

AML12 Cells | 300643

General information

Description

AML12 cells, also known as Alpha Mouse Liver 12 cells, are a non-tumorigenic epithelial cell line derived from the liver of a transgenic mouse. These cells were initially developed to provide a suitable in vitro model for studying the hepatocyte function and liver biology of the adult mouse. AML12 cells express characteristics typical of differentiated hepatocytes, including the production of albumin, transferrin, and other liver-specific proteins, making them an invaluable resource for research in toxicology, drug metabolism, and liver disease.

The cell line was established from hepatocytes isolated from a mouse harboring a transgene for human transforming growth factor alpha (TGF-alpha), under the control of the mouse metallothionein-I promoter. This genetic alteration contributes to the immortalization of the cells without disrupting their differentiated state. AML12 cells maintain a stable phenotype and karyotype under standard cell culture conditions, which includes a unique requirement for dexamethasone and insulin-transferrin-selenium in the growth medium to promote proliferation and maintain hepatocyte-specific functions.

Organism

Mouse

Tissue

Liver

Applications

3D cell culture, High-throughput screening, Toxicology

Synonyms

AML-12, AML 12, Alpha Mouse Liver 12

Characteristics

Age

3 months

Gender

Male

Morphology

Epithelial

Cell type

Hepatocyte

Growth properties

Adherent

Identifiers / Biosafety / Citation

Citation

AML12 (Cytion catalog number 300643)

Biosafety level

1

AML12 Cells | 300643**Expression / Mutation**

Products	The cells express high levels of human TGF alpha and lower levels of mouse TGF alpha.
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Handling

Culture Medium	DMEM:Ham's F12, w: 3.1 g/L Glucose, w: 1.6 mM L-Glutamine, w: 15 mM HEPES, w: 1.0 mM Sodium pyruvate, w: 1.2 g/L NaHCO ₃ (Cytion article number 820400a)
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Medium supplements	Supplement the medium with 10% FBS, 10 microgram/mL insulin, 5.5 microgram/mL transferrin, 5 ng/mL selenium, 40 ng/mL dexamethasone
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Passaging solution	Accutase
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Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
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Freeze medium	CM-1 (Cytion catalog number 800100)
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Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.