Product sheet

KG-1 Cells | 300208

General information

Description	KG-1 is a human acute myelogenous leukemia (AML) cell line derived from the bone marrow of an adult patient with erythroleukemia. This cell line is a valuable model for studying hematopoietic differentiation and leukemia, particularly due to its unique characteristics, including the expression of several hematopoietic markers. KG-1 cells are classified as immature myeloid cells that resemble early progenitor cells, which makes them a useful tool for investigating the early stages of myeloid lineage commitment and the molecular mechanisms driving leukemogenesis. KG-1 cells exhibit a high degree of plasticity, which allows them to differentiate into various hematopoietic lineages under the right experimental conditions. This characteristic is particularly important for research in understanding the regulation of hematopoiesis and the development of therapeutic strategies aimed at targeting leukemic stem cells. Additionally, KG-1 cells are known to express markers such as CD34, HLA-DR, and CD13, which are critical in both normal and malignant hematopoiesis, making them an excellent model for flow cytometry and other immunophenotyping studies. KG-1 has also been employed in drug discovery and toxicity testing, where its responsiveness to differentiation agents and chemotherapeutic drugs can be evaluated. As with all in vitro models, it is important to recognize that KG-1 cells are for research use only and not suitable for therapeutic or in vivo applications.
Organism	Human
Tissue	Bone marrow
Disease	Acute myelogenous leukemia
Synonyms	KG1

Characteristics

Age	59 years
Gender	Male
Ethnicity	Caucasian
Cell type	Myeloblast
Growth properties	Suspension

Regulatory Data

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Citation	KG-1 (Cytion catalog number 300208)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_0374

Biomolecular Data

Antigen expression	HLA A30, A31, B35, Cw4
Isoenzymes	G6PD, B, PGM1, 1-2, PGM3, 0, ES-D, 1, Me-2, 1, AK-1, 0, GLO-1, 2
Viruses	EBNA (EBNA): negative
Reverse transcriptase	Negative

Handling

Culture Medium	IMDM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 25 mM HEPES, w: 1.0 mM Sodium pyruvate, w: 3.024 g/L NaHCO3 (Cytion article number 820800a)
Supplements	Supplement the medium with 10% FBS
Doubling time	45 hours
Subculturing	Transfer the cell suspension into sterile centrifuge tubes. Collect the cells by spinning down at 300xg for 3 minutes. Discard the supernatant and resuspend the pelleted cells in fresh cell culture medium. Adjust to an optimal cell density between 1 - 3 x 10^5 cells/ml. Split the cells when a maximum cell density of 1 - 2 x 10^6 cells/ml is reached.
Fluid renewal	Every 3 days
Post-Thaw Recovery	Allow the cells to recover from the freezing process for at least 24 hours.
Freeze medium	As a cryopreservation medium, use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality control / Genetic profile / HLA

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.