Product sheet

SW780 Cells | 305098

General information

Description	The patient had preoperative chemotherapy(Thiotepa).The cells have a reported plating efficiency of 41%.
Organism	Human
Tissue	Urinary bladder
Disease	Bladder carcinoma
Synonyms	SW-780, SW 780

Characteristics

Age	80 years
Gender	Female
Ethnicity	European
Morphology	Epithelial
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	SW780 (Cytion catalog number 305098)
Biosafety level	1

Expression / Mutation

Tumorigenic	Yes

Handling

Culture Medium	Leibovitz's L-15, w: 2.0 mM L-Glutamine, 0.55 g/L NaHCO3 (We do not supply this product; please consider other suppliers. Please let us know if you need further assistance)

SW780 Cells | 305098

Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	1:2 to 1:4
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

SW780 Cells | 305098

STR profile

Amelogenin: x,x
CSF1PO: 10,11
D13S317: 11,12
D16S539: 9,11
D5S818: 11,12
D7S820: 9,1
TH01: 6
TPOX: 8
vWA: 16,19
D3S1358: 16,18
D21S11: 29,3
D18S51: 15
Penta E: 12
Penta D: 9
D8S1179: 13
FGA: 19,2
D6S1043: 11,2
D2S1338: 19,22
D12S391: 20,25
D19S433: 13,15