

## Human NKp46 (Luc) Jurkat Reporter Cell Data Sheet

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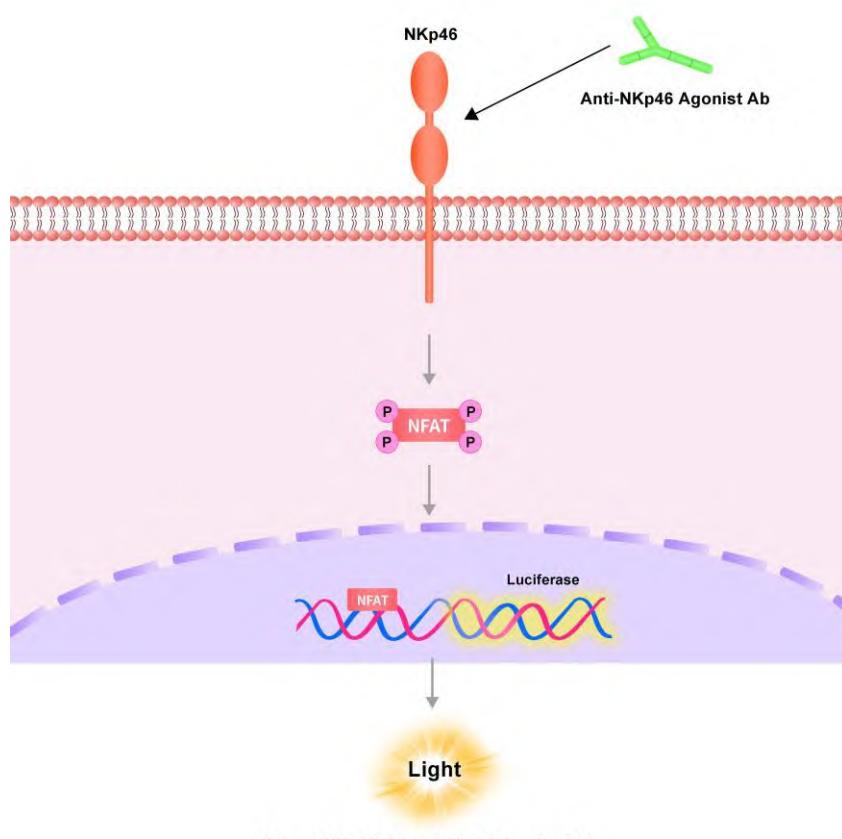
Catalog No.	Size
SCJUR-STF130	2 × (1 vial contains ~5×10 <sup>6</sup> cells)

### • Description

The Human NKp46 (Luc) Jurkat Reporter Cell was engineered to not only express the NFAT response element driving luciferase expressing systems, but also express the receptor full length human NKp46 (Uniprot: O76036-1). When cocultured with anti-human Nkp46 agonist antibody, the interaction of agonist antibody and Nkp46 on the surface of Human NKp46 (Luc) Jurkat Reporter Cell results in NFAT-mediated luminescence.

### • Application

- Screen for anti-human NKp46 agonist antibody.



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## • Cell Line Profile

Cell line	Human NKp46 (Luc) Jurkat Reporter Cell
Host Cell	Jurkat
Property	Suspension
Complete Growth Medium	RPMI-1640 + 10% FBS
Selection Marker	Puromycin (5 µg/mL) + Hygromycin B (20 µg/mL)
Incubation	37°C with 5% CO <sub>2</sub>
Doubling Time	16-20 hours
Transduction Technique	Lentivirus

## • Materials Required for Cell Culture

- RPMI Medium 1640 (Gibco, Cat. No. 11875-093)
- Fetal bovine serum (CellMax, Cat. No. SA211.02)
- Puromycin (InvivoGen, Cat. No. ant-pr-5b)
- Hygromycin B (Invitrogen, Cat. No. 10687010)

**Note:** For selection antibiotics, we highly recommend using the specified brand. The activity of antibiotics may vary between manufacturers, so if you choose to use a different brand, it is essential to validate whether the concentration recommended in the culture medium is suitable. Regardless of the brand used, we recommend maintaining a backup culture without selection antibiotics to avoid potential cell loss due to inappropriate antibiotic concentration.

