

# Human CEACAM6 ELISA Pair Set

( CD66c )

Catalog Number: SEK10823

To achieve the best assay results, this manual must be read carefully before using this product and the assay is run as summarized in the General ELISA protocol.

#### **BACKGROUND**

Carcinoembryonic antigen-related cell adhesion molecule 6 ( CEACAM6 ), also known as nonspecific crossreacting antigen (NCA) and CD66c, is one of seven human CEACAM family members within the immunoglobulin superfamily. CEACAM family members are a set of widely expressed proteins involved in several biological functions, including cell adhesion, migration, signal transduction, and the regulation of gene expression. Abnormal overexpression and downregulation of some CEACAMs have been described in tumor cells. Monoclonal antibodies grouped in the CD66 cluster recognize CEACAM members. Ectopic CD66 expression is commonly detected in B-cell lineage acute lymphoblastic leukemia (ALL). Human CEACAMs include type I transmembrane proteins (CEACAM-1, -3, and -4) and GPI-linked molecules (CEACAM- 5 through -8). There is no human CEACAM-2. Human CEACAM6 is a GPI-linked membrane protein consisting of a signal sequence, an extracellular domain (ECD) containing one N-terminal V-type Ig-like domain and two C2-type Iglike domains, and a hydrophobic C-terminal propeptide. The GPI membrane anchor is attached at the C-terminus following cleavage of the propeptide. CEACAM-6 is expressed by granulocytes and their progenitors. It is also expressed by epithelia of various organs and is upregulated in pancreatic and colon adenocarcinomas, as well as hyperplastic polyps. Resistance to adhesion-related apoptosis in tumor cells is conferred in the condition of CEACAM6 overexpression. CEACAM6 is emerging as an important determinant of the malignant phenotype in a range of cancers. CEACAM6 plays a key role in pancreatic adenocarcinoma cellular invasiveness. Levels of CEACAM6 expression can modulate pancreatic adenocarcinoma cellular invasiveness in a c-Src-dependent manner. This pathway warrants further investigation as a target for therapy.

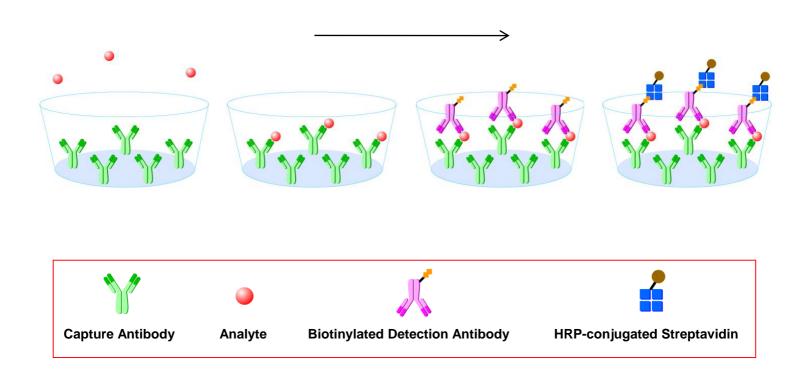
#### PRINCIPLE OF THE TEST

The Sino Biological ELISA Pair Set is a solid phase sandwich ELISA (Enzyme-Linked Immunosorbent Assay). It utilizes a monoclonal antibody specific for CEACAM6 / CD66c coated on a 96-well plate. Standards and samples are added to the wells, and any CEACAM6 / CD66c present binds to the immobilized antibody. The wells are washed and a biotinylated rabbit anti- CEACAM6 / CD66c polyclonal antibody is then added, producing an antibody-antigen-antibody "sandwich". To produces color in proportion to the amount of CEACAM6 / CD66c present in the sample strepavidin-HRP and TMB substrate solution are loaded. The absorbances of the microwell are read at 450 nm.

#### **INTENDED USE**

- The human CEACAM6 / CD66c ELISA Pair Set is for the quantitative determination of human CEACAM6 / CD66c.
- This ELISA Pair Set contains the basic components required for the development of sandwich ELISAs.

