

Phospho-p38 alpha MAPK (Thr180/Tyr182) and Pan p38 MAPK ELISA Kit

Catalog #: PEL- P38-T180-T

User Manual

Last Revised: March 5, 2025

Introduction

RayBio® Phospho-p38 alpha MAPK (Thr180/Tyr182) and Pan p38 alpha MAPK ELISA kit is an in vitro enzyme-linked immunosorbent assay for the measurement of Human Mouse, and Rat phospho-p38 alpha MAPK (Thr180/Tyr182) and pan p38 alpha MAPK. An anti-p38 alpha MAPK (Thr180/Tyr182) (half plate, red marker on left side) and anti-pan p38 alpha MAPK antibody (half plate, black marker on right side) has been coated onto a 96-well plate. Controls and Samples are pipetted into the wells and phosphorylated (left side) and pan (right side) p38 alpha MAPK present in a sample is bound to the wells by the immobilized antibody. The wells are washed, and anti-p38 alpha MAPK antibody is used to detect phosphorylated or pan p38 alpha MAPK. After washing away unbound antibody, HRP-conjugated anti-rabbit IgG is pipetted into the wells. The wells are again washed, a TMB substrate solution is added to the wells and color develops in proportion to the amount of p38 alpha MAPK (Thr180/Tyr182) or pan p38 alpha MAPK bound. The Stop Solution changes the color from blue to yellow, and the intensity of the color is measured at 450 nm.

Storage / Stability

The entire kit may be stored at -20°C for up to 6 months from the date of shipment. Avoid repeated freeze-thaw cycles. For prepared reagent storage, see kit contents on the next page.

Kit Components

Name	Catalog #	Size / Qty	Description	Storage / Stability After Preparation
Anti-Phospho-P38 Microplate	PEL-P38-T-A	96 wells	Microplate coated with anti-phospho-P38 MAPK (Thr180/Tyr182) antibody (half plate, red marker on left side) and anti-p38 alpha MAPK antibody (half plate, black marker on right side).	1 month at -20°C*
Positive Control	HELAA003-1	1 vial	Lyophilized powder from treated Hela cell lysate.	1 week at -80°C
P38 alpha MAPK Detection Antibody	PEL-P38-T180-C	2 vials	Rabbit anti-p38 alpha MAPK detection antibody. 1 vial is enough to assay half the microplate.	5 days at 4°C
HRP-Conjugated anti-rabbit IgG	EL-ItemD1	25 µl	500x concentrated HRP-conjugated anti-rabbit IgG.	Do not store and reuse.
Wash Buffer	EL-ITEMB	25 ml	20X concentrated wash buffer	1 month at 4°C
Assay Diluent B	EL-ITEME	15 ml	5X concentrated assay diluent	1 month at 4°C
Lysis Buffer	EL-Lysis	5 ml	2X cell lysate buffer	1 month at 4°C
TMB One-Step Substrate Reagent	EL-TMB	12 ml	3,3',5,5'-tetramethylbenzidine (TMB) in buffer solution	N/A
Stop Solution	EL-STOP	8 ml	0.2 M sulfuric acid	N/A

*Return unused wells to the pouch containing desiccant pack, reseal along entire edge.

Additional Materials Required

- Microplate reader capable of measuring absorbance at 450 nm
- Protease and Phosphatase inhibitors.
- Precision pipettes to deliver 2 µl to 1 ml volumes
- Adjustable 1-25 ml pipettes for reagent preparation
- 100 ml and 1 liter graduated cylinders
- Log-log graph paper or computer and software for ELISA data analysis.

- Absorbent paper
- Tubes to prepare standard or sample dilutions
- Distilled or deionized water
- Shaker

Sample Preparation

For the initial experiment, we recommend a serial dilution, such as a 5-fold to 50-fold dilution, for your cell lysates with prepared Assay Diluent (see Reagent Preparation step 2) before use.

Note: The fold dilution of sample used depends on the abundance of phosphorylated proteins and should be determined empirically. More of the sample can be used if signals are too weak. If signals are too strong, the sample can be diluted further.

