

DF-1 Cells | 305016

General information

Description

DF-1 cells are a continuous cell line derived from chicken embryonic fibroblasts, specifically from East Lansing Line (ELL-0) chickens. The cell line is noted for its lack of endogenous avian leukosis virus, which is a common contaminant in many other chicken cell lines. This characteristic makes DF-1 cells particularly valuable for virology research, especially in studies involving the propagation and genetic manipulation of viruses that infect birds, such as avian influenza and Marek's disease virus.

In addition to their use in virology, DF-1 cells are utilized in various areas of cellular and molecular biology research. They have a robust growth rate and a fibroblast-like morphology, making them suitable for in vitro experiments that require a stable avian cell environment. These cells have been instrumental in gene expression studies, especially concerning the effects of viral and other genetic elements in avian species. The genetic stability and susceptibility to transfection also make DF-1 an excellent model for studying gene function and regulation in a controlled environment.

Organism

Chicken

Tissue

Embryo

Synonyms

DF1, UMNSAH-DF-1, UMNSAH-DF 1, UMNSAH-DF1, UMNSAH/DF1, UMNSAH/DF#1, Douglas Foster-1, UMNSAH/DF-1

Characteristics

Age

10 days gestation

Morphology

Fibroblast

Growth properties

Adherent

Identifiers / Biosafety / Citation

Citation

UMNSAH/DF-1 (Cytion catalog number 305016)

Biosafety level

1

Expression / Mutation

Tumorigenic

No

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Handling

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 1.5 g/L NaHCO ₃ , w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
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Medium supplements	Supplement the medium with 10% FBS
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Passaging solution	Accutase
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Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
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Split ratio	1:2 to 1:4
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Fluid renewal	2 to 3 times per week
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Freeze medium	CM-1 (Cytion catalog number 800100)
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Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.