- Penicillin-Streptomycin (Gibco, Cat. No. 15140-122)
- Complete Growth Medium: RPMI-1640 + 10% FBS, 1%P/S
- Culture Medium: RPMI-1640 + 10% FBS, Puromycin (5 µg/mL), Hygromycin B (20 µg/mL), 1%P/S
- Freeze Medium: 90% FBS, 10% (V/V) DMSO
- T-75 Culture flask (Corning, Cat. No. 430641)
- Cryogenic storage vials (SARSTEDT, Cat. No. 72.379.007)
- Thermostat water bath
- Centrifuge (Cence, Model: L550)
- Cell counter (MONWEI, Model: SmartCell200A Plus)
- CO<sub>2</sub> Incubator (Thermo, Model: 3111)
- Biological Safety Cabinet (Thermo, Model: 1389)

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## • Recovery

1. Thaw the vial by gently agitating it in a 37°C water bath. To minimize the risk of contamination, ensure the cap remains out of the water. Thawing should be completed quickly, typically within 3-5 minutes.
2. After thawing, promptly remove the vial from the water bath and decontaminate it by spraying with 70% ethanol. From this point onward, all operations must be performed under strict aseptic conditions.
3. Transfer the contents of the vial to a centrifuge tube containing 4.0 mL of complete growth medium.
4. Count viable cells and centrifuge at approximately 1000 rpm for 5 minutes.
5. Discard the supernatant and resuspend the cell pellet in an appropriate amount of fresh **complete growth medium**. Adjust the cell density of the suspension to  $1\times10^6$  viable cells/mL and transfer cells to an appropriate size vessel.
6. Incubate at 37°C with 5% CO<sub>2</sub> incubator.

## • Subculture

Cell viability may be low after thawing, and full recovery (viability >90%) may take up to 1-2 weeks. Once the cell density reaches approximately  $2\times10^6$  viable cells/mL, adjust the density to a range of  $2\times10^5$ - $5\times10^5$  viable cells/mL by either adding the fresh **culture medium** or replacing the existing culture medium. Avoid allowing the cell density to exceed  $3\times10^6$  cells/mL, as this may negatively impact cell performance in subsequent passages. T-75 flasks are recommended for subculturing.

• **Subculturing Frequency:** It is recommended to subculture every 3-4 days, adjusting the frequency based on the cell density in your specific culture system.

**Note:** After recovery, maintain the cells for 1-2 passages in the complete growth medium not containing the selection marker, if the cells are in good condition (viability >90%), transition to the culture medium containing the selection marker during subculturing.

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## • *Cryopreservation*

1. Count viable cells and harvest the cell suspension.
2. Centrifuge at 1000 rpm for 5 min at room temperature and resuspend cells in ice cold freezing medium to a concentration of  $5\times10^6$  to  $1\times10^7$  cells/mL.
3. Aliquot the cell suspension into cryogenic storage vials. Place the vials in a programmable cooler or an insulated box placed in a  $-80^{\circ}\text{C}$  freezer overnight, then transfer to liquid nitrogen storage for long-term storage.