#### ASSAY PROCEDURE SUMMARY



This Pair Set has been configured for research use only and is not to be used in diagnostic procedures.

## MATERIALS PROVIDED

Bring all reagents to room temperature before use.

Capture Antibody - 1 mg/mL of mouse anti-CEACAM6 monoclonal antibody. Dilute to a working concentration of 2  $\mu$ g/mL in PBS before coating.

**Detection Antibody** - 0.4 mg/mL of biotinylated rabbit anti-CEACAM6 polyclonal antibody. Dilute to working concentration of 0.5 μg/mL in detection antibody diluteion buffer before use.

**Standard** - Each vial contains 88 ng of recombinant CEACAM6. Reconstitute standard powder with 1 mL detection antibody dilution buffer. A seven-point standard curve using 2-fold serial dilutions in sample dilution buffer, and a high standard of 1000 pg/mL is recommended.

**Streptavidin-HRP** - 50  $\mu$ L of streptavidin conjugated to horseradish-peroxidase. 1:2000 Dilution in detection antibody dilution buffer before use.

#### **SOLUTIONS REQUIRED**

PBS - 136.9 mM NaCl, 10.1 mM Na<sub>2</sub>HPO<sub>4</sub>, 2.7 mM KCl, 1.8 mM KH<sub>2</sub>PO<sub>4</sub>, pH 7.4, 0.2 μm filtered

TBS - 25mM Tris, adjust pH to 7.4 by HCl

Wash Buffer - 0.05% Tween20 in TBS, pH 7.2 - 7.4

Blocking Buffer - 2% BSA in Wash Buffer

Sample dilution buffer - 0.1% BSA in wash buffer, pH 7.2 - 7.4, 0.2 µm filtered

**Detection antibody dilution buffer** - 0.5% BSA in wash buffer, pH 7.2 - 7.4, 0.2 µm filtered.

Substrate Solution: To achieve best assay results, fresh substrate solution is recommended

Substrate stock solution - 10 mg/ml TMB (Tetramethylbenzidine) in DMSO

Substrate dilution buffer - 0.05M Na<sub>2</sub>HPO<sub>4</sub> and 0.025M citric acid; adjust pH to 5.5

Substrate working solution - For each plate dilute 250 µl substrate stock solution in 25ml substrate dilution

buffer and then add  $80 \mu l 0.75\% H_2O_2$ , mix it well

Stop Solution - 2 N H<sub>2</sub>SO<sub>4</sub>

#### **PRECAUTION**

The Stop Solution suggested for use with this Pair Set is an acid solution. Wear eye, hand, face, and clothing protection when using this material.

#### **STORAGE**

**Capture Antibody**: Aliquot and store at  $-20^{\circ}$ C to  $-80^{\circ}$ C for up to 6 months from date of receipt. Avoid repeated freeze-thaw cycles.

**Detection Antibody**: Aliquot and store at -20°C to -80°C for up to 6 months from date of receipt. Avoid repeated freeze-thaw cycles.

**Standard**: Store lyophilized standard at  $-20^{\circ}$ C to  $-80^{\circ}$ C for up to 6 months from date of receipt. Aliquot and store the reconstituted standard at  $-20^{\circ}$ C to  $-80^{\circ}$ C for up to 1 month. Avoid repeated freeze-thaw cycles.

**Streptavidin-HRP**: Store at  $4^{\circ}$ C and protect it from prolonged exposure to light. **DO NOT FREEZE!** It is stable for up to 6 months from date of receipt.

#### **GENERAL ELISA PROTOCOL**

#### **Plate Preparation**

- 1. Dilute the capture antibody to the working concentration in PBS. Immediately coat a 96-well microplate with 100µL per well of the diluted capture antibody. Seal the plate and incubate overnight at 4°C.
- 2. Aspirate each well and wash with at least 300µl wash buffer, repeating the process two times for a total of three washes. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining wash buffer by inverting the plate and blotting it against clean paper towels.
- 3. Block plates by adding 300 µL of blocking buffer to each well. Incubate at room temperature for a minimum of 1 hour.
- 4. Repeat the aspiration/wash as in step 2. The plates are now ready for sample addition.

