Product sheet

KHM-5M Cells | 305148

General information

Description	The KHM-5M cell line is an important model derived from a patient with undifferentiated thyroid carcinoma complicated by neutrophilia and malignant pleurisy. This cell line is characterized by its significant production of neutrophil chemotactic factors, specifically human interleukin 8 (IL-8) and granulocyte-macrophage colony-stimulating factor (GM-CSF). These factors are crucial in the recruitment and activation of neutrophils, which play a pivotal role in the immune response and inflammation. The KHM-5M cells were shown to possess extreme chemotactic activity, a trait that was substantiated through in vitro experiments using conditioned media from the cells and the modified Boyden chamber technique. Additionally, KHM-5M cells were transplanted into nude rats, where the infiltration of neutrophils was observed in and around the transplanted tumor tissue. This finding underscores the relevance of KHM-5M as a model for studying the interactions between tumor cells and the immune microenvironment, particularly in relation to neutrophil recruitment and function. The cell line also serves as a valuable tool for investigating the molecular mechanisms underlying cytokine production in cancer and the subsequent modification of pathological features. Through DNA cloning techniques, the chemotactic activities attributed to IL-8 and GM-CSF were confirmed, solidifying the KHM-5M cell line as a significant resource for research into cytokine-driven tumor-immune interactions.
Organism	Human
Tissue	Thyroid
Disease	Thyroid gland anaplastic carcinoma
Metastatic site	Pleural effusion
Synonyms	KHM/5M, KHM5M

Characteristics

Age	65 years
Gender	Male
Morphology	Fibroblast
Growth properties	Adherent

Identifiers / Biosafety / Citation

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Citation	KHM-5M (Cytion catalog number 305148)
Biosafety level	1

Expression / Mutation

Handling

Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Doubling time	27 hours
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	1:2 to 1:5
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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STR profile

Amelogenin: x,x
CSF1PO: 12,13
D13S317: 8,11
D16S539: 10
D5S818: 12
D7S820: 10,11
TH01: 7
TPOX: 8
vWA: 18
D3S1358: 15
D21S11: 28,31
D18S51: 16,19
Penta E: 11,18
Penta D: 9,11
D8S1179: 13
FGA: 22,23
D6S1043: 13,19
D2S1338: 19,23
D12S391: 18,21
D19S433: 14