Product sheet

OS-RC-2 Cells | 305086

General information

Description	The OS-RC-2 cell line is a human renal cell carcinoma (RCC) model established from the tumor of a Japanese male patient diagnosed with clear cell RCC. This cell line exhibits hallmark features of RCC, including the presence of numerous long microvilli on its surface and glycogen granules within its cytoplasm, as observed through electron microscopy. These characteristics align closely with the features of proximal tubular epithelial cells, thought to be the origin of clear cell RCC. OS-RC-2 has proven to be tumorigenic in immunocompromised mice, where the histopathological features of xenograft tumors strongly resemble the original patient tumor. Chromosomal analyses of OS-RC-2 reveal a hypodiploid modal number of 40, with evidence of a marker chromosome and a specific translocation between chromosomes 2 and 13. Additionally, a large subset of the cell population exhibits a hypotetraploid karyotype with a modal number of 75. These genetic features make OS-RC-2 a valuable model for studying chromosomal aberrations and tumor biology in RCC. Further research using OS-RC-2 has shed light on the role of cytokines in RCC, including tumor necrosis factor-alpha (TNF-?) and interleukin-6 (IL-6). Studies have demonstrated that while TNF-? does not induce DNA synthesis or cell proliferation in OS-RC-2, it can stimulate IL-6 production at high concentrations. These findings contribute to understanding the complex interplay of cytokines in RCC progression and the tumor microenvironment, making OS-RC-2 a useful tool for investigating therapeutic interventions in RCC.
Organism	Human
Tissue	Kidney
Disease	Clear cell renal cell carcinoma
Synonyms	OSRC2, RC-2

Characteristics

Age	52 years
Gender	Male
Ethnicity	Asian
Morphology	Epithelial
Growth properties	Adherent

Identifiers / Biosafety / Citation

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Citation	OS-RC-2 (Cytion catalog number 305086)
Biosafety level	1

Expression / Mutation

Tumorigenic	Yes

Handling

Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	1:2 to 1:4
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.