Product sheet

C3H/10T1/2 Cells | 305164

General information

Description	The C3H/10T1/2, Clone 8 cell line is a murine fibroblast cell line derived from C3H mouse embryo tissues. This cell line is widely utilized in biological research due to its capacity to differentiate into a variety of cell types when treated with appropriate agents. The C3H/10T1/2 cells exhibit characteristics typical of fibroblasts but have the remarkable ability to undergo transformation into adipocytes, chondrocytes, or osteoblasts under specific experimental conditions. This makes them an invaluable model for studying mesenchymal differentiation, tissue engineering, and carcinogenesis. These cells are particularly noted for their use in research involving the mechanisms of action of carcinogens and the genetic regulation of cellular transformation. C3H/10T1/2, Clone 8 cells are sensitive to contact inhibition and maintain a stable phenotype under standard culture conditions, which is critical for reproducible results in experiments. Furthermore, their responsiveness to a variety of chemical and environmental stimuli makes them an excellent model for toxicology studies, examining the effects of various substances on cellular behavior and differentiation pathways.
Organism	Mouse
Tissue	Embryo
Synonyms	C3H/10T1/2 Clone 8, C3H/10T1/2-clone8, C3H/10T1/2 CL8, C3H10T1/2 clone8, C3H10T1/2CL8, 10T1/2(clone8), 10T1/2, C3H10T1-2, C3H10T1/2, C3H-10T1/2, C3H 10T1/2, C3H/10T1/2

Characteristics

Age	Embryo
Morphology	Fibroblast
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	C3H/10T1/2, Clone 8 (Cytion catalog number 305164)
Biosafety level	1

Expression / Mutation

Tumorigenic	No

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Handling

Culture Medium	BME, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 1.5 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (We do not supply BME; please consider other suppliers. Please let us know if you need further assistance)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	1:2 to 1:4
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.