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DATASHEET

Neurofilament, Light Polypeptide (NEFL) Antibody Pair

Catalogue No.:abx370679

Neurofilament, Light Polypeptide (NEFL) Antibody Pair for use in Competitive ELISA assay development. The 10 × 96 tests size contains:

- 30 µg Neurofilament, Light Polypeptide (NEFL) rabbit polyclonal capture antibody,
- 200 µg Neurofilament, Light Polypeptide (NEFL) biotin-conjugated Competitor (NEFL and BSA coupling complexes),
- 30 µg Neurofilament, Light Polypeptide (NEFL) standard.

It is recommended to use this antibody pair with abx098959 Antibody Pair Support Kit (Competitive Method).

Target:	Neurofilament, Light Polypeptide (NEFL)
Reactivity:	Human
Tested Applications:	ELISA
Recommended dilutions:	Dilute the Capture Antibody 40-fold with Coating Buffer. Dilute the biotin-conjugated Competitor 300-fold with Biotin-Conjugated Competitor Diluent. Optimal dilutions/concentrations should be determined by the end user.
Form:	Liquid (Capture Antibody and biotin-conjugated Competitor)
Reconstitution:	Reconstitute the standard with Standard Diluent. The volume, and therefore standard concentration, should be determined by the end user.
Test Range:	123.5 pg/ml - 10000 pg/ml
Storage:	Store at 2 to 8 °C for up to one month. Aliquot and store at -80 °C for up to one year. Avoid repeated freeze/thaw cycles. All solutions should be made fresh before the experiment.
Standard Form:	Lyophilized
ELISA Type:	Competitive
Capture Antibody Host:	Rabbit
Capture Antibody Clonality:	Polyclonal
Capture Antibody Conjugation:	Unconjugated
Detection Antibody Conjugation:	Biotin
Buffer:	The capture antibody and biotin-conjugated competitor both contain 0.1% sodium azide.

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Directions for use:

Bring all components to room temperature (18-25°C) and briefly spin or centrifuge the vials before use. Working solutions should be prepared and used immediately.

Recommended Procedure:

1. Dilute the Capture Antibody to working concentration using Coating Buffer. Immediately coat the 96-well plate with diluted Capture Antibody (100 µl per well). Seal the plate and incubate at 4 °C overnight or at 37 °C for 2 hours
2. Aspirate the wells and wash with Wash Buffer (350 µl per well) and allow to soak for 1-2 min. Remove the liquid by inverting and tapping the plate on to absorbent paper.
3. Block the plate with Blocking Buffer (200 µl per well) at 37 °C for 1.5 hours.
4. Repeat the aspiration/wash process in Step 2.
5. Add 50 µl of standards or sample into the appropriate wells, followed by 50 µl of working biotin-conjugated Competitor. Cover with a plate sealer and incubate at 37 °C for 1 hour.
6. Repeat the aspiration/wash process in Step 2.
7. Add appropriately diluted Streptavidin HRP (100 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 30 min.
8. Repeat the aspiration/wash process in Step 2, for a total of 5 times.
9. Add Substrate Solution (90 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 10-20 min. Keep the plate in the dark and avoid exposure to light.
10. Add Stop Solution (50 µl per well). Tap the side of the plate to ensure thorough mixing.
11. Measure the absorbance immediately using a microplate reader set at 450 nm.

Note:

This product is for research use only.