

## ActiveMax® Human VCAM-1 / CD106 µBeads, premium grade (for cells)

Cat. No. MBS-C019

### ● Product Information

| Product   | Size                      | Amount                  |
|---|---------------------------|-------------------------|
| ActiveMax® Human VCAM-1 / CD106 µBeads, premium grade (for cells) | 2.5 mg                    | $2.5 \times 10^7$ beads |
|   | 10 mg (2.5 mg $\times$ 4) | $1.0 \times 10^8$ beads |

### ● Product Description

ActiveMax® Human VCAM-1 / CD106 µBeads, premium grade (for cells) are uniform, superparamagnetic beads of 5.5 µm in diameter immobilized with Human VCAM-1 / CD106 Protein, expressed from human 293 cells (HEK293) and contains AA Phe 25 - Glu 698(Accession # P19320-1).

ActiveMax® Human VCAM-1/CD106 µBeads, premium grade (for cells) are produced under sterile manufacturing conditions (ISO 5), and no animal- or human-derived components are used throughout the production process. It is produced under our rigorous quality control system that includes a comprehensive set of tests including sterility and endotoxin tests.

### ● Product Applications

ActiveMax® Human VCAM-1 / CD106 µBeads, premium grade (for cells) is designed to synergistically activate Notch signaling in hematopoietic stem/progenitor cells ((HSPCs). The synergistic interactions between Notch ligand Delta-like 4 and VCAM-1 are leveraged to enhance Notch signaling to support efficient T-cell lineage commitment, so the beads is recommended to use for in vitro production of induced pluripotent stem cell derived T cells coordinating with ActiveMax® Human DLL4 µBeads, premium grade (Cat. No. MBS-C013).

*The Product performance has been carefully validated and tested for compatibility for cell culture use or any other applications in the early preclinical stage. For use in clinical phases, we also offer a custom GMP protein service that tailors to your needs. We will work with you to customize and develop a GMP-grade product in accordance with your requests that also meets the requirements for raw and ancillary materials use in cell manufacturing of cell-based therapies.*

### ● Formulation

Lyophilized in PBS with 0.1% HSA, pH 7.4. Trehalose is added as protectant before lyophilization.

### ● Reconstitution

Please see Certificate of Analysis for specific instructions.

*For best performance, we strongly recommend you to follow the reconstitution protocol provided in the Certificate of Analysis.*

### ● Storage

This product is stable in storage under the following conditions:

- -20°C for 12 months in lyophilized state.
- -70°C for 3 months under sterile conditions after reconstitution.

Please avoid repeated freeze-thaw cycles after reconstitution. Immediate use after reconstitution is highly recommended.

### ● Important Note

This product is for research use only and not intended for therapeutic or in vivo diagnostic use.

## ● General guidelines

It is recommended to reconstitute the lyophilized ActiveMax® Human VCAM-1 / CD106  $\mu$ Beads, premium grade (for cells) with sterile deionized water to a stock solution of 5 mg/mL ( $5 \times 10^7$  beads/mL) under ISO 5 clean conditions. Separate into working aliquots and store at  $-70^\circ\text{C}$  immediately. Upon reconstitution, immediate use is recommended for best performance.

Use a magnetic separator that is suitable for your equipment and application. Allow the beads to separate for at least 1 minute before removing supernatant. The  $\mu$ Beads are dense and will settle very quickly. Be sure that any  $\mu$ Beads mixture is homogenous before use or aliquoting.

## ● Preparing $\mu$ Beads for use

Washing the ActiveMax® Human VCAM-1 / CD106  $\mu$ Beads, premium grade (for cells) to remove trehalose from the formulation buffer before use.

1. Resuspend the Magnetic Beads in the vial (i.e. vortex for  $>30$  sec, or tilt and rotate for 5 min).
2. Transfer the desired volume of Magnetic Beads to a sterile tube.
3. Add an equal volume of sterile PBS buffer, or at least 1 mL, and mix (vortex for 5 sec, or keep on a roller for at least 2 min).
4. Place the tube on a magnet for 1 min and let the beads settle before discarding the supernatant.
5. Remove the tube from the magnet and resuspend the washed  $\mu$ Beads in the same volume of desired cell culture medium as the initial volume of added  $\mu$ Beads in **step 2**.

## ● Inducing T cell lineage differentiation

1. Seed CD34+ CD45+ hematopoietic stem cells with suitable cell density on culture plates.
2. Add the prepared ActiveMax® Human VCAM-1 / CD106  $\mu$ Beads and ActiveMax® Human DLL4  $\mu$ Beads (Cat. No. MBS-C013) with optimizing quantity and ratio for co-culture with the cells.
3. After 7-day culture intervals, the cells are counted and subjected to a full media change, including re-addition of the two  $\mu$ beads to modulate Notch-1 signaling.
4. Harvest the cells for flow cytometric analysis of the expression of T-cell progenitor markers, CD7 and CD5 on day 14.

*For use in vitro, ActiveMax® Human VCAM-1 / CD106  $\mu$ Beads, premium grade (for cells) need to be optimized by the user according to their own experiments.*

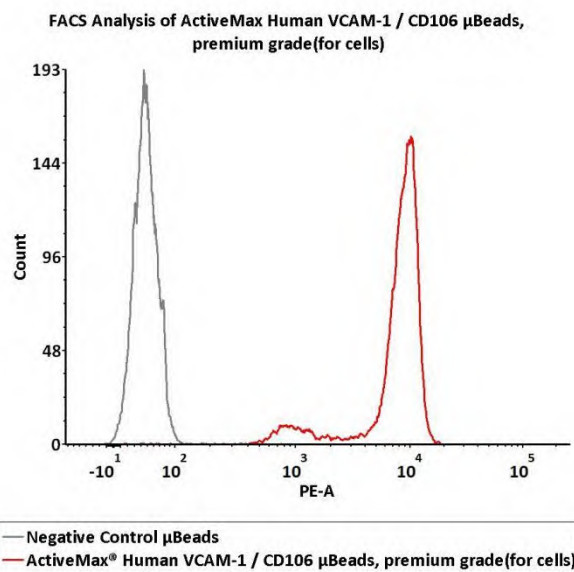
## ● Removing $\mu$ Beads from cells

1. Collect the mixture of the cells and  $\mu$ Beads after co-culturing, before centrifuging and discarding the supernatant.
2. Resuspend the pellet in suitable volume of the relevant medium, and then transfer the mixture of cells and  $\mu$ Beads into a new tube.
3. Place the tube next to a magnet for 1–2 minutes until the  $\mu$ Beads have moved to the side of the tube.
4. Transfer the supernatant containing the cells to a new tube for use.

## ● Contact Information

If you have any questions, please contact our technical support team at: [TechSupport@acrobiosystems.com](mailto:TechSupport@acrobiosystems.com)

- **Conjugated human VCAM-1 / CD106 analyzed by FACS**



**Assay of human VCAM-1 / CD106 protein on the  $\mu$ Beads surface by Flow cytometry.** The human VCAM-1 / CD106 conjugated on the  $\mu$ Beads (Cat. No. MBS-C019) surface were fluorescently stained using PE labeled anti-human VCAM-1 / CD106 antibody and analyzed by flow cytometry (QC tested).