Reagent Preparation

1. Bring all reagents and samples to room temperature (18 - 25°C) before use.
2. Assay Diluent B should be diluted 5-fold with deionized or distilled water before use.
3. Lysis Buffer should be diluted 2-fold with deionized or distilled water (for cell lysate and tissue lysate). We also recommend the addition of protease and phosphatase inhibitors (not included) to the lysis buffer prior to use.
4. Preparation of Positive Control: Briefly spin the Standard Vial. Add 400µl of prepared 1X Assay Diluent to positive control vial. Gently mix the powder to allow it to dissolve thoroughly. If a precipitate is seen in the solution after mixing, this can be removed by a quick centrifuge of the positive control vial, and then pipetting the supernatant only for the assay.
5. If the Wash Buffer (20X) concentrate contains visible crystals, warm to room temperature and mix gently until dissolved. Dilute 20 ml of Wash Buffer concentrate into deionized or distilled water to yield 400 ml of 1X Wash Buffer.
6. Preparation of P38 MAPK Detection Antibody: Briefly spin the vial of rabbit anti -P38-MAPK. Add 100 µl of 1X Assay Diluent into the vial to prepare a phospho detection antibody concentrate. Pipette up and down to mix gently (the concentrate can be stored at 4°C for 5 days or at -80°C for one month). The concentrate should then be diluted 55-fold with 1X Assay Diluent and used in step 5 of the Assay Procedure.
7. Preparation of HRP-conjugated anti-rabbit IgG: Briefly spin the vial of HRP-conjugated anti-rabbit IgG concentrate before use. HRP-conjugated anti-rabbit IgG should be diluted 500-fold with 1X Assay Diluent and used in step 7 of the Assay Procedure.

Assay Procedure

1. Bring all reagents and samples to room temperature (18 - 25°C) before use. It is recommended to run all positive controls and samples in at least duplicate.
2. Label removable 8-well strips as appropriate for your experiment.
3. Add 100 µl of each sample or positive control (see Reagent Preparation step 4) into appropriate wells (see the following 96 well microplate format). Cover the wells and incubate for 2.5 hours at room temperature or overnight at 4°C with gentle shaking.

Anti-p38 alpha MAPK Anti-pan p38 MAPK
(Thr180/Tyr182)

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												

4. Discard the solution and wash 4 times with 1X Wash Solution. Wash by filling each well with Wash Buffer (300 µl) using a multi-channel pipette or auto-washer. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
5. Add 100 µl of prepared 1X P38 MAPK (see Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking.
6. Discard the solution. Repeat the wash as in step 4.
7. Add 100 µl of prepared HRP-conjugated anti-rabbit IgG solution (see Reagent Preparation step 7) to each well. Incubate for 1 hour at room temperature with gentle shaking.
8. Discard the solution. Repeat the wash as in step 4.
9. Add 100 µl of TMB One-Step Substrate Reagent to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking.
10. Add 50 µl of Stop Solution to each well. Read at 450 nm immediately.

Assay Procedure Summary

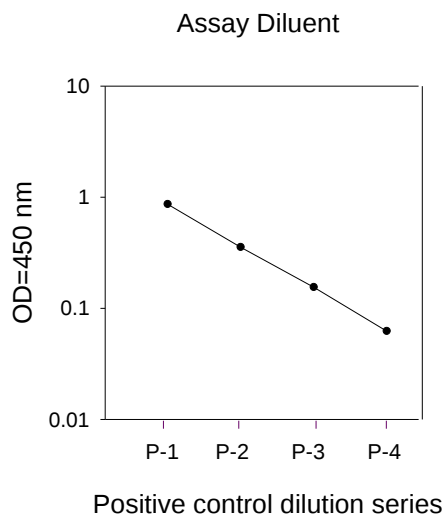
1. Prepare all reagents, samples and standards as instructed.
2. Add 100 μ l of each positive control and sample to each well. Incubate 2.5 hours at room temperature or overnight at 4°C with gentle shaking.
3. Add 100 μ l prepared detection antibody to each well. Incubate for 1 hour at room temperature with gentle shaking.
4. Add 100 μ l prepared HRP-conjugated anti-rabbit solution. Incubate for 1 hour at room temperature with gentle shaking.
5. Add 100 μ l TMB One-Step Substrate Reagent to each well. Incubate 30 minutes at room temperature.
6. Add 50 μ l Stop Solution to each well. Read at 450 nm immediately.

