

DATASHEET

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Myxovirus Resistance 1 (MX1) Antibody Pair

Catalogue No.:abx130433

Capture Antibody Clonality:

Detection Antibody Clonality:

Capture Antibody Conjugation:

Myxovirus Resistance 1 (MX1) Antibody Pair for use in Sandwich ELISA assay development. The 5 × 96 tests size contains:

- 600 µg Myxovirus Resistance 1 (MX1) mouse monoclonal capture antibody,
- 50 µg Myxovirus Resistance 1 (MX1) biotin-conjugated mouse monoclonal antibody,
- 10 µg Myxovirus Resistance 1 (MX1) standard.

It is recommended to use this antibody pair with abx098958 Antibody Pair Support Kit (Sandwich Method).

Target: Myxovirus Resistance 1 (MX1) Human Reactivity: **Tested Applications: ELISA Recommended dilutions:** Dilute the Capture Antibody 500-fold with Coating Buffer. Dilute the biotin-conjugated Detection Antibody 1000-fold with Detection Antibody Diluent. Optimal dilutions/concentrations should be determined by the end user. Form: Liquid (Capture Antibody and Detection Antibody) Reconstitution: Reconstitute the standard with 1.0 ml Standard Diluent. The volume, and therefore standard concentration, should be determined by the end user. Test Range: 1.56 ng/ml - 100 ng/ml Storage: Store at 2 to 8 °C for up to one month. Aliquot and store at -80 °C for up to one year. Avoid repeated freeze/thaw cycles. All solutions should be made fresh before the experiment. **Concentration:** 600 μg/0.24 ml (Capture Antibody), 50 μg/0.12 ml (Detection Antibody), 10 μg/vial (Standard) Standard Form: Lyophilized **ELISA Type:** Sandwich **Capture Antibody Host:** Mouse **Detection Antibody Host:** Mouse

Monoclonal

Monoclonal

Unconjugated



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Detection Antibody Conjugation: Biotin

Buffer: The capture and detection antibody both contain 0.1% sodium azide.

Directions for use: Bring all components to room temperature (18-25°C) and briefly spin or centrifuge the vials before use. Working solutions should be prepared and used immediately.

Recommended Procedure:

- 1. Dilute the Capture Antibody to working concentration using Coating Buffer. Immediately coat the 96-well plate with diluted Capture Antibody (100 µl per well). Seal the plate and incubate at 4 °C overnight or at 37 °C for 2 hours
- 2. Aspirate the wells and wash with Wash Buffer (350 µl per well) and allow to soak for 1-2 min. Remove the liquid by inverting and tapping the plate on to absorbent paper.
- 3. Block the plate with Blocking Buffer (200 µl per well) at 37 °C for 1.5 hours.
- 4. Repeat the aspiration/wash process in Step 2.
- 5. Add 100 µl of standards or sample into the appropriate wells. Cover with a plate sealer and incubate at 37 °C for 1 hour.
- 6. Repeat the aspiration/wash process in Step 2.
- 7. Add appropriately diluted Biotin-Conjugated Detection Antibody (100 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 1 hour.
- 8. Repeat the aspiration/wash process in Step 2.
- 9. Add appropriately diluted Streptavidin HRP (100 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 30 min.
- 10. Repeat the aspiration/wash process in Step 2.
- 11. Add Substrate Solution (90 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 10-20 min. Keep the plate in the dark and avoid exposure to light.
- 12. Add Stop Solution (50 µl per well). Tap the side of the plate to ensure thorough mixing.
- 13. Measure the absorbance immediately using a microplate reader set at 450 nm.

Note: This product is for research use only.