

**Human Gastric Carcinoma N87 Cells**
**ORDER INFORMATION**

**Name of Cells:** Human Gastric Carcinoma N87 Cells (**N87 cells**)  
**Catalogue Number:** **cAP-0058**  
**Product Format:** Frozen Vials  
**Cell Number:** >5 x 10<sup>5</sup>/vial

**General Information**

Human Gastric Carcinoma N87 Cells (N87 cells) were established from the liver metastatic site of a male gastric carcinoma patient. The cells can be cultivated with Universal Full Growth Medium (cAP-01B) and they have a minimum population doubling capacity > 12 when cultured following the detailed protocol described below.

**Characterization of the cells**

**N87 cells** are negative for HIV-1, HBV, HCV, and mycoplasma

**Product Use:** N87 cells are for **Research Use Only.**

**Shipping status:** Frozen vials in dry ice package.

**Handling of Arriving Cells**

When you receive the cells in a frozen vial, you can transfer the vial of cells into a -80°C freezer for short period storage or a liquid nitrogen tank for long term storage. Thaw the cells in a 37°C water bath, and then transfer the cells in a T25 flask pre-coated with human Fibronectin (20ug/ml, **cAP-42**) as described in details in Subculture Protocol.

**1. Subculture Protocol:**

- A) Pre-coating of T25 flasks: Add 1ml of Fibronectin (**cAP-42**) into one T25 flask and make sure whole surface of the flask is covered with the coating solution and leave flask at room temperature for overnight or 37°C for 1 hour. Dispose excessive fibronectin by aspiration.
- B) Rinse the cells in T25 flask with 5ml HBSS (**Room Temperature, RT**) once.
- C) Add 2ml of Trypsin/EDTA (**RT**) (cAP-23) into one T25 flask (make sure the whole surface of the T25 flask is covered with Trypsin/EDTA), and gently dispose the excessive Trypsin/EDTA solution **within 20 seconds** with aspiration.
- D) Leave the T25 flask with the cells at **RT** for 1 minute (the cells usually will detach from the surface within 1-2 minutes). You can monitor the cells under microscope and when most of cells become rounded up, hit the flask against the bench surface, and the cells will move on the surface of the flask when monitoring under microscope.
- E) Add 5ml Trypsin Neutralization Buffer and spin the cells down with 800g for 5 minutes.
- F) Re-suspend the cell pellet with 15ml of Universal Full Growth Medium and the cell suspension is transferred directly into 3 pre-coated T25 flasks (5ml each, and the cells are sub-cultured at 1:3 ratios)
- G) Change medium every 2-3 days and cells usually become confluent within 7-8 days (when split at a 1:3 ratio).

**Related products**

|                                              |         |       |                 |
|----------------------------------------------|---------|-------|-----------------|
| Quick Coating Solution                       | cAP-01  | 240ml | Angio-Proteomie |
| <b>Universal Full Growth Medium</b>          | cAP-01B | 500ml | Angio-Proteomie |
| Pericyte Basal Medium                        | cAP-09C | 500ml | Angio-Proteomie |
| HBSS w/o Ca <sup>2+</sup> , Mg <sup>2+</sup> | cAP-11  | 100ml | Angio-Proteomie |
| Trypsin/EDTA Solution                        | cAP-23  | 100ml | Angio-Proteomie |
| Trypsin Neutralization Solution              | cAP-28  | 100ml | Angio-Proteomie |

**Caution:** Handling human tissue derived products is potentially bio-hazardous. Although each cell strain is tested negative for HIV, HBV and HCV DNA, diagnostic tests are not necessarily 100% accurate; therefore, proper precautions must be taken to avoid inadvertent exposure. Always wear gloves and safety glasses when working these materials. Never mouth pipette. We recommend following the universal procedures for handling products of human origin as the minimum precaution against contamination.