

GFP-Expressing Human Colorectal Adenocarcinoma (LS 174T)
ORDER INFORMATION

Name of Cells: GFP-Expressing Human Colorectal Adenocarcinoma (LS 174T) (**GFP-LS 174T**)
Catalogue Number: **cAP-0045GFP**
Product Format: Frozen Vials
Cell Number: $>5 \times 10^5$ /vial

General Information

Human Colorectal Adenocarcinoma (LS 174T) cells are isolated from a Dukes's type B colorectal adenocarcinoma tissue by centrifugal force following enzymatic digestion. GFP-expressing Human Colorectal Adenocarcinoma (GFP-LS 174T) are selected from LS 174T cells after infected with lentiviruses expressing GFP with puromycin. The cells are shipped in a frozen vial with more than 5×10^5 cells/vial. The Universal Full Growth Medium (cAP-01B) are recommended to culture the cells and these cells have a minimum population doubling capacity > 20 when cultured following the detailed protocol described below.

Characterization of the cells

GFP-LS 174T are negative for HIV-1, HBV, HCV, and mycoplasma and with a minimum $>99.9\%$ GFP positive cells

Product Use: **GFP-LS 174T** are for **Research Use Only**.

Shipping status: Frozen vials in dry ice package.

Handling of Arriving Cells

When you receive the cells in a frozen vial, you can transfer the vial of cells into a -80°C freezer for short period storage or a liquid nitrogen tank for long term storage. Thaw the cells in a 37°C water bath, and then transfer the cells in a T25 flask pre-coated with Quick coating solution (cAP-01) as described in details in Subculture Protocol.

1. Subculture Protocol:

- A) Pre-coating of T25 flasks: Add 2ml of Quick Coating Solution (**cAP-01**) into one T25 flask and make sure whole surface of the flask is covered with the coating solution. Five minutes later, dispose excessive Quick Coating Solution by aspiration and the flask is ready to be used (no need for overnight incubation when using Quick Coating Solution).
- B) Rinse the cells in T25 flask with 5ml HBSS (**Room Temperature, RT**) twice.
- C) Add 2ml of Trypsin/EDTA (**RT**) (cAP-23) into one T25 flask (make sure the whole surface of the T25 flask is covered with Trypsin/EDTA), and gently dispose the excessive Trypsin/EDTA solution **within 20 seconds** with aspiration.
- D) Leave the T25 flask with the cells at **RT** for 1 minute (the cells usually will detach from the surface within 1-2 minutes). You can monitor the cells under microscope and when most of cells become rounded up, hit the flask against the bench surface, and the cells will move on the surface of the flask when monitoring under microscope.
- E) Add 5ml Trypsin Neutralization Buffer and spin the cells down with 800g for 5 minutes.
- F) Re-suspend the cell pellet with 15ml of Universal Full Growth Medium and the cell suspension is transferred directly into 3 pre-coated T25 flasks (5ml each, and the cells are sub-cultured at 1:3 ratios)
- G) Change medium every 2-3 days and cells usually become confluent within 7-8 days (when split at a 1:3 ratio).

Related products

Quick Coating Solution	cAP-01	240ml	Angio-Proteomie
Pericyte Growth Medium	cAP-09	500ml	Angio-Proteomie
Universal Full Growth Medium	cAP-01B	500ml	Angio-Proteomie
Pericyte Basal Medium	cAP-09C	500ml	Angio-Proteomie
HBSS w/o Ca^{2+} , Mg^{2+}	cAP-11	100ml	Angio-Proteomie
Trypsin/EDTA Solution	cAP-23	100ml	Angio-Proteomie
Trypsin Neutralization Solution	cAP-28	100ml	Angio-Proteomie

Caution: Handling human tissue derived products is potentially bio-hazardous. Although each cell strain is tested negative for HIV, HBV and HCV DNA, diagnostic tests are not necessarily 100% accurate; therefore, proper precautions must be taken to avoid inadvertent exposure. Always wear gloves and safety glasses when working these materials. Never mouth pipette. We recommend following the universal procedures for handling products of human origin as the minimum precaution against contamination.