Product sheet

NCI-H1650 Cells | 305059

General information

Description	The NCI-H1650 cell line is derived from a human non-small cell lung carcinoma (NSCLC), specifically adenocarcinoma, and is widely used in cancer research due to its distinctive genetic profile and relevance in drug testing. This cell line features mutations in key oncogenic and tumor suppressor pathways, including a deletion in the PTEN gene and an activating mutation in EGFR. These genetic alterations make NCI-H1650 a suitable model for studying mechanisms of tumorigenesis and therapeutic resistance in NSCLC, especially in the context of targeted therapies aimed at the EGFR signaling pathway. The deletion of PTEN in NCI-H1650 results in the loss of phosphatase activity, which deregulates the PI3K/AKT signaling pathway, contributing to tumor progression and resistance to certain therapeutic agents. The activating EGFR mutation, commonly observed in lung adenocarcinoma, renders the cell line particularly sensitive to tyrosine kinase inhibitors like erlotinib. However, the co-occurrence of these genetic changes often necessitates combination therapies to overcome adaptive resistance mechanisms that involve compensatory signaling pathways, such as mTOR or MET. In addition to its genetic and signaling characteristics, NCI-H1650 has been included in numerous studies examining somatic mutations, copy number variations, and epigenetic alterations in cancer cell lines. Its response to inhibitors of EGFR and PI3K pathways highlights its utility in preclinical drug discovery and personalized medicine strategies. This cell line serves as a representative model for investigating the interplay between oncogenic drivers and therapeutic vulnerabilities in lung adenocarcinoma.
Organism	Human
Tissue	Lung
Disease	Minimally invasive lung adenocarcinoma
Metastatic site	Pleural effusion
Synonyms	NCI-H1650, H-1650, H1650_CO, NCIH1650

Characteristics

Age	27 years
Gender	Male
Ethnicity	European
Morphology	Epithelial

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Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	NCI-H1650 (Cytion catalog number 305059)
Biosafety level	1

Expression / Mutation

Handling

Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	1:2 to 1:4
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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STR profile

Amelogenin: x,x
CSF1PO: 11
D13S317: 11
D16S539: 11,12
D5S818: 11
D7S820: 8,9
TH01: 09. Mrz
TPOX: 11
vWA: 18
D3S1358: 18
D21S11: 30
D18S51: 10
Penta E: 12
Penta D: 8
D8S1179: 12
FGA: 20
D6S1043: 13
D2S1338: 19
D12S391: 22
D19S433: 15