Product sheet

CAL-62 Cells | 305114

General information

Description	The CAL-62 cell line was established from the right lobe of the thyroid gland of a 70-year-old Caucasian woman in 1988 and has been extensively used in the study of thyroid anaplastic carcinoma. These human epithelial-like cells exhibit a distinctive monolayer growth pattern and demonstrate pronounced tumorigenic properties, making them a significant model for in vivo studies of thyroid cancer progression. When transplanted into immunodeficient nude mice, CAL-62 cells have shown a robust capability to form tumors, providing a practical and effective model to analyze tumor dynamics and evaluate potential therapeutic strategies in real-time biological settings. Characterized by a rapid proliferation rate with a doubling time of approximately 24 hours, CAL-62 enables accelerated research outputs in studies that are time-sensitive, enhancing the efficiency of experimental workflows in cancer research. Genetic characterization of this cell line reveals the presence of the KRAS p.G12R mutation and alterations at the 9p21.3 locus, indicating complex genetic underpinnings associated with thyroid anaplastic carcinoma. This cell line?s stable epithelial phenotype and inherent radioresistance further underscore its utility in uncovering novel insights into the pathophysiology of aggressive thyroid cancers and in the development of new therapeutic modalities. The unique attributes of CAL-62, including its aggressive tumor-forming ability and genetic markers, make it a pivotal resource in the ongoing efforts to better understand and treat thyroid anaplastic carcinoma.
Organism	Human
Tissue	Thyroid
Disease	Thyroid gland anaplastic carcinoma
Synonyms	Cal-62, CAL 62, Cal 62, CAL62, Centre Antoine Lacassagne-62

Characteristics

Age	70 years
Gender	Female
Ethnicity	European
Morphology	Epithelial
Growth properties	Adherent

Identifiers / Biosafety / Citation

CAL-62 Cells | 305114

Citation	CAL-62 (Cytion catalog number 305114)
Biosafety level	1

Expression / Mutation

Handling

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 1.5 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Doubling time	24 hours
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	1:2 to 1:5
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.