



## **TCF/LEF Luciferase Reporter- HEK293 Cell Line**

**Catalog number: RC1019**

This package insert must be read in its entirety before using this product.  
For research use only. Not for use in diagnostic procedures.

### **TCF/LEF Luciferase Reporter-HEK293 Cell Line**

**Catalog Number:** RC1019, **Storage:** Immediately upon receipt, store in liquid nitrogen. (Ship on dry ice.)

**Contents:** Each vial contains  $2 \sim 3 \times 10^6$  cells in 1 ml of 90% FBS + 10% DMSO.

**Description:** The TCF/LEF Luciferase Reporter cell line is a stably transfected HEK293 cell line which expresses Renilla luciferase reporter gene under the control of the TCF/LEF response element. This cell line is designed to monitor the transcriptional activity of TCF/LEF and can be used for studying Wnt signaling pathways as well as screening of activators or inhibitors that affect the TCF/LEF transcriptional activity.

**Applications:** Functional Assay

**Application Notes:** Functional Assay, detecting the transcriptional activity of TCF/LEF

**Application Details:** Dilute the sample so that the expected range of concentrations fall within the detection range of this kit.

If the expected range of concentration is unknown, a pilot test should be conducted to decide the optimal dilution ratio for your samples.

Some PubMed article(s) citing the expression level of this target are as follows:

Boster Bio's internal QC testing used:

## Application:

Monitor the TCF/LEF induction activity. Screen for activators or inhibitors of the TCF/LEF signaling pathway.

## Culture conditions:

Cells should be grown at 37°C with 5% CO<sub>2</sub> using DMEM medium supplemented with 10% FBS and 1% Pen/Strep, plus 3 µg/ml of Puromycin. It is recommended to quickly thaw the frozen cells upon receipt or from liquid nitrogen in a 37°C water-bath, transfer to a tube containing 10 ml of growth medium without Puromycin, spin down cells, resuspend cells in pre-warmed growth medium without Puromycin, transfer resuspended cells to T25 flask and culture in 37°C-CO<sub>2</sub> incubator. Leave the T25 flask in the incubator for 2~4 days without disturbing or changing the medium until cells completely recover viability and become adherent. Once cells are over 90% adherent, remove growth medium and passage the cells through trypsinization and centrifugation. At first passage, switch to growth medium containing Puromycin. Cells should be split before they reach complete confluence. To passage the cells, detach cells from culture vessel with Trypsin/EDTA, add complete growth medium and transfer to a tube, spin down cells, resuspend cells and seed appropriate aliquots of cells suspension into new culture vessels. Subcultivation ration = 1:10 to 1:20 weekly.

## Functional validation:

A. Response of TCF/LEF HEK293 cells to lithium chloride (LiCl) (Figure 1). 1. Harvest TCF/LEF HEK293 cells and seed cells into a white solid-bottom 96-well microplate in 100 µl of growth medium at  $5 \times 10^4$  cells/well. 2. Incubate cells at 37°C in a CO<sub>2</sub> incubator for overnight. 3. The next day, stimulate cells with various concentrations of LiCl. 4. Incubate at 37°C in a CO<sub>2</sub> incubator for 6-16 hours.

## TCF/LEF Luciferase Reporter-HEK293 Cell Line (RC1019) Images

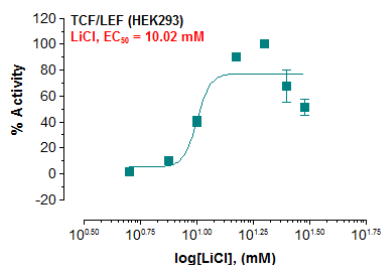


Fig-1: Induction of TCF/LEF activity by LiCl in TCF/LEF HEK293 cells.

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