

Hela 229 Cells | 305056**General information****Description**

The HeLa 229 cell line is a clonal derivative of the original HeLa cell line, which was the first human cell line to be continuously cultured. HeLa cells were derived from cervical cancer cells taken from Henrietta Lacks in 1951. The HeLa 229 subline is utilized in various areas of biomedical research, including cancer research, drug development, and toxicology, due to its robust growth and adaptability under laboratory conditions.

One of the main characteristics of the HeLa 229 cell line is its aggressive growth and proliferation, reflecting the cancerous origin of the cells. This makes it particularly useful for studies requiring high cell yields and rapid growth, such as high-throughput screening for drug discovery. HeLa 229 cells are also highly amenable to genetic manipulation, allowing researchers to introduce foreign genes or specific mutations to study their effects on cell behavior and pathology.

HeLa 229 cells continue to be a critical model in virology, as they are susceptible to a wide variety of viruses. This susceptibility makes them an excellent tool for studying viral life cycles, host-virus interactions, and the efficacy of antiviral compounds. The cell line has also been instrumental in advancing our understanding of fundamental cellular processes, such as DNA replication, transcription, and apoptosis.

Despite their utility, the use of HeLa cells, including HeLa 229, raises ethical considerations regarding consent and the origins of the cell line, as the cells were originally obtained without the consent of Henrietta Lacks or her family. However, ongoing research with HeLa cells continues to contribute significantly to science, driven by their unique characteristics and historical importance in the development of modern cell biology.

Organism

Human

Tissue

Cervix

Disease

Human papillomavirus-related endocervical adenocarcinoma

Synonyms

HeLa-229, HeLa229

Characteristics**Age**

31 years

Gender

Female

Morphology

Epithelial

Growth properties

Adherent

Identifiers / Biosafety / Citation

Hela 229 Cells | 305056

Citation	Hela 229 (Cytion catalog number 305056)
-----------------	---

Biosafety level	1
------------------------	---

Expression / Mutation**Handling**

Culture Medium	EMEM, w: 2 mM L-Glutamine, w: 1.5 g/L NaHCO ₃ , w: EBSS, w: 1 mM Sodium pyruvate, w: NEAA (Cytion article number 820100c)
-----------------------	--

Medium supplements	Supplement the medium with 10% FBS
---------------------------	------------------------------------

Passaging solution	Accutase
---------------------------	----------

Doubling time	26 hours
----------------------	----------

Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
---------------------	---

Split ratio	1:2 to 1:5
--------------------	------------

Fluid renewal	2 to 3 times per week
----------------------	-----------------------

Freeze medium	CM-1 (Cytion catalog number 800100)
----------------------	-------------------------------------

Hela 229 Cells | 305056

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

Hela 229 Cells | 305056

STR profile

Amelogenin: x,x
CSF1PO: 9,1
D13S317: 12,13.3
D16S539: 9,1
D5S818: 11,12
D7S820: 8,12
TH01: 7
TPOX: 8,12
vWA: 16,18
D3S1358: 15,18
D21S11: 27,28
D18S51: 16
Penta E: 7,17
Penta D: 8,15
D8S1179: 12,13
FGA: 18,21
D6S1043: 18
D2S1338: 17
D12S391: 20,25
D19S433: 13,14