#### **Product sheet**



## Mv.1.Lu Cells | 305192

### **General information**

### **Description**

The Mv.1.Lu cell line, also known as CCL 64, originates from the lung tissue of a fetal mink (Mustela vison). It is an epithelial-like cell line known for its flat, contact-inhibited growth in monolayer cultures, exhibiting a regular polygonal morphology. This cell line has been widely utilized in virological studies due to its broad permissivity for various mammalian Type C viruses, including murine and feline sarcoma viruses, making it a preferred system for focus formation assays and viral transformation studies.

This cell line's ability to support viral replication and transformation has made it an essential tool in understanding viral pathogenesis and host-pathogen interactions. Mv.1.Lu cells are also used in pulmonary physiology research, benefiting from their origin in lung tissue. Studies on their growth characteristics, including their response to various media and culture conditions, have underscored their adaptability and stability in laboratory settings.

Organism Neovison vison (American mink)

Tissue Lung

**Synonyms** Mv 1 Lu (NBL-7), NBL-7, Mv 1 Lu, MV 1 LU, Mv1.Lu, Mv.1.Lu, MV-1-Lu, Mink, Mink Lung

## **Characteristics**

**Age** Near term fetus

Gender Mixed sex

Morphology Epithelial

Growth properties

Adherent

### **Identifiers / Biosafety / Citation**

**Citation** Mv.1.Lu (Cytion catalog number 305192)

**Biosafety level** 2

### **Expression / Mutation**

## **Handling**

## **Product sheet**



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| Culture<br>Medium     | EMEM, w: 2 mM L-Glutamine, w: 1.5 g/L NaHCO3, w: EBSS, w: 1 mM Sodium pyruvate, w: NEAA (Cytion article number 820100c)   |
|-----------------------|---|
| Medium<br>supplements | Supplement the medium with 10% FBS  |
| Passaging solution    | Accutase  |
| Subculturing          | Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium. |
| Split ratio           | 1:2 to 1:4  |
| Fluid renewal         | 2 to 3 times per week   |
| Freeze<br>medium      | CM-1 (Cytion catalog number 800100)   |

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### Handling of cryopreserved cultures

- 1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
- 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
- 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
- 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
- 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
- 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
- 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
- 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

## Quality control / Genetic profile / HLA

### **Sterility**

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.