Product sheet

A549/DDP Cells | 305047

General information

Description	The A549/DDP cell line is a drug-resistant variant of the A549 cell line, which itself is a model of human alveolar basal epithelial adenocarcinoma. This variant has been specifically selected for its resistance to cisplatin (DDP), a common chemotherapy drug used in the treatment of various cancers, including lung cancer. The development of the A549/DDP cell line enables researchers to study the mechanisms underlying chemoresistance, which is a major challenge in cancer therapy. In research, the A549/DDP cell line is utilized to investigate the biochemical pathways involved in cisplatin resistance. This includes the exploration of changes in gene expression, protein function, and cellular metabolism that confer resistance to cisplatin. The cell line is also valuable in the screening of new drugs or drug combinations that can overcome resistance, providing insights that are crucial for the development of more effective therapeutic strategies against lung cancer. Moreover, studies using the A549/DDP cell line contribute to a better understanding of the molecular basis of lung cancer progression and metastasis in the context of chemoresistance. This cell line serves as a critical tool for translational research, bridging experimental findings to potential clinical applications in oncology.
Organism	Human
Tissue	Lung

Characteristics

Morphology	Epithelial
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	A549/DDP (Cytion catalog number 305047)
Biosafety level	1

Expression / Mutation

Handling

Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)

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Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	1:2 to 1:4
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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STR profile

Amelogenin: x,y
CSF1PO: 10,12
D13S317: 11
D16S539: 11,12
D5S818: 11
D7S820: 8,11
TH01: 8,9.3
TPOX: 8,11
vWA: 14
D3S1358: 16
D21S11: 29
D18S51: 14,17
Penta E: 7,11
Penta D: 9
D8S1179: 13,14
FGA: 23
D6S1043: 11,13
D2S1338: 24
D12S391: 18
D19S433: 13