

RayBio[®] Human IL-6R *Speed*ELISA Kit

Catalog #: ELHS-IL6sR

User Manual
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Caution:
Extraordinarily useful information enclosed



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RayBiotech, Inc.

RayBio[®] Human IL-6R **SpeedELISA**
Protocol

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Please read the entire manual carefully before starting your experiment

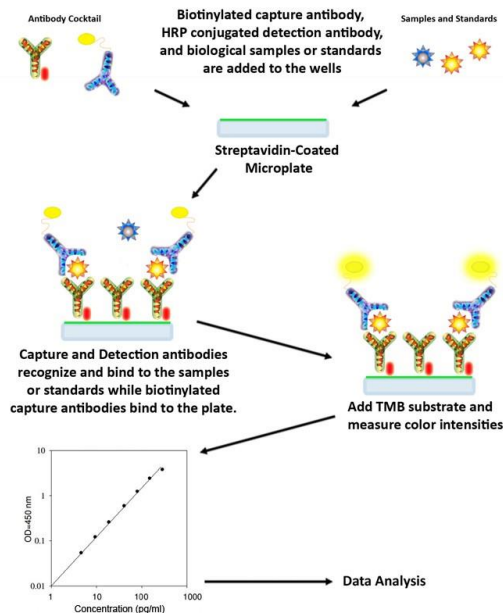
I. INTRODUCTION

The RayBio[®] Human IL-6R **SpeedELISA** kit is an in vitro enzyme-linked immunosorbent assay for the quantitative measurement of Human IL-6R in biological sample such as serum, plasma, and cell culture supernatants*. This **SpeedELISA** employs a biotinylated capture and a HRP-conjugated detection antibody which immunocaptures the sample analyte in solution. The microplate in the kit is pre-coated with streptavidin. The biotinylated capture antibody/protein/HRP-conjugated detection antibody mixture is pipetted into the wells and the biotinylated Human IL-6R present in a sample is bound to the wells by the immobilized streptavidin. After incubation, the wells are washed to remove unbound material.

A TMB substrate solution is added to the wells and color develops in proportion to the amount of Human IL-6R bound. The Stop Solution changes the color from blue to yellow, and the intensity of the color is measured at 450 nm.

***Note:** Biotin interferes with this assay. If your samples contain biotin (commonly found in certain cell culture media such as RPMI 1640, or F-12K), we recommend using the standard RayBio Sandwich ELISA kit.

II. How It Works



III. STORAGE

The entire kit may be stored at -20°C for up to 1 year from the date of shipment. Avoid repeated freeze-thaw cycles. The kit may be stored at 4°C for up to 6 months. For extended storage, it is recommended to store at -80°C. For prepared reagent storage, see table below.

IV. REAGENTS

| Component | Size / Description | Storage / Stability After Preparation |
|---|--|---------------------------------------|
| Streptavidin-Coated Microplate (Item A) | 96 wells (12 strips x 8 wells) coated with streptavidin. | 1 month at 4°C |
| Wash Buffer Concentrate (20X) (Item B) | 25 ml of 20X concentrated solution. | 1 month at 4°C |
| Standard Protein (Item C) | 2 vials of Human IL-6R. 1 vial is enough to run each standard in duplicate. | 1 week at -80°C |
| Biotinylated-Conjugated Capture Antibody (Item F) | 2 vials of Biotinylated anti-Human IL-6R. 1 vial is enough to assay half microplate. | 5 days at 4°C |
| HRP Conjugated Detection Antibody (Item G) | 2 vials of HRP conjugated anti-Human IL-6R. 1 vial is enough to assay half the microplate. | 5 days at 4°C |
| TMB One-Step Substrate Reagent (Item H) | 12 ml of 3,3',5,5'-tetramethylbenzidine (TMB) in buffer solution. | N/A |
| Stop Solution (Item I) | 8 ml of 0.2 M sulfuric acid. | N/A |
| Assay Diluent (Item E2) | 15 ml of 5X concentrated buffer | 1 month at 4°C |

