Product sheet

MC3T3-E1 Subclone 24 Cells | 305186

General information

Description	MC3T3-E1 Subclone 24 cells expressly represent a preosteoblast cell type, which plays a crucial role in bone formation. Morphologically, they exhibit a fibroblast-like appearance, characterized by their elongated shape and spindle-like structures. This particular subclone is derived from the calvaria tissue, a skull region contributing to bone formation. One of the critical applications of MC3T3-E1 Subclone 24 Cells lies in 3D cell culture, where researchers can study the behavior and interactions of these cells within a three-dimensional environment. This method offers a more physiologically relevant model than traditional two-dimensional cell cultures, allowing for a better understanding of the intricate processes involved in bone formation. While these cells possess numerous advantages, it's important to note their specific characteristics. MC3T3-E1 Subclone 24 Cells have been observed to exhibit poor osteoblast differentiation when exposed to ascorbic acid, a key component for promoting bone cell growth. Furthermore, they do not form a mineralized extracellular matrix, a crucial step in creating bone tissue. The doubling time of MC3T3-E1 Subclone 24 Cells is approximately 90.5 hours.
Organism	Mouse
Tissue	Bone
Applications	3D cell culture, Differentiation studies

Characteristics

Age	1 day
Gender	Unspecified
Morphology	Fibroblast
Cell type	Osteoblast
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	MC3T3-E1 Subclone 24 (Cytion catalog number 300640)
Biosafety level	1

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Expression / Mutation

Receptors expressed	parathyroid hormone-related protein (PTHrP) receptor
Protein expression	Collagen, bone sialoprotein (BSP), osteocalcin (OCN), parathyroid hormone (PTH)
Tumorigenic	Yes, in immunosuppressed mice

Handling

Culture Medium	Alpha MEM, w: 2.0 mM stable Glutamine, w: Ribonucleosides, w: Deoxyribonucleosides, w: 1.0 mM Sodium pyruvate, w: 2.2g/L NaHCO3, w/o: Ascorbic acid (GIBCO, Catalog No. A1049001. We do not supply this product; please consider other suppliers. Please let us know if you need further assistance.)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Freeze medium	CM-1 (Cytion catalog number 800100)

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Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.