

Human TL1A/TNFSF15 ELISA Kit

Catalog Number: CEA-C090

Assay Tests: 96 tests

For Research Use Only. Not For Use in Diagnostic or Therapeutic Procedures

CEA-C090-EN01

IMPORTANT: Please carefully read this user guide before performing your experiment.

Product information

This kit is specifically designed for the accurate quantitation of human TL1A/TNFSF15 from cell culture supernates,

serum and plasma.

The principle of this assay employs a quantitative sandwich enzyme immunoassay approach. Initially, a microplate

is coated with a capture antibody. Then, samples and biotinylated capture antibody are added to the wells. After

the removal of any unbound materials through washing, streptavidin-HRP (SA-HRP) conjugate is added to the

wells. Streptavidin has a very high affinity for biotin, so it binds to the biotinylated capture antibody that is already

bound to the target antigen. After washing, a substrate specific to HRP is added to the wells. HRP catalyzes a

reaction that converts the substrate into a detectable signal, often a color change or luminescence, depending

on the substrate used. This enzymatic reaction amplifies the signal, allowing for higher sensitivity in detecting the

target analyte. The intensity of the signal is measured using a spectrophotometer.

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NOTE:

1. This kit is for research use only and is not for use in diagnostic or therapeutic applications.

2. Please do not use the kit after the expiration date indicated on the kit label.

3. Do not mix or substitute reagents with those from other lots or sources.

Manufactured and distributed by

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Contents

The kit contains sufficient reagents for 96 wells.

Catalog	Contents	Amount
CEA090-C01	Pre-coated Anti-TL1A Antibody Microplate	1 plate
CEA090-C02	Human TL1A Standard	80 ng×2
CEA090-C03	Biotin-Anti-TL1A Antibody Con. Solution	100 μL
CEA090-C04	Biotin-Antibody Dilution Buffer	8 mL
CEA090-C05	Streptavidin-HRP Con. Solution	500 μL
CEA090-C06	Streptavidin-HRP Dilution Buffer	15 mL
CEA090-C07	20× Washing Buffer	50 mL
CEA090-C08	Sample Dilution Buffer	15 mL×2
CEA090-C09	Substrate Solution	12 mL
CEA090-C10	Stop Solution	6 mL

NOTE: Bubbles in microplate wells do not affect the experiment and require no action. Proceed with the experimental procedures and methods described below.

Storage

Keep the unopened kit stored at 2-8 °C. Avoid using the kit beyond its expiration date. For opened kit and reconstituted reagents, with the exception of the two contents listed in following table, others can be stored for up to 30 days at 2-8 °C.

Contents	Storage conditions
Pre-coated Anti-TL1A Antibody Microplate	Return unused wells to the foil pouch, reseal along entire edge. May be stored for up to 1 month at 2-8°C.
Human TL1A Standard	Aliquot and store for up to 1 month at -70°C in a freezer. Avoid repeated freeze-thaw cycles.

NOTE: Streptavidin-HRP Con. Solution and Substrate Solution should avoid light.

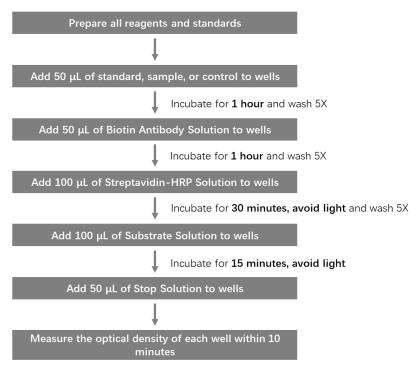
IMPORTANT: Bring all reagents to room temperature before use. If crystals have formed in buffer solution, place the buffer solution in an 37°C incubator until the crystals have completely dissolved and bring the solution back to room temperature before use.

Required materials not supplied.

Instrument	Microplate reader capable of measuring absorbance at 450 nm
Reagents	Deionized, ultrapure or distilled water
50 mL and 500 mL graduated cylinders Consumables Pipettes and pipette tips	

Workflow

Analyte: TL1A



NOTE: Incubation temperature is 18 $^{\circ}$ C-25 $^{\circ}$ C

Prepare the working buffers and standard dilutions.

Prepare the working buffers.