V. ADDITIONAL MATERIALS REQUIRED

1. Microplate reader capable of measuring absorbance at 450 nm.
2. Precision pipettes to deliver 2 µl to 1 ml volumes.
3. Adjustable 1-25 ml pipettes for reagent preparation.
4. 100 ml and 1 liter graduated cylinders.
5. Absorbent paper.
6. Distilled or deionized water.
7. Log-log graph paper or computer and software for ELISA data analysis.
8. Tubes to prepare standard or sample dilutions.

VI. REAGENT PREPARATION

1. Bring all reagents and samples to room temperature (18 - 25°C) before use.
2. Assay Diluents should be diluted 5-fold with deionized or distilled water before use.
3. Briefly spin down the biotinylated Capture Antibody vial (Item F) before use. Add 1.9 ml of 1X Assay Diluent into the vial to prepare the capture antibody solution. Pipette up and down to mix gently. This capture antibody solution will be used in step 5 of Part VI: Reagent Preparation.
4. Briefly spin down the HRP-conjugated Detection Antibody vial (Item G) before use. Add 1.9 ml of 1X Assay Diluent into the vial to prepare the detection antibody solution. Pipette up and down to mix gently. This detection antibody solution will be used in the following step.
5. Prepare the Antibody Cocktail: a 1:1 mixture of the biotinylated capture antibody and the HRP-conjugated detection antibody. This antibody cocktail will be used in step 3 of Part VII: Assay Procedure.
For example: Add 1 ml of prepared biotinylated capture antibody solution and 1 ml HRP-conjugated detection antibody solution into a tube (transfer 1.9 ml biotinylated Capture Antibody vial, Item F into 1.9 ml HRP-conjugated Detection Antibody vial, Item G. This yields Antibody Cocktail, enough to assay half the plate). Mix well and use immediately.
6. Sample Dilution: 1X Assay Diluent (Item E2) should be used for dilution of serum, plasma, and cell culture supernatant samples. The suggested dilution for normal serum/plasma is 40 fold.

Note: Levels of IL-6R may vary between different samples. Optimal dilution factors for each sample must be determined by the investigator.

7. Preparation of standard: Briefly spin a vial of Item C. Add 400 µl of 1X Assay Diluent (Item E2; Assay Diluent should be diluted 5-fold with deionized or distilled water before use) into Item C vial to prepare a 25 ng/ml standard. Dissolve the powder thoroughly by a gentle mix. Add 80 µl of IL-6R standard solution (25 ng/ml) from the vial of Item C, into a tube with 420 µl 1X Assay Diluent to prepare a 4000 pg/ml standard solution. Pipette 300 µl 1X Assay Diluent into each tube. Use the 4000 pg/ml standard solution to produce a dilution series (shown below). Mix each tube thoroughly before the next transfer. 1X Assay Diluent serves as the zero standard (0 pg/ml).

**80 μ l Standard +
420 μ l**

200 μ l

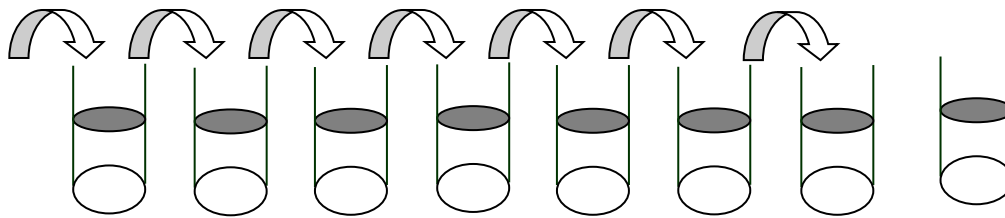
200 μ l

200 μ l

200 μ l

200 μ l

200 μ l



**4000
pg/ml**

**1600
pg/ml**

**640
pg/ml**

**256
pg/ml**

**102.4
pg/ml**

**40.96
pg/ml**

**16.38
pg/ml**

**0
pg/ml**

8. If the Wash Concentrate (20X) (Item B) contains visible crystals, warm to room temperature and mix gently until dissolved. Dilute 20 ml of Wash Buffer Concentrate into deionized or distilled water to yield 400 ml of 1X Wash Buffer.