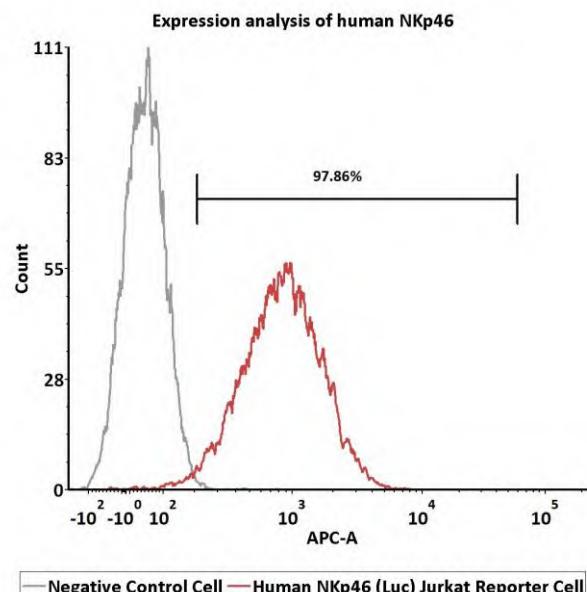
**Note:** It is recommended to establish a cell bank at the earliest possible passage for long-term use.

## • *Storage*

Cells must be received in a frozen state on dry ice and should be transferred to liquid nitrogen or a  $-80^{\circ}\text{C}$  freezer immediately upon receipt. If stored in a  $-80^{\circ}\text{C}$  freezer, it is recommended to limit the storage period to no more than two weeks. For long-term preservation, transfer the cells to liquid nitrogen is highly recommended.

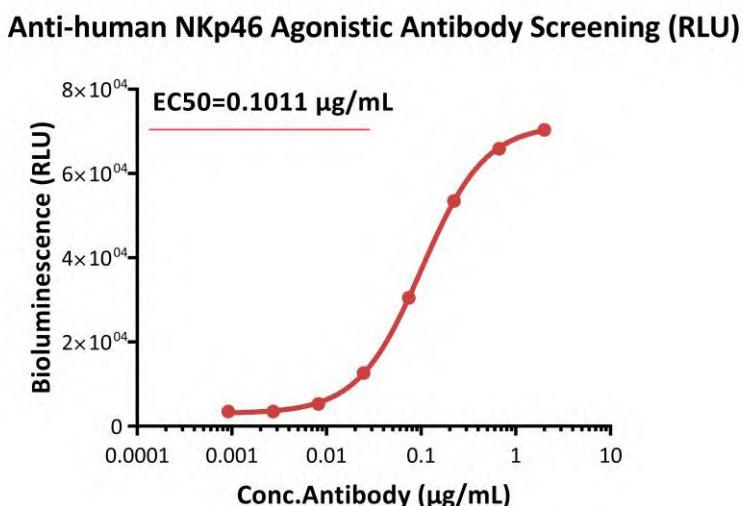
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## • Receptor Assay



**Fig1. Expression analysis of human NKp46 on Human NKp46 (Luc) Jurkat Reporter Cell by FACS.** Cell surface staining was performed on Human NKp46 (Luc) Jurkat Reporter Cell or negative control cell using APC-labeled Anti-human NKp46 antibody.

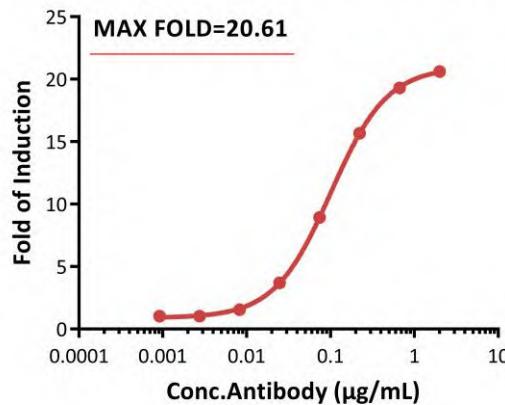
## • Application



**Fig2. Agonistic activity analysis of anti-human NKp46 antibody (RLU).** This reporter cell was incubated with serial dilutions of anti-human NKp46 antibody. The EC50 of anti-human NKp46 antibody was approximately 0.1011  $\mu\text{g/mL}$ .