- 1. 1×Washing Buffer: Dilute 50 mL 20×Washing Buffer with deionized or distilled water to 1000 mL.
- Biotin-Anti-TL1A Antibody Solution: Add 60 μL of Biotin-Anti-TL1A Antibody Con. Solution to 6 mL Biotin-Antibody Dilution Buffer, thoroughly mix. The solution was freshly prepared just before use.
- 3. Streptavidin-HRP Solution: Add 360 µL of Streptavidin-HRP Con. Solution to 12 mL of Streptavidin-HRP Dilution Buffer, thoroughly mix. The solution was freshly prepared just before use.

Prepare the reconstituted standard.

Add 1 mL ultrapure water to the provided lyophilized product (CEA090-C02), dissolve at room temperature for 15-30 minutes, and mix by gently pipetting. The concentration of reconstituted human TL1A Standard is 80 ng/mL.

NOTE: Avoiding vigorous shaking or vortexing. The reconstituted solution should be stored at -70°C. The freeze-thaw cycle should not exceed 1 time.

Prepare the standard serial dilutions.

- 1. Label a tube "Cm". Add 100 μ L of the reconstituted human TL1A Standard and 900 μ L of Sample Dilution Buffer to tube Cm, gently mix well.
- 2. Label 7 tubes, one for each standard point: Std.-1, Std.-2, Std.-3, Std.-4, Std.-5, Std.-6, Std.-7.
- 3. Add 100 μ L of the liquid from **Cm** and 900 μ L of Sample Dilution Buffer to tube Std.-1, thoroughly mix (Std.-1 =800 pg/mL).
- 4. Prepare 1:1 serial dilutions for the standard curve as follows: Add 300 μL of Sample Dilution Buffer to each tube (Std.-2, Std.-3, Std.-4, Std.-5, Std.-6, Std.-7).
- 5. Transfer 300 μ L of liquid from Std.-1 to the tube Std.-2, and thoroughly mix (Std.-2 = 400 pg/mL).
- 6. Continue to transfer 300 μ L of liquid from previous dilution tube to the next dilution tube until add liquid to tube Std.-7.
- 7. Sample Dilution Buffer serves as zero standard (blank).

PROCEDURE OF ASSAY

- 1. Add 50 μ L of TL1A Standard, sample, or control to wells. Seal the plate with microplate sealing film. Incubate at room temperature (18-25 °C) for **1 hour**.
- 2. Aspirate each well and add 300 μ L of 1×Washing Buffer to each well, gently tap the plate for **1 minute**. Remove any remaining Washing Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels. Repeat the wash process four times for a total of five washes.
- 3. Add 50 μ L Biotin-Anti-TL1A Antibody Solution to each well, Seal the plate with microplate sealing film. Incubate at room temperature (18-25 °C) for **1 hour.**
- 4. Repeat step 2.
- 5. Add 100 μL of Streptavidin-HRP Solution to each well. Seal the plate with microplate sealing film. Incubate at room temperature (18-25 °C) for 30 minutes, avoid light.
- 6. Repeat step 2.
- 7. Add 100 μ L of Substrate Solution to each well. Seal the plate with microplate sealing film and incubate at room temperature (18-25 °C) for **15 minutes, avoid light**.
- Add 50 μL of Stop Solution to each well. Tap the plate gently to ensure thorough mixing.
 Note: the color in the wells should change from blue to yellow.
- 9. Read the absorbance at 450nm and 630nm using Microplate reader within 10minutes.

 *Note: To reduce the background noise, subtract the readings at 630nm from the readings at 450nm.

CALCULATION OF RESULTS

- 1. Compute the average of the duplicated readings for every standard, control, and sample. Then, subtract the average optical density (O.D.) of the zero standard(blank).
- 2. Establish a standard curve by processing the data using computer software capable of executing a **four-parameter logistic (4-PL)** curve fitting.
- 3. Normal range of Standard curve: $R^2 \ge 0.9900$.
- 4. If the OD value of the sample to be tested is higher than the highest standard, the sample shall be diluted with dilution buffer and assay repeated.

Typical data

Note: For each experiment, a standard curve needs to be set for each microplate, and the specific OD value may vary depending on different laboratories, testers, or equipment. The following example data is for reference only. The sample concentration was calculated based on the results of the standard curve.

