

RPMI 1788 Cells | 300318

General information

Description	The RPMI 1788 cell line was derived from the peripheral blood of an apparently normal patient. The cells are EBNA positive.
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Organism	Human
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Tissue	Peripheral blood
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Synonyms	RPMI-1788, RPMI1788, Roswell Park Memorial Institute 1788, GM02131, GM2131, GM02131A, GM17219
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Characteristics

Age	33 years
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Gender	Male
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Ethnicity	Caucasian
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Morphology	Round cells
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Cell type	B lymphoblast
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Growth properties	Adherent
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Regulatory Data

Citation	RPMI 1788 (Cytion catalog number 300318)
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Biosafety level	2
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NCBI_TaxID	9606
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CellosaurusAccession	CVCL_2710
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Biomolecular Data

Antigen expression	HLA A2, Aw33, B7, B14
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Isoenzymes G6PD, B

Viruses EBNA-pos

Reverse transcriptase Negative

Products IgM (lambda light chain), lymphotoxin (LT) also known as tumor necrosis factor beta (TNF-beta, TNF beta)

Karyotype Human male, hypodiploid, stable

Handling

Culture Medium RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO₃ (Cytion article number 820700a)

Supplements Supplement the medium with 10% FBS

Subculturing Maintain cultures by periodically adding or replacing the medium. Initiate cultures with a density of 5×10^5 cells/ml and keep the cell concentration within the range of 3×10^5 to 1×10^6 cells/ml for optimal growth.

Seeding density 1×10^5 cells/mL

Post-Thaw Recovery Low viability after thawing. Good recovery after 8 days

Freeze medium As a cryopreservation medium, use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C, 5% CO₂, humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality control / Genetic profile / HLA

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Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.