; after reading the subsequent contents of this manual, please note the following specially:

- The operation should be carried out in strict accordance with the provided instructions.
- Store the unused strips in a sealed foil bag at 2-8°C.
- Always avoid foaming when mixing or reconstituting protein solutions.
- Pipette reagents and samples into the center of each well, avoid bubbles.
- The samples should be transferred into the assay wells within 15 minutes of dilution.
- We recommend that all standards, testing samples are tested in duplicate.
- Using serial diluted sample is recommended for first test to get the best dilution factor.
- If the blue color develops too light after 15 minutes incubation with the substrate, it may be appropriate to extend the incubation time (Do not over-develop).
- Avoid cross-contamination by changing tips, using separate reservoirs for each reagent.
- Avoid using the suction head without extensive wash.
- Do not mix the reagents from different batches.
- Stop Solution should be added in the same order of the Substrate Solution.
- TMB developing agent is light-sensitive. Avoid prolonged exposure to the light.

**ALWAYS REFER TO LOT SPECIFIC PROTOCOL PROVIDED WITH EACH KIT FOR INSTRUCTIONS.**

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## Scientific Background

In bone physiology, the C-terminal telopeptide (or more formally, carboxy-terminal collagen crosslinks, and known by the acronym CTX) is a telopeptide that can be used as a biomarker in the serum to measure the rate of bone turnover. It can be useful in assisting clinicians to determine a patient's nonsurgical treatment response as well as evaluate a patient's risk of developing complications during healing following surgical intervention. The test used to detect the CTX marker is called the Serum CrossLaps, and it is more specific to bone resorption than any other test currently available. The CTX test measures for the presence and concentration of a crosslink peptide sequence of type I collagen, found, among other tissues, in bone. This specific peptide sequence relates to bone turnover because it is the portion that is cleaved by osteoclasts during bone resorption, and its serum levels are therefore proportional to osteoclastic activity at the time the blood sample is drawn

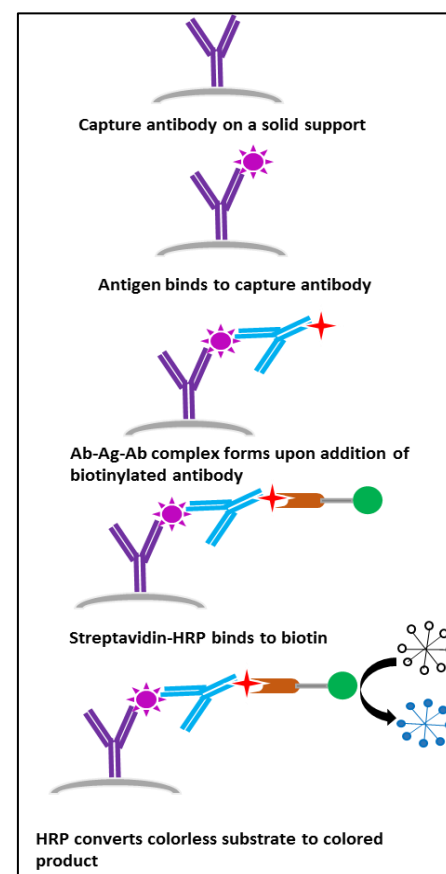
## Intended use

The kit is used to quantify the Mouse CTX-1 in serum/ plasma, cell culture supernatant and other suitable sample solution.

<b>Standard range</b>	62.5 – 4000 pg/mL
<b>Assay time</b>	4.5 hours
<b>Validity</b>	Six months
<b>Store at</b>	2-8 °C

## Assay principle

The Mouse CTX-1 ELISA Kit is based on standard sandwich enzyme-linked immunosorbent assay technology. Anti-Mouse CTX-1 specific antibody has been pre-coated onto 96-well plate. Mouse CTX-1 present in the standards/ samples bind to the capture antibody. Subsequently, biotinylated anti-Mouse CTX-1 detection antibody is added to form an Ab-Ag-Ab sandwich. After a washing step, streptavidin-HRP is added and the unbound conjugate is removed with wash buffer. Next, addition of HRP substrate, TMB, results in the production of a blue colored product that changes to yellow after the addition of acidic Stop Solution. The density of yellow color is directly proportional to the amount of Mouse CTX-1 captured on plate.



