

MARC-145 Cells | 305006**General information****Description**

The MARC-145 cell line is a monkey kidney epithelial cell line derived from the African green monkey (*Cercopithecus aethiops*). This cell line is particularly notable for its use in virology, especially in the propagation of porcine reproductive and respiratory syndrome virus (PRRSV), a significant pathogen in the swine industry. MARC-145 cells have been instrumental in studying PRRSV due to their high susceptibility to the virus, making them a valuable tool for viral isolation, propagation, and vaccine development.

MARC-145 cells are characterized by their epithelial morphology and robust growth in vitro, which facilitates large-scale production of PRRSV for research purposes. Additionally, they have been used to explore the mechanisms of PRRSV infection, including virus entry, replication, and host-pathogen interactions. This cell line has also been utilized in the development and testing of antiviral compounds and PRRSV vaccines, contributing significantly to efforts aimed at controlling this economically important disease in pigs.

Organism

Chlorocebus pygerythrus (Vervet monkey)

Tissue

Embryonic kidney

Synonyms

Marc-145, MARC 145, Marc 145, MARC145, Marc145, Meat Animal Research Center-145

Characteristics**Age**

Foetus

Morphology

Epithelial

Growth properties

Adherent

Identifiers / Biosafety / Citation**Citation**

MARC-145 (Cytion catalog number 305006)

Biosafety level

1

Expression / Mutation**Handling****Culture Medium**

DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 1.5 g/L NaHCO₃, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)

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Medium supplements	Supplement the medium with 10% FBS
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Passaging solution	Accutase
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Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
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Split ratio	1:2 to 1:4
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Fluid renewal	2 to 3 times per week
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Freeze medium	CM-1 (Cytion catalog number 800100)
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Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.