VII. ASSAY PROCEDURE

1. Bring all reagents and samples to room temperature (18 - 25°C) before use. It is recommended that all standards and samples be run at least in duplicate.
2. Label removable 8-well strips as appropriate for your experiment.
3. Add 70 µl of the Antibody Cocktail (see Reagent Preparation step 5) into appropriate wells. Cover wells and incubate for 10 minutes at room temperature.
4. Add 50 µl of standard (see Reagent Preparation step 6) or each sample (see Reagent Preparation step 7) into each well with the Antibody Cocktail added in Step 3. Cover wells and incubate for 2 hours at room temperature.
5. Discard the solution and wash 4 times with 1x Wash Solution. Wash by filling each well with Wash Buffer (300 µl) using a multi-channel Pipette or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
6. Add 100 µl of TMB One-Step Substrate Reagent (Item H) to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking.
7. Add 50 µl of Stop Solution (Item I) to each well. Read at 450 nm immediately.

VIII. ASSAY PROCEDURE SUMMARY

1. Prepare all reagents, samples and standards as instructed.
2. Add 70 µl of the Antibody Cocktail to the appropriate wells and incubate for 10 minutes at room temperature.
3. Add 50 µl of sample or standard to each well and incubate for 2 hours at room temperature.
4. Wash wells.
5. Add 100 µl of TMB One-Step Substrate Reagent to each well. Incubate for 30 minutes at room temperature.

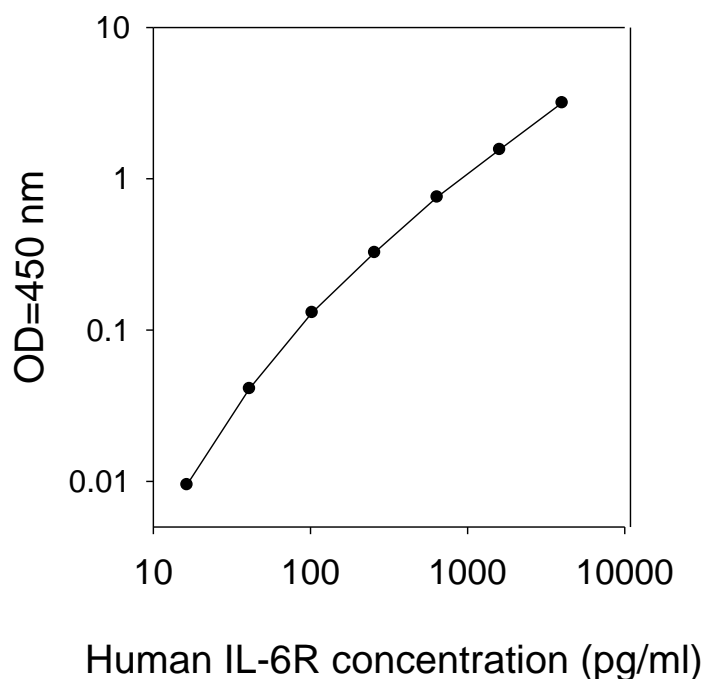
6. Add 50 μ l of Stop Solution to each well. Read at 450 nm immediately.

IX. CALCULATION OF RESULTS

Calculate the mean absorbance for each set of duplicate standards, controls and samples, and subtract the average zero standard optical density. Plot the standard curve on log-log graph paper or using Sigma plot software, with standard concentration on the x-axis and absorbance on the y-axis. Draw the best-fit straight line through the standard points.