## Materials supplied

1. Mouse CTX-1 standard:	4 ng/mL × 2
2. 96-well plate pre-coated with anti-Mouse CTX-1 Ab:	1
3. Sample Diluent buffer :	12 ml × 2
4. Detection antibody:	1 vial, dilution 1:60
5. Streptavidin-HRP:	1 vial dilution 1:40
6. Antibody Diluent buffer:	12 ml
7. Streptavidin HRP diluent buffer:	12 ml
8. TMB substrate:	12 ml × 2
9. Stop Solution:	12 ml
10. 20 × Wash Buffer:	25 ml
11. Plate sealers	2
12. Package insert	1

## Materials required but not supplied

- 1x PBS.
- Standard plate reader capable of measuring absorbance at 450 nm.
- Adjustable pipettes and disposable pipette tips.
- Multi-channel pipettes, manifold dispenser or automated microplate washer.
- Distilled water.
- Absorbent paper.
- Materials used for sample preparation.

## Sample Preparation and storage

Store samples to be assayed within 24 hours at 2-8°C. For long-term storage, aliquot and freeze samples at -20°C. Avoid repeated freeze-thaw cycles.

- Cell culture supernatant, tissue lysate or body fluids: Remove particulates by centrifugation, analyze immediately or aliquot and store at -20°C
- Serum: Allow the serum to clot in a serum separator tube (about 4hours) at room temperature. Centrifuge at approximately 1000 X g for 15 min. Analyze the serum immediately or aliquot and store frozen at -20°C. Dilution factor: 1:20 to 1:30.
- Plasma: Collect plasma using heparin as an anticoagulant. Centrifuge for 15 min at 1000 x g within 30 minutes of collection. Analyze immediately or aliquot and store frozen at -20°C. EDTA and citrate are not recommended as the anticoagulant. Diluted like that of serum.

## Reagent Preparation

### Standard

- Mouse CTX-1: Standard solution should be prepared no more than 2 hours prior to the experiment. Two tubes of standard (4 ng /ml) are included in each kit. Use one tube for each experiment.
- Label 6 Eppendorf tubes with 2000 pg/ml, 1000 pg/ml, 500 pg/ml, 250 pg/ml, 125 pg/ml, 62.5 pg/ml, respectively. Then make 2-fold serial dilution from 4000 pg/ml to 62.5 pg/ml in 1.5 ml tubes with sample diluent buffer.
- Make sure each tube has  $\geq 250 \mu\text{l}$  of standard.

**Note:** *The standard solutions are best used within 2 hours.*

### Preparation of detection anti-Mouse CTX-1 antibody working solution

- The stock solution is stable at 2-8 °C for up to 1 month. After opening the vial use within 1 month. For long-term storage, please aliquot and store at -20 °C. Avoid freeze-thaw cycles.
- The working solution should be prepared no more than 2 hours prior to the experiment

- The reagent is supplied as 60X concentrate. Empty the total contents into 10.62 ml of Antibody Diluent Buffer or prepare the solution separately in a volume as needed. The solution should be mixed thoroughly.
- The total volume should be: 0.1 ml/well x the number of wells (Allowing 0.1-0.2 ml more than total volume).

#### **Preparation of Streptavidin-HRP working solution**

- The solution should be prepared no more than 1 hour prior to the experiment.
- The total volume should be: 0.1 ml/well x the number of wells (allowing 0.1-0.2 ml more than total volume).
- Streptavidin-HRP should be diluted 1:40 with Streptavidin-HRP Diluent buffer and mixed thoroughly.

#### **Wash Buffer**

- If crystals have formed in the 20X wash buffer, warm to room temperature and mix gently until the crystals have completely dissolved.
- Dilute 25 ml Wash Buffer Concentrate (20X) to a total volume of 500 ml with distilled water.

#### **Assay Procedure**

Bring all reagents to room temperature before use. Mouse CTX-1 standard curve should be prepared for each experiment. The user will decide sample dilution factor by rough estimation of Mouse CTX-1 concentration in samples.

