

## Boster's PCR PreMix (No-Dye) 8-tube strips, 20 µl x 96 reactions

SKU: MB1007

### I. Introduction

The Boster's PCR PreMix contains DNA polymerase, dNTPs and reaction buffer in a premixed format that is freeze-dried into an individual packet.

### II. Storage

Store at -20°C

### III. Application

- Routine PCR
- Primer Extension
- TA Cloning
- Gene Sequencing

### III. Content

Components	20µl Reaction	50µl Reaction
Top DNA Polymerase	1U	2.5U
dNTP(dATP,dCTP,dGTP,dTTP)	Each 250 µM	Each 250 µM
Reaction Buffer with 1.5mM MgCl <sub>2</sub>	1X	1X
Stabilizer	0	0

### IV. Advantages

- **Speed:** Substantial reduction in PCR setup time.
- **Stability:** PCR Premix is the powerful technology for convenient and easy to perform DNA amplification. It contains DNA polymerase, dNTPs, and reaction buffer in a premixed format, freeze-dried into a pellet. The patented chemical stabilizer of this product enables to maintain the activity of pre mixture for over 2 years in the freezer. It ensures superior amplification efficiency with experiment stability and uniform activity of polymerase in the process of PCR.
- **Reproducibility and Yield:** Batch manufacturing under strict ISO 9001 quality control conditions guarantees reproducibility. Minimal handling during reaction set up provides improved accuracy.
- **Simplicity:** Each product contains an application specific enzyme in an easy to re-suspended, lyophilized premix of dNTPs, reaction buffer, and a stabilizer. The premix is ready to use, thus offering virtually no reaction set up time.

### V. Notice to Purchaser

This enzyme is specifically optimized for increasing base incorporation rate by inactivating 5' → 3' exonuclease activity. Therefore, this is not recommended to use for Real Time PCR using Taqman® probe.

### VI. Protocol

1. Add template DNA and primers into PCR PreMix tubes.
2. Add distilled water into PCR PreMix tubes to a total volume of 20 µl or 50µl. Do not calculate the dried pellet.

Ex) Reaction Mix

Components	20µl Reaction	50µl Reaction
Template DNA	Variable (1~10 µl)	Variable (1 ~ 25 µl)
Forward Primer (10 pmole/µl)	0.5µl~2µl	1 µl ~ 5µl
Reverse Primer (10 pmole/µl)	0.5µl~ 2 µl	1 µl ~ 5µl
D.W.	Variable (6 µl ~ 18 µl)	Variable (15µl ~ 47µl)
Total Volume	20µl	50µl

- Note 1: Amount of template

Template DNA	Amount of template	
	20µl Reaction	50µl Reaction
Bacteriophage λ, Plasmid DNA	100fg~200ng	100fg~500ng
Total Genomic DNA	1ng ~500ng	1ng ~ 1µg

3. Dissolve the lyophilized blue pellet by flicking your finger or pipetting, and briefly spin down
4. (Optional) If Necessary, overlay mineral oil. This step is unnecessary when using a thermal cycler with top heating.
5. Perform the reaction under the following conditions.

- In case of routine PCR

Step	Temp	Time	Cycles
Pre-Denaturation	95°C	5 min	1 Cycle
Denaturation	95°C	20 sec	25~35 cycles
Annealing	45~65°C	20 sec	
Extension	72°C	30sec ~ 1min/kb	
Final Extension	72°C	Optional Normally 3~6mins	1 cycle

- In case Primer's T<sub>m</sub> value is more than 65°C or PCR product size is more than 5kb.

Step	Temp	Time	Cycles
Pre-Denaturation	95°C	5 min	1 Cycle
Denaturation	95°C	20 sec	30~35 cycles
Anneal/Extension	68°C	1 min/kb	
Final Extension	72°C	Optional. Normally 3~5 min	1 Cycle

6. Maintain the reaction at 4°C~8°C after cycling. The samples can be stored at -20°C until use.
7. Load samples on agarose gel with adding a loading-dye mixture and perform electrophoresis.