Product sheet

SNU-1 Cells | 305076

General information

Description	The SNU-1 cell line is derived from the gastric carcinoma of a human adult and is widely utilized in gastric cancer research. This cell line provides an important model for studying the molecular and cellular mechanisms underlying gastric adenocarcinoma, a common and often deadly form of stomach cancer. SNU-1 cells are particularly valuable for investigating the genetic alterations and signaling pathways involved in the pathogenesis of gastric cancer, as well as for developing and testing new therapeutic strategies. SNU-1 cells exhibit an epithelial morphology and are characterized by the expression of markers typical of gastric epithelial cells and adenocarcinoma, such as carcinoembryonic antigen (CEA) and cytokeratins. They are often used in studies exploring the role of oncogenes, tumor suppressor genes, and other molecular factors in gastric cancer progression. Researchers employ SNU-1 cells to assess the efficacy and mechanisms of action of chemotherapeutic agents, targeted therapies, and combination treatments. Additionally, SNU-1 cells serve as a model for understanding the tumor microenvironment and the interactions between cancer cells and stromal cells. The relevance of the SNU-1 cell line in gastric cancer research highlights its importance in advancing our knowledge of this malignancy and in the development of effective treatments for gastric cancer patients.
Organism	Human
Tissue	Stomach
Disease	Adenocarcinoma
Synonyms	SNU1, NCI-SNU-1

Characteristics

Age	44 years
Gender	Male
Ethnicity	Asian
Morphology	Epithelial
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	SNU-1 (Cytion catalog number 305076)

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Biosafety level	1

Expression / Mutation

Receptors expressed	Vasoactive intestinal peptide(VIP), expressed
Antigen expression	Blood Type O, Rh?, The cells express the surface glycoproteins carcinoembryonic antigen(CEA) and TAG 72.

Handling

Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Medium supplements	Supplement the medium with 10% heat-inactivated FBS
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	1:2 to 1:4
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.