1. Add 100 µl of sample or standards per well. Add 0.1 ml of the sample diluent into the control well (Zero well). Cover with an adhesive strip and incubate at room temperature for 2 hours with shaking (500 rpm).  
Note: We recommend that each Mouse CTX-1 standard solution and each sample is measured in duplicate.
2. Aspirate each well and wash with Wash Buffer, repeating the process two times for a total of three washes. Wash by filling each well with Wash Buffer (300 µl) using a squirt bottle, manifold dispenser, or auto-washer. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining Wash Buffer by aspirating or by inverting the plate and blotting it against clean paper towels.
3. Add 100 µl of the Detection Antibody working solution to each well. Cover with a new adhesive strip and incubate at room temperature for 2 hours with shaking (500 rpm).
4. Repeat the aspiration/wash as in step 2.
5. Add 100 µl of the working solution of Streptavidin-HRP to each well. Cover the plate and incubate at room temperature for 30 min. Avoid placing the plate in direct light.
6. Repeat the aspiration/wash as in step 2 for three times.
7. Add 200 µl of TMB substrate solution to each well. Cover and incubate at room temperature for 20 -40 min (500 rpm shaking) until a gradient develops and you see visible color in the 2<sup>nd</sup> lowest concentration well. Protect from light. Do not over-develop.
8. Add 100 µl Stop Solution to each well. Mix well.

9. Read the Optical Density (O.D.) at 450 nm using a microtiter plate reader immediately.

**Note: if there's no plate shaker available, do all the incubations at 37 °C.**

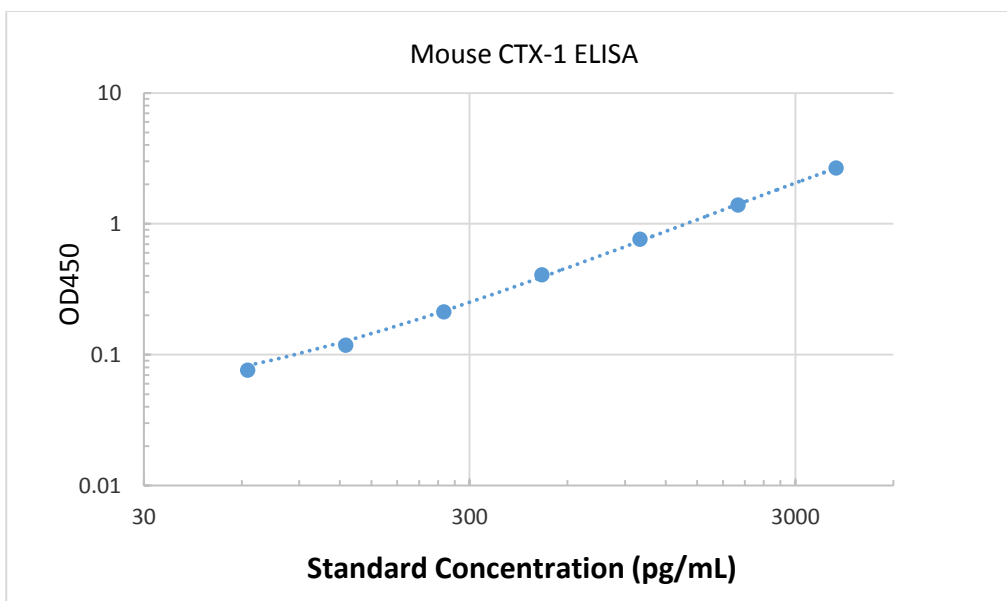
## Result calculation

For calculation, (the relative O.D.450) = (the O.D.450 of each well) – (the O.D.450 of Zero well). The standard curve can be plotted as the relative O.D.450 of each standard solution (Y) vs. the respective concentration of the standard solution (X). The Mouse CTX-1 concentration of the samples can be interpolated from the standard curve.

**Note:** if the samples measured were diluted, multiply the dilution factor to the concentrations from interpolation to obtain the concentration before dilution

## Typical data

This standard curve was generated at the Novatein Biosciences laboratory for demonstration purpose only. A standard curve must be run with each assay.



## Sensitivity

The sensitivity or minimum detectable dose (MDD) of Mouse CTX-1 was determined to be 30 pg/ml. MDD is defined as the Mouse CTX-1 concentration resulting in an O.D.<sub>450</sub> value that is 2 standard deviations higher than blank.