A. TYPICAL DATA

These standard curves are for demonstration only. A standard curve must be run with each assay.



B. SENSITIVITY

The minimum detectable dose of Human IL-6R was determined to be 17 pg/ml.

Minimum detectable dose is defined as the analyte concentration resulting in an absorbance that is 2 standard deviation higher than that of the blank (diluent buffer).

C. RECOVERY

Recovery was determined by spiking various levels Human IL-6R into human serum, plasma and cell culture media. Mean recoveries are as follows:

| Sample Type | Average % Recovery | Range (%) |
|--------------------|--------------------|-----------|
| Serum | 88.83 | 84-101 |
| Plasma | 84.46 | 80-89 |
| Cell culture media | 78.44 | 76-82 |

D. LINEARITY

| Sample Type | | Serum | Plasma | Cell Culture Media |
|-------------|-----------------------|-------|--------|--------------------|
| 1:2 | Average % of Expected | 90.18 | 93.74 | 103.7 |
| | Range (%) | 86-95 | 91-98 | 99-108 |
| 1:4 | Average % of Expected | 85.11 | 104.1 | 126.6 |
| | Range (%) | 81-89 | 99-111 | 117-136 |

E. REPRODUCIBILITY

Intra-Assay CV%: <10%

Inter-Assay CV%: <12%

X. SPECIFICITY

This ELISA kit shows no cross-reactivity with the following cytokines tested: human Angiogenin, BDNF, BLC, CNTF, ENA- 78, FGF-4, IL-1 alpha, IL-1 beta, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-11, IL-12 p70, IL-12 p40, IL-13, IL-15, I-309, IP-10, FGF-4, FGF-6, FGF- 7, G-CSF, GDNF, GM-CSF, IFN-gamma, IGFBP-2, IGFBP-3, IGFBP-4, Leptin (OB), MCP-1, MCP-2, MCP-3, MDC, MIF, MIG, MIP-1 alpha, MIP-1 beta, MIP-1 delta, PARC, PDGF, RANTES, SCF, SDF-1 alpha, TARC, TGF-beta, TIMP-1, TIMP-2, TNF-alpha, TNFbeta, TPO, VEGF.

XI. TROUBLESHOOTING GUIDE

| Problem | Cause | Solution |
|---------------------|--|--|
| Poor standard curve | <ul style="list-style-type: none"> • Inaccurate pipetting • Improper standard dilution | <ul style="list-style-type: none"> • Check pipettes • Briefly centrifuge Item C and dissolve the powder thoroughly by gently mixing |
| Low signal | <ul style="list-style-type: none"> • Improper preparation of standard and/or biotinylated antibody (Item G) • Too brief incubation times Inadequate reagent volumes or improper dilution | <ul style="list-style-type: none"> • Briefly spin down vials before opening. Dissolve the powder thoroughly. • Ensure sufficient incubation time; assay procedure step 2 may be done overnight Check pipettes and ensure correct preparation |
| Large CV | <ul style="list-style-type: none"> • Inaccurate pipetting • Air bubbles in wells | <ul style="list-style-type: none"> • Check pipettes • Remove bubbles in wells |
| High background | <ul style="list-style-type: none"> • Plate is insufficiently washed Contaminated wash buffer | <ul style="list-style-type: none"> • Review the manual for proper wash. If using a plate washer, ensure that all ports are unobstructed. • Make fresh wash buffer |
| Low sensitivity | <ul style="list-style-type: none"> • Improper storage of the ELISA kit • Stop solution | <ul style="list-style-type: none"> • Store your standard at <-70°C after reconstitution, others at 4°C. Keep substrate solution protected from light. • Add stop solution to each well before reading plate |

RayBio[®] ELISA Kits

Over 2,500 ELISA kits available, visit www.RayBiotech.com/ELISA-Kits.html for details.

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