

**Human Mesenchymal Stem Cells - Chorion Villi | 300646****General information****Description**

Human Mesenchymal Stem Cells (MSCs) derived from chorion villi represent a highly versatile population of multipotent stromal cells capable of differentiating into multiple lineages, including adipocytes, osteoblasts, and chondrocytes. These cells are isolated from the chorionic villi, a part of the placenta that plays a critical role in maternal-fetal exchange. Chorion villi are unique in that they are composed of both fetal and maternal tissues, providing a distinct microenvironment that contributes to the robust self-renewing and differentiation capabilities of the MSCs derived from this source. The MSCs from chorion villi exhibit a more primitive phenotype compared to MSCs derived from adult tissues, often displaying a higher proliferation rate and broader differentiation potential. These characteristics make them particularly valuable for research in regenerative medicine, tissue engineering, and disease modeling.

These MSCs have been rigorously demonstrated *in vitro* to differentiate into adipocytes, osteoblasts, and chondrocytes when cultured in lineage-specific differentiation media, underscoring their potential for applications in tissue regeneration and disease modeling. The unique origin of these cells from the chorionic villi imparts them with specific immunomodulatory properties, which may differ from MSCs derived from other sources such as bone marrow or adipose tissue. This distinction is crucial for studies focusing on immune-related conditions or developing allogeneic cell therapies.

MSCs are cryopreserved at early passages in a specialized cryomedium, ensuring their viability and functionality post-thaw. Each cryovial contains a minimum of  $1 \times 10^6$  cells with a viability rate ranging between 92% and 95%, as determined by the Trypan Blue dye exclusion test. These cells are sourced from healthy donors who have provided informed consent, ensuring ethical collection practices. Each batch undergoes stringent quality control assessments, including thorough testing for cell identification, purity, potency, and viability. These measures guarantee that the cultured MSCs are of high quality and appropriate for research applications, excluding therapeutic or *in vivo* use.

**Organism** Human**Tissue** Chorion Villi**Applications** Drug testing, regenerative medicine, disease research**Characteristics****Age** Please inquire**Gender** Please inquire**Ethnicity** Caucasian**Morphology** Well-spread spindle shaped, fibroblast-like morphology for at least within 5 passages. Fewer than 2% cells exhibit spontaneous myofibroblast-like morphology within each passage.

**Human Mesenchymal Stem Cells - Chorion Villi | 300646****Cell type** Stem cell**Growth properties** Adherent**Identifiers / Biosafety / Citation****Citation** Human Mesenchymal Stem Cells, Whartons Jelly (HMSC-WJ) (Cytion catalog number 300685)**Biosafety level** 1**Expression / Mutation****Antigen expression** A comprehensive panel of markers, including CD73/CD90/CD105 (positive) and CD14/CD34/CD45/HLA-DR (negative), are used in flow cytometry analysis to identify cultivated MSCs (P2-P3) prior to cryopreservation. These markers are recommended by the ISCT MSC committee.**Viruses** Donor is negative for HBV (PCR), Treponema pallidum (PCR), and HIV-1/2 (IFA). Cells are negative for HBV, HCV, HSV1, HSV2, CMV, EBV, HHV6, Toxoplasma gondii, Treponema pallidum, Chlamydia trachomatis, Ureaplasma urealyticum, and Ureaplasma parvum.**Handling****Culture Medium** Alpha MEM, w: 2.0 mM stable Glutamine, w/o: Ribonucleosides, w/o: Deoxyribonucleosides, w: 1.0 mM Sodium pyruvate, w: 2.2g/L NaHCO<sub>3</sub>**Medium supplements** Supplement the medium with 10% FBS, 2 ng/mL bFGF**Passaging solution** Trypsin-EDTA**Subculturing** Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.**Seeding density** 1 to 3 x 10<sup>4</sup> cells/cm<sup>2</sup>**Fluid renewal** First fluid renewal after 24 hours, then every 2 to 3 days.

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### Freeze medium

CM-1 (Cytion catalog number 800100)

### Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

## Quality control / Genetic profile / HLA

### Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.