Product sheet

NCI-H196 Cells | 300390

General information

Description	NCI-H196 is a small-cell lung cancer (SCLC) cell line used to study the mechanisms of cancer progression, chemotherapy resistance, and cellular responses to oxidative stress. Research involving NCI-H196 has demonstrated its sensitivity to the cytotoxic effects of pyrrolidine dithiocarbamate (PDTC), a pro-oxidant agent. PDTC induces S-phase cell cycle arrest and significantly reduces the viability of NCI-H196 cells in a dose-dependent manner. This cytotoxicity is attributed to the induction of oxidative stress, as evidenced by increased reactive oxygen species (ROS) and changes in the expression of oxidative stress-related genes. The addition of antioxidants like N-acetyl-L-cysteine (NAC) can effectively reverse PDTC-induced cytotoxicity, confirming the role of oxidative stress in cell death. Further studies have shown that PDTC enhances the cytotoxicity of cisplatin, a first-line chemotherapy drug used for SCLC treatment. Combining low doses of cisplatin with non-toxic concentrations of PDTC leads to synergistic cytotoxicity in NCI-H196 cells. This combination therapy is believed to be effective due to PDTC?s downregulation of ATP7A, a copper efflux transporter associated with cisplatin resistance. By inhibiting ATP7A, PDTC may increase intracellular copper and sensitize NCI-H196 cells to cisplatin, highlighting its potential as an adjunct therapy for SCLC.
Organism	Human
Tissue	Lung
Disease	Lung small cell carcinoma
Metastatic site	Pleural effusion
Applications	3D cell culture, Cancer research
Synonyms	NCI-H196, H-196, NCIH196

Characteristics

Age	68 years
Gender	Male
Ethnicity	European
Growth properties	Adherent

Identifiers / Biosafety / Citation

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Citation	NCI-H196 (Cytion catalog number 300390)
Biosafety level	1

Expression / Mutation

Handling

Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Freeze medium	CM-1 (Cytion catalog number 800100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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STR profile

Amelogenin: x,y
CSF1PO: 10
D13S317: 9
D16S539: 11
D5S818: 12
D7S820: 10,11
TH01: 6
TPOX: 11
vWA: 19
D3S1358: 15
D18S51: 17,19
Penta E: 8,12
Penta D: 10
D8S1179: 13,15
FGA: 22,23
D6S1043: 13
D2S1338: 17,2
D12S391: 19
D19S433: 14