Human TL1A Standard (pg/mL)	OD _{450nm-630nm}	R ² =0.9998
800	2.507	37
400	1.515	sity
200	0.850	Density
100	0.469	
50	0.254	Optical 1
25	0.144	0
12.5	0.091	0 200 400 600 800 100 Conc.[pg/mL]
Blank	0.040	Gone.[pg/me]

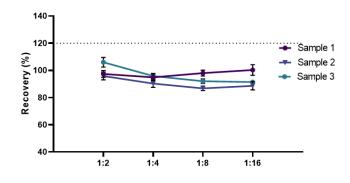
PERFORMANCE CHARACTERISTICS

1. Sensitivity

The minimum detectable concentration (MDC) of TL1A is typically less than 7.5 pg/mL. The MDC was determined by adding two standard deviations to the mean optical density value of twenty four zero standard replicates and calculating the corresponding concentration.

2. Linearity

Three samples (Serum) spiked with high concentrations of TL1A were serially diluted with dilution buffer to produce samples with values within the dynamic range of the assay and then assayed. The average recovery of TL1A for serum samples is 94.80%.



3. Intra-Assay Precision

Ten replicates of each of 3 samples containing different TL1A concentrations were tested in one assay. Acceptable criteria: CV < 10%.

Sample Concentration (pg/mL)	Mean (pg/mL)	SD	Numbers	CV (%)
600	549.488	17.586	8	3.2
100	94.020	3.575	8	3.8
37.5	34.513	1.281	8	3.7

4. Inter-Assay Precision

Three samples containing different concentrations of TL1A were tested in independent assays. Acceptable criteria: CV<15%.

Sample Concentration (pg/mL)	Mean (pg/mL))	SD	Numbers	CV (%)
600	579.412	34.893	24	6.0
100	94.692	3.165	24	3.3
37.5	36.554	3.204	24	8.8

5. Recovery

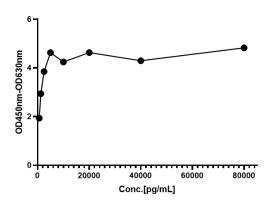
Recombinant TL1A was spiked into 3 human serum samples, and then analyzed. The average recovery of TL1A for serum samples is 96.47%.

Sample ID	Conc Measured (pg/mL)	Conc Added (pg/mL)	Conc Recovered (pg/mL)	Recovery (%)
	626.00	600	596.84	99.47
1	127.74	100	98.58	98.58
1	68.83	37.5	39.68	105.80
	32.40			
2	586.53	600	553.38	92.23
	128.73	100	95.58	95.58

	67.91	37.5	34.76	92.70
	36.83			
	602.30	600	577.82	96.30
3	117.98	100	93.50	93.50
3	59.76	37.5	35.28	94.07
	27.20			

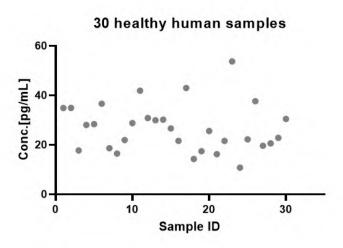
6. Hook Effect

Not be affected by the concentration of TL1A up to 5000 pg/mL.



7. Sample Values

30 healthy serum samples were evaluated for the concentrations of human TL1A in assay. The detection rate is 100%.



TROUBLESHOOTING GUIDE

Problem	Cause	Solution	
Poor standard curve	* Inaccurate pipetting	* Check pipettes	
Large CV	* Inaccurate pipetting* Air bubbles in wells	* Check pipettes * Remove bubbles in wells	
High background	* Plate is insufficiently washed* Contaminated wash buffer	* Review the manual for proper wash. * Make fresh wash buffer	
Very low readings across the plate	* Incorrect wavelengths * Insufficient development time	* Check filters/reader * Increase development time	
Samples are reading too high, but standard curve looks fine	* Samples contain cytokine levels above assay range	* Dilute samples and run again	
Drift	* Interrupted assay set-up * Reagents not at room temperature	* Assay set-up should be continuous - have all standards and samples prepared appropriately before commencement of the assay * Ensure that all reagents are at room temperature before pipetting into the wells unless otherwise instructed in the antibody inserts	