Product sheet

TK6 Cells | 300357

General information

Description	TK6 is a lymphoblast cell line derived from the spleen of a 5-year-old male diagnosed with hereditary spherocytosis. This cell line is particularly notable for being heterozygous at the thymidine kinase (TK) locus, which underpins its utility in genetic research. The heterozygosity at the TK locus allows the TK6 cells to serve as a sensitive model for detecting forward mutations, providing a robust platform for mutagenicity testing and genetic toxicology studies. The cells are employed extensively in assays designed to quantitatively detect forward mutations at three loci, including resistance to trifluorothymidine at the tk locus. This capability makes TK6 an invaluable tool in the pharmaceutical and chemical industries for evaluating the mutagenic potential of new compounds. The cell line's unique genetic background and its relevance to disease make it a critical resource for studies focused on understanding mutation processes and evaluating the cytogenetic effects of chemical exposures in a controlled environment.
Organism	Human
Tissue	Spleen
Synonyms	TK-6, H2BT

Characteristics

Age	5 years
Gender	Male
Cell type	Lymphoblast
Growth properties	Suspension

Regulatory Data

Citation	TK6 (Cytion catalog number 300357)
Biosafety level	2
NCBI_TaxID	9606
CellosaurusAccession	CVCL_0561

Biomolecular Data

Handling

Culture Medium	RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Supplements	Supplement the medium with heat inactivated 10% FBS, 2,5% horse serum
Subculturing	Initiate cultures with a cell density of 5 x 10^5 cells/ml and maintain them within the range of 1 x 10^5 to 1 x 10^6 cells/ml. For subculturing, transfer the cell suspension to a fresh cell culture flask pre-filled with the correct volume of fresh culture medium.
Seeding density	1 x 10^5 cells/mL
Fluid renewal	2 to 3 times per week
Freeze medium	As a cryopreservation medium, use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality control / Genetic profile / HLA

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.