Product sheet

Vero E6 Cells | 305008

General information

Description	Vero E6 cells, also known as Vero C1008 or Vero 76 clone E6, are a continuous line of epithelial cells derived from the kidney of the African green monkey, Chlorocebus sabaeus. The Vero clone E6, a subline of Vero cells, is particularly noted for its utility in virology research due to its high susceptibility to a wide range of viruses, including coronaviruses like SARS-CoV and SARS-CoV-2, the Ebola virus, and the Zika virus. The cell line is crucial in the production of vaccines, such as those for Japanese Encephalitis vaccine, due to their capacity for virus culture and isolation. The cells have played a pivotal role in the development of COVID therapeutics, including the testing of the polymerase inhibitor remdesivir. With their ability to support the replication of a variety of viruses, Vero E6 cells facilitate compound screening and the evaluation of antiviral efficacy. Their role in clinical trials extends to the assessment of anti-inflammatory drugs like dexamethasone and the study of gene products like the P-glycoprotein (pgp protein) encoded by the pgp gene. Vero E6 cells lack the interferon-β gene, which partly explains their high susceptibility to viral infections; this deficiency prevents them from mounting an effective innate antiviral response. In summary, Vero E6 cells are a valuable resource in the field of virology and biomedicine, providing a versatile platform for antiviral screening, the study of replication in Vero, and aiding in the quest for understanding retroviral sequences.
Organism	Chlorocebus sabaeus (Green monkey)
Tissue	Normal Kidney

Characteristics

Age	Adult
Morphology	Epithelial
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	Vero E6 (Cytion catalog number 305008)
Biosafety level	1

Expression / Mutation

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Handling

Culture Medium	EMEM, w: 2 mM L-Glutamine, w: 1.5 g/L NaHCO3, w: EBSS, w: 1 mM Sodium pyruvate, w: NEAA (Cytion article number 820100c)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Doubling time	22 hours
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	1: 2 to 1: 4
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100)

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Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.