## Spiking and Recovery

Recovery was determined by spiking the following matrices with various concentrations of Mouse CTX-1.

Sample Type	Average Recovery (%)	Range (%)
Cell lysates	94.1	86-98
Serum	95.7	87-103



## Reproducibility

- Inter-assay- <8.8%
- Intra-assay- <6.5%

## Specificity

This kit recognizes both natural and recombinant Mouse CTX-1.

## Sample Dilution

Dilution of 1:2 – 1:50 is recommended. The user may need to determine the dilution factor in a preliminary experiment. If required, samples should be diluted in sample diluent buffer.

*For trouble shooting information please visit the following website:*

*<http://www.novateinbio.com/en/content/15-tech-info> OR*

*Email us at [techsupport@novateinbio.com](mailto:techsupport@novateinbio.com)*

Plate Layout

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# Troubleshooting Information

## High Background

Probable Cause:	Solution/ Action
<b>High incubation temperature:</b>	Incubate at room temperature (25 °C) throughout the procedure
<b>Insufficient washing of the plate:</b>	Fill the wells with wash buffer and aspirate completely for the next wash Increase the number of washes Add soak time (20-30 seconds) in between the washes Use automated plate washer, if available and check that all the channels are operating properly
<b>Concentrated streptavidin-HRP</b>	Streptavidin-HRP was not diluted properly Dilute the streptavidin-HRP as mentioned in the manual
<b>Light exposure during substrate incubation</b>	The TMB substrate is light sensitive and turns to blue color in the presence of light. The incubation must be carried out in dark.
<b>Stop solution not added</b>	Color will continue to develop if stop solution is not added
<b>Diluents came with the kit were not used</b>	Standards/ sample, detection antibody and streptavidin-HRP must be diluted in the respective buffers came with the kit. Do not use buffers from other kits
<b>Contaminated solutions</b>	Prepare fresh working solutions

## Poor Standard Curve

Probable Cause:	Solution/ Action
<b>Improper standard reconstitution:</b>	Spin the vial briefly before opening Reconstitute the standard as mentioned in the manual. After reconstitution, leave it atleast for 10 minutes at room temperature Do not store and reuse diluted standards
<b>Curve fitting problem:</b>	Log transform the values on both axes Use 4-PL/ 5-PL curve fitting programs
<b>Incubation temperature/ time</b>	Use the recommended standard incubation conditions
<b>Poor dilutions</b>	Pipetting error. Check pipetting technique and calculations. Use calibrated pipettes.

## No Signal

Probable Cause:	Solution/ Action
Omission of reagent(s):	Read the manual entirely. Check that all the reagents are added in the correct order as stated in the manual
Incorrect detection antibody was used:	Use the detection antibody came with the kit
Chromogen solutions were mixed improperly	Use the recommended procedure to prepare the TMB substrate
HRP inhibitor in sample/ buffers	Check that the samples/ buffers do not have sodium azide as it will inhibit peroxidase reaction.
Vigorous washing	If the washing is done manually, pipette the wash buffer gently.
Dried wells	Do not allow the wells to dry out during the assay. Seal with the supplied adhesive cover during incubations
Improper plate reader settings	Check the wavelength and read the plate again

## Erratic duplicate OD values

Probable Cause:	Solution/ Action
Insufficient washing of the plate	Fill the wells with wash buffer and aspirate completely for the next wash Increase number of washes Add soak time (20-30 seconds) in between the washes Use automated plate washer, is available and check that all the channels are functioning properly
Poor dilutions	Pipetting error. Check pipetting technique and calculations. Use calibrated pipettes.
Improper mixing of samples/ buffers	Mix the samples well before pipetting Thoroughly mix the working solutions of detection antibody/ streptavidin-HRP
Contamination from other wells	Do not reuse the adhesive covers from previous assay setups Change pipette tips during reagent addition. If same pipette tip is being used to dispense reagents, care should be taken, not to touch the solution in the well
Precipitates in the samples/ buffer	If precipitates are visible in wash buffer concentrate, keep it at 37 °C for 10-15 minutes until no precipitates are visible Centrifuge the samples to remove particulate matter
Dried wells	Do not allow the wells to dry out during the assay. Seal with the supplied adhesive cover during incubations

