Product sheet

SU-DHL-4 Cells | 305106

General information

Description	The SU-DHL-4 cell line is derived from a lymphoblast-like cell isolated from the peritoneal effusion of a 38-year-old Caucasian male patient. This cell line represents a model of diffuse large B-cell lymphoma (DLBCL), one of the most common types of non-Hodgkin lymphoma in adults. The establishment of this cell line has provided valuable insights into the biology of DLBCL, especially concerning the cellular and molecular mechanisms underlying lymphomagenesis and tumor progression. In research, SU-DHL-4 cells have been extensively utilized to study the efficacy and mechanism of action of various chemotherapeutic and targeted therapeutic agents, reflecting their importance in lymphoma treatment research. The cells express several key immunophenotypic markers associated with B-cell lineage such as CD19 and CD20, which are crucial for the development and function of B-lymphocytes. These markers also make SU-DHL-4 an excellent target for testing B-cell-specific therapies, including monoclonal antibodies and small molecule inhibitors that disrupt critical signaling pathways involved in lymphoma cell survival and proliferation.
Organism	Human
Tissue	Peritoneal effusion
Disease	Diffuse large B-cell lymphoma
Synonyms	SUDHL4, Sudhl4, SUDHL-4, Sudhl-4, SuDHL 4, SUD-4, SUD4, SU4, Stanford University-Diffuse Histiocytic Lymphoma-4, DHL-4, DHL4

Characteristics

Age	38 years
Gender	Male
Ethnicity	European
Morphology	Lymphoblast
Growth properties	Suspension

Identifiers / Biosafety / Citation

Citation	SU-DHL-4 (Cytion catalog number 305106)
Biosafety level	1

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Expression / Mutation

Protein expression	IgG+, Kappa+, IgM-, IgA-, IgD-, Lambda-, This cell line has relatively high expression levels of Bax, Bak, AIF, high caspase-9 activity.

Handling

Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Medium supplements	Supplement the medium with 10% FBS
Doubling time	40 hours
Subculturing	Maintain cultures by periodically adding or replacing the medium. Initiate cultures with a density of 2 x 10^5 cells/ml and keep the cell concentration within the range of 1 x 10^5 to 1 x 10^6 cells/ml for optimal growth.
Split ratio	1:2 to 1:6
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.