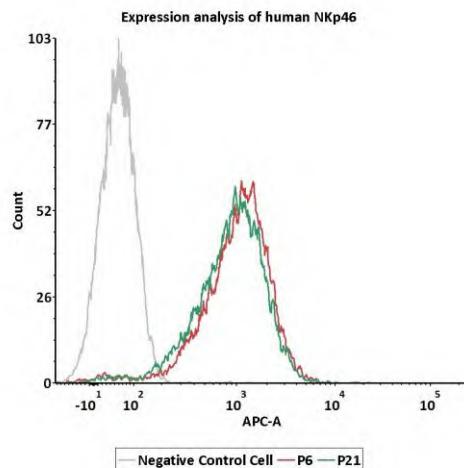
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## Anti-human NKp46 Agonistic Antibody Screening (FOLD)



**Fig3. Agonistic activity analysis of anti-human NKp46 antibody (FOLD).** This reporter cell was incubated with serial dilutions of anti-human NKp46 antibody. The max induction fold was approximately 20.61.

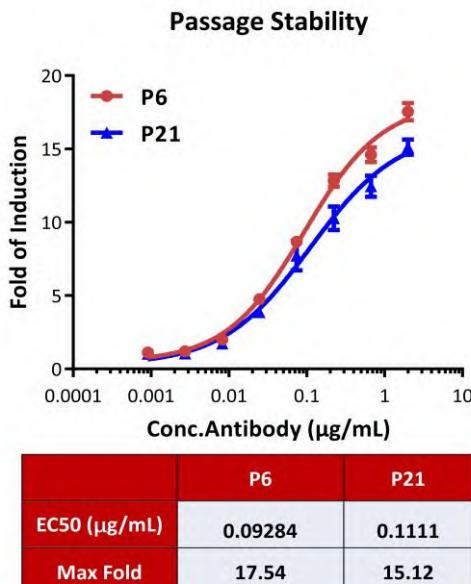
### • Passage Stability



Passage	MFI for NKp46 (APC)
P6	1059.76
P21	919.11

**Fig4. Passage stability analysis of receptor expression by FACS.** Flow cytometry surface staining of human NKp46 on Human NKp46 (Luc) Jurkat Reporter Cell demonstrates consistent mean fluorescent intensity across passage 6-21.

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**Fig4. Passage stability analysis by Signaling Bioassay.** The continuously growing Human NKp46 (Luc) Jurkat Reporter Cell was stimulated with serial dilutions of anti-human NKp46 antibody. Anti-human NKp46 antibody stimulated response demonstrates passage stabilization (fold induction and EC50) across passage 6-21.

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## • *Related Products*

<u>Products</u>	<u>Cat.No.</u>
HEK293/Human PD-L1, GFP Tag Stable Cell Line	CHEK-ATP002
HEK293/Human 4-1BB Ligand / TNFSF9 Stable Cell Line	CHEK-ATP039
HEK293/Human 4-1BB / TNFRSF9 Stable Cell Line	CHEK-ATP038
Human PD-1/LAG-3 (Luc) Jurkat Reporter	SCJUR-STF063
Human PD-1 (Luc) Jurkat Reporter Cell	SCJUR-STF064
Human LAG-3 (Luc) Jurkat Reporter Cell	SCJUR-STF065
Raji/Human PD-L1 Stable Cell Line	SCRAJ-STT075
Raji/Human CD155 Stable Cell Line	SCRAJ-STT076
CHO/Human LILRB4 Stable Cell Line	SCCHO-ATP087
HEK293/Human LILRB4 Stable Cell Line	CHEK-ATP088
Raji/Human HVEM Stable Cell Line	SCRAJ-STF108
CHO/Human LIGHT Stable Cell Line	SCCHO-ATP109
CHO/Human BTLA Stable Cell Line	SCCHO-ATP110
HEK293/Human PD-1 Stable Cell Line	CHEK-ATP143
HEK293/Human HVEM Stable Cell Line	CHEK-ATP147
HEK293/Human NKp46 Stable Cell Line	CHEK-ATP153
HEK293/Human ITPRIPL1 Stable Cell Line	CHEK-ATP203