Product sheet

RH-35 Cells | 305210

General information

Description	The H4-II-E (also referred to as RH-35) cell line is a derivative of the Reuber H-35 rat hepatoma. This cell line originated from a liver tumor induced in a male ACI rat by exposure to the chemical carcinogen N-2-fluorenyldiacetamide. When transplanted into ACI rats, H4-II-E cells form rapidly growing tumors with histological features characteristic of poorly differentiated hepatomas. They are particularly sensitive to the induction of aryl hydrocarbon hydroxylase (AHH) activity, making them a robust system for studying enzymatic responses to polycyclic aromatic hydrocarbons and dioxin-like compounds. H4-II-E cells also serve as a model for studying cellular responses to carcinogens and radiation, given their clonogenicity and the ability to assay long-term cell survival post-treatment. Their application extends to exploring the mechanisms of enzyme induction, xenobiotic metabolism, and liver-specific toxicology. These attributes make H4-II-E an invaluable tool in cancer research and toxicological screening.
Organism	Rat
Tissue	Liver
Disease	Rat hepatocellular carcinoma
Synonyms	H4II, H-35tc2, Reuber-H-35 hepatoma tissue culture 2, Reuber H-35 tc2, Reuber H35 tc2, H-35 Reuber tc2, H35 Reuber tc2, RH-35 tc2, RH35 tc2, H-35 tc2, H35 tc2

Characteristics

Gender	Male
Morphology	Epithelial
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	RH-35 (Cytion catalog number 305210)
Biosafety level	1

Expression / Mutation

Handling

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Culture Medium	Ham's F12, w: 1.0 mM stable Glutamine, w: 1.0 mM Sodium pyruvate, w: 1.1 g/L NaHCO3 (Cytion article number 820600a)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	1:2 to 1:4
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.