

Abbexa Ltd, Innovation Centre, Cambridge Science Park, Cambridge, CB4 0EY, UK
Telephone: +44 (0) 1223 755950 - Fax: +44 (0) 1223 755951 - E-Mail: info@abbexa.com

Uromodulin (UMOD) Antibody Pair

Catalogue No.: abx370002

Uromodulin (UMOD) Antibody Pair for use in Sandwich ELISA assay development. The 10 × 96 tests size contains:

- 220 µg Uromodulin (UMOD) mouse monoclonal capture antibody,
- 20 µg Uromodulin (UMOD) biotin-conjugated mouse monoclonal detection antibody,
- 800 ng Uromodulin (UMOD) standard.

It is recommended to use this antibody pair with [abx098958 Antibody Pair Support Kit \(Sandwich Method\)](#).

Target:	Uromodulin (UMOD)
Reactivity:	Human
Tested Applications:	ELISA
Recommended dilutions:	Dilute the Capture Antibody 140-fold with Coating Buffer. Dilute the biotin-conjugated Detection Antibody 1100-fold with Detection Antibody Diluent. Optimal dilutions/concentrations should be determined by the end user.
Form:	Liquid (Capture Antibody and Detection Antibody)
Reconstitution:	Reconstitute the standard with Standard Diluent. The volume, and therefore standard concentration, should be determined by the end user.
Test Range:	0.625 ng/ml - 40 ng/ml
Storage:	Store at 2 to 8 °C for up to one month. Aliquot and store at -80 °C for up to one year. Avoid repeated freeze/thaw cycles. All solutions should be made fresh before the experiment.
Concentration:	220 µg/0.7 ml (Capture Antibody), 20 µg/0.09 ml (Detection Antibody), 800 ng/vial (Standard)
Standard Form:	Lyophilized
ELISA Type:	Sandwich
Capture Antibody Host:	Mouse
Detection Antibody Host:	Mouse
Capture Antibody Clonality:	Monoclonal
Detection Antibody Clonality:	Monoclonal
Capture Antibody Conjugation:	Unconjugated

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Detection Antibody Conjugation: Biotin

Buffer: The capture and detection antibody both contain 0.1% sodium azide.

Directions for use: Bring all components to room temperature (18-25°C) and briefly spin or centrifuge the vials before use. Working solutions should be prepared and used immediately.

Recommended Procedure:

1. Dilute the Capture Antibody to working concentration using Coating Buffer. Immediately coat the 96-well plate with diluted Capture Antibody (100 µl per well). Seal the plate and incubate at 4 °C overnight or at 37 °C for 2 hours
2. Aspirate the wells and wash with Wash Buffer (350 µl per well) and allow to soak for 1-2 min. Remove the liquid by inverting and tapping the plate on to absorbent paper.
3. Block the plate with Blocking Buffer (200 µl per well) at 37 °C for 1.5 hours.
4. Repeat the aspiration/wash process in Step 2.
5. Add 100 µl of standards or sample into the appropriate wells. Cover with a plate sealer and incubate at 37 °C for 1 hour.
6. Repeat the aspiration/wash process in Step 2.
7. Add appropriately diluted biotin-conjugated Detection Antibody (100 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 1 hour.
8. Repeat the aspiration/wash process in Step 2.
9. Add appropriately diluted Streptavidin HRP (100 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 30 min.
10. Repeat the aspiration/wash process in Step 2.
11. Add Substrate Solution (90 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 10-20 min. Keep the plate in the dark and avoid exposure to light.
12. Add Stop Solution (50 µl per well). Tap the side of the plate to ensure thorough mixing.
13. Measure the absorbance immediately using a microplate reader set at 450 nm.

Note: This product is for research use only.