Typical Data

Calculate the mean absorbance for each set of duplicate positive controls and samples and subtract the average zero standard optical density.

A. Positive Control

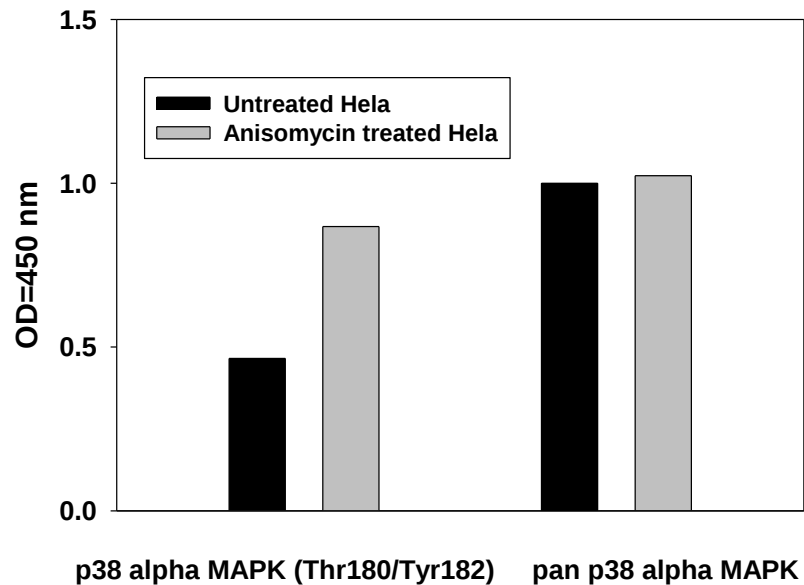
Hela cells were treated with Anisomycin at 37°C for 10 min. Solubilize cells at 4×10^7 cells/ml in Cell Lysate Buffer. Serial dilutions of lysates were analyzed in this ELISA.



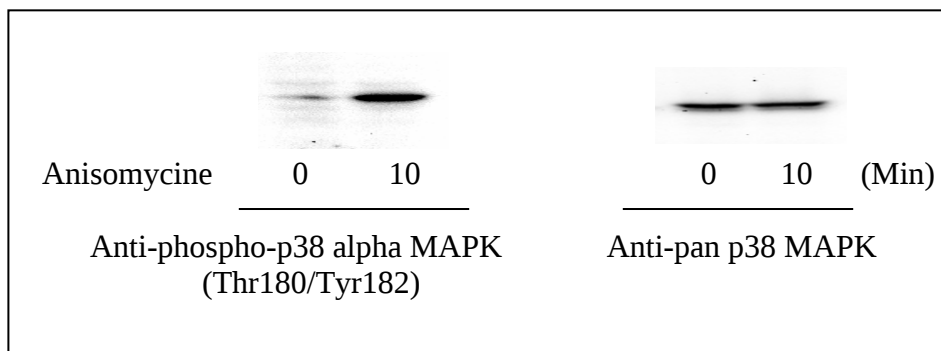
B. Anisomycin Stimulation of Hela Cell Lines

Hela cells were treated or untreated with Anisomycin for 10 min at 37°C. Cell lysates were analyzed using this phosphor ELISA and Western Blot.

i. ELISA



ii. Western-Blot Analysis



Troubleshooting Guide

Problem	Cause	Solution
Low signal in samples	- Sample concentration is too low - Improper preparation of detection antibody - Too brief incubation times - Inadequate reagent volumes or improper dilution	- Increase sample concentration Briefly spin down vials before opening. Dissolve the powder thoroughly. - Ensure sufficient incubation time; assay procedure step 3 may be done overnight Check pipettes and ensure correct preparation
High signal in samples	Sample concentration is too high	Reduce sample concentration
Large CV	Inaccurate pipetting Air bubbles in wells	- Check pipettes - Remove bubbles in wells
High background	- Plate is insufficiently washed - Contaminated wash buffer	- Review the manual for proper wash. If using a plate washer, ensure that all ports are unobstructed. - Make fresh wash buffer
Low sensitivity	- Improper storage of the ELISA kit - Stop solution - Improper primary or secondary antibody dilution	- Store your standard at <- 70°C after reconstitution, others at 4°C. Keep substrate solution protected from light. - Add stop solution to each well before reading plate - Ensure correct dilution