Product sheet

Panc 10.05 Cells | 300599

General information

Description	The Panc 10.05 cell line is a human pancreatic ductal adenocarcinoma (PDAC) cell line, which is used in studies exploring the biology of pancreatic cancer and potential therapeutic interventions. Like other PDAC cell lines, Panc 10.05 cells are often employed in research focused on understanding the tumor microenvironment, cancer cell proliferation, and mechanisms of resistance to chemotherapy. This cell line, along with others such as BxPC-3 and HPAF-II, has been used to test the effects of novel anti-cancer agents, including iron chelators like deferasirox (DFX). Studies have shown that DFX exhibits dose-dependent antiproliferative activity against Panc 10.05 cells by inducing apoptosis and arresting the cell cycle in the S-phase. Panc 10.05 has also been used to explore the role of inflammation and immune modulation in pancreatic cancer. For example, in co-culture models with macrophages, Panc 10.05 cells were shown to interact with tumor-associated macrophages (TAMs), creating a pro-inflammatory microenvironment. This interaction leads to the activation of the NLRP3 inflammasome, which plays a critical role in promoting tumor growth and immune evasion. Inhibition of the NLRP3 inflammasome by specific inhibitors like MCC950 has been shown to reduce the pro-inflammatory cytokine response and tumor cell proliferation, highlighting its potential as a therapeutic target. Overall, the Panc 10.05 cell line serves as a robust model for studying both the direct effects of therapeutic agents and the complex interactions within the tumor microenvironment in pancreatic cancer, aiding in the development of new treatment strategies for this aggressive disease.
Organism	Human
Tissue	Pancreas
Disease	Pancreatic ductal adenocarcinoma
Applications	3D cell culture, Cancer research
Synonyms	Panc-10.05, Panc10.05, PANC-10-05, PANC 1005, PANC1005, Panc1005, Pa16C, PL12, PL-12

Characteristics

Age	81 years
Gender	Male
Ethnicity	European
Morphology	Epithelial
Cell type	Epithelial cell

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Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	Panc 10.05 (Cytion catalog number 300599)
Biosafety level	1

Expression / Mutation

Protein expression	cytokeratin 7, cytokeratin 18
Antigen expression	MHC class I +, MHC class II -
Oncogenes	K-ras+
Tumorigenic	Yes, forms tumors in nude or SCID mice

Handling

Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Medium supplements	Supplement the medium with 20% heat-inactivated FBS, 10 Units/mL human recombinant insulin
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Freeze medium	CM-1 (Cytion catalog number 800100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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STR profile

Amelogenin: x,x
CSF1PO: 12
D13S317: 12
D16S539: 9,12
D5S818: 13
D7S820: 8,9
TH01: 6,9.3
TPOX: 11
vWA: 16
D3S1358: 14
D21S11: 30
D18S51: 15
Penta E: 11,13
Penta D: 12
D8S1179: 13,14
FGA: 20
D6S1043: 17
D2S1338: 17,18
D12S391: 17,2
D19S433: 13,14