## **Assay Procedure**

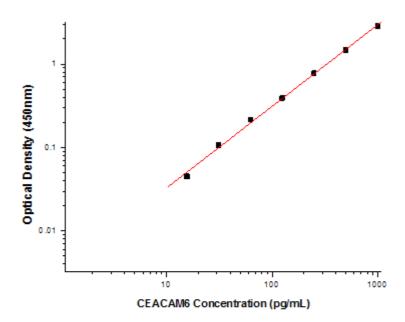
- 1. Add 100 μL of sample or standards in sample dilution buffer per well. Seal the plate and incubate 2 hours at room temperature.
- 2. Repeat the aspiration/wash as in step 2 of plate preparation.
- 3. Add 100  $\mu$ L of the detection antibody, diluted in antibody dilution buffer, to each well. Seal the plate and incubate 1 hour at room temperature.
- 4. Repeat the aspiration/wash as in step 2 of plate preparation.
- 5. Add 100 μL of Streptavidin-HRP to each well. Incubate for 1 hour at room temperature.
- 6. Repeat the aspiration/wash as in step 2 of plate preparation.
- 7. Add 200 µL of substrate solution to each well. Incubate for 20 minutes at room temperature ( if substrate solution is not as requested, the incubation time should be optimized ). Avoid placing the plate in direct light.
- 8. Add 50 µL of stop solution to each well. Gently tap the plate to ensure thorough mixing.
- 9. Determine the optical density of each well immediately, using a microplate reader set to 450 nm.

#### **CALCULATION OF RESULTS**

- Calculate the mean absorbance for each set of duplicate standards, controls and samples. Subtract the mean zero standard absorbance from each.
- Construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph.
- To determine the concentration of the unknowns, find the unknowns' mean absorbance value on the y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the x-axis and read the concentration. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.
- Alternatively, computer-based curve-fitting statistical software may also be employed to calculate the concentration of the sample.

#### **TYPICAL DATA**

This standard curve is only for demonstration purposes. A standard curve should be generated for each assay.



Concentration ( pg/mL)	Zero standard subtracted OD
0	0
15.63	0.045
31.25	0.107
62.5	0.215
125	0.392
250	0.777
500	1.476
1000	2.833

#### PERFORMANCE CHARACTERISTIC

#### **SENSITIVITY**

The minimum detectable dose of human CEACAM6 / CD66c was determined to be approximately **15.6 pg/ml**. This is defined as at least three times standard deviations above the mean optical density of 10 replicates of the zero standard.

# TROUBLE SHOOTING

Problems	Possible Sources	Solutions
No signal	Incorrect or no Detection Antibody was added	Add appropriate Detection Antibody and continue
	Substrate solution was not added	Add substrate solution and continue
	Incorrect storage condition	Check if the kit is stored at recommended condition and used before expiration date
Poor Standard Curve	Standard was incompletely reconstituted or was inappropriately stored	Aliquot reconstituted standard and store at -70 $^{\circ}\!$
	Imprecise / inaccurate pipetting	Check / calibrate pipettes
	Incubations done at inappropriate temperature, timing or agitation	Follow the general ELISA protocol
	Background wells were contaminated	Avoid cross contamination by using the sealer appropriately
Poor detection value	The concentration of antigen in samples was too low	Enriching samples to increase the concentration of antigen
	Samples were ineffective	Check if the samples are stored at cold environment. Detect samples in timely manner
High Background	Insufficient washes	Use multichannel pipettes without touching the reagents on the plate
		Increase cycles of washes and soaking time between washes
	TMB Substrate Solution was contaminated	TMB Substrate Solution should be clear and colorless prior to addition to wells
	Materials were contaminated.	Use clean plates, tubes and pipettes tips
Non-specificity	Samples were contaminated	Avoid cross contamination of samples
	The concentration of samples was too high	Try higher dilution rate of samples

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# Human CEACAM6 / CD66c ELISA Pair Set Notes