Product sheet

B16-F0 Cells | 300308

General information

Description	The B16-F0 cell line is a murine melanoma cell line derived from the B16 mouse melanoma. This cell line is widely used in cancer research due to its high metastatic potential and ability to form tumors when injected into syngeneic mice. B16-F0 cells are particularly useful for studying the molecular mechanisms underlying melanoma progression and metastasis, as well as for testing the efficacy of anti-cancer drugs and therapeutic interventions in preclinical models. Notably, the B16-F0 cell line is the parent cell line from which other variants, such as B16-F1, B16-F10, and B16-BL6, have been derived through selective procedures aimed at enhancing specific metastatic properties. B16-F0 cells exhibit a typical epithelial morphology and grow adherently in culture. They are known to express various melanoma-associated antigens, making them a valuable tool for immunological studies and for developing melanoma vaccines. Additionally, these cells are often employed in studies involving gene expression, signaling pathways, and the tumor microenvironment. Researchers utilize B16-F0 cells to explore the interactions between melanoma cells and the immune system, particularly focusing on immune evasion and suppression mechanisms. The characterization of B16-F0 and its derived lines provides a comprehensive framework for understanding the invasive and metastatic behaviors of melanoma, with B16-F1, B16-F10, and B16-BL6 each representing stages of increasing metastatic and invasive activity, thereby serving as critical models in the study of cancer progression and therapeutic response.
Organism	Mouse
Tissue	Skin
Disease	Mouse melanoma
Synonyms	B16/F0, B16F0

Characteristics

Gender	Male
Morphology	mixture of spindle-shaped and epithelial-like cells
Cell type	Epithelial
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	B16-F0 (Cytion catalog number 300308)

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Biosafety level	1

Expression / Mutation

Tumorigenic	Yes, in syngeneic mice
Products	Melanin

Handling

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 1.5 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Freeze medium	CM-1 (Cytion catalog number 800100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.