

Human Immunodeficiency Virus type 1 (HIV-1) gp120 / Glycoprotein 120 ELISA Pair Set

Catalog Number: SEK11233 Lot Number: KW15AU2602

To achieve the best assay results, this manual must be read carefully before using this product and the assay is run as summarized in the General ELISA protocol.

BACKGROUND

The HIV-1 gp120 envelope protein, a glycoprotein that is part of the outer layer of the virus, which is an essential component in the multi-tiered viral entry process. It presents itself as viral membrane spikes consisting of 3 molecules of gp120 linked together and anchored to the membrane by gp41 protein. Gp120 is essential for viral infection as it facilitates HIV entry into the host cell and this is its best-known and most researched role in HIV infection. However, it is becoming increasingly evident that gp120 might also be facilitating viral persistence and continuing HIV infection by influencing the T cell immune response to the virus. The surface protein gp120 attaches the virus to the host lymphoid cell by binding to the primary receptor CD4. Gp120 binding to its receptor CD4 and co-receptor, CXCR4 or CCR5 is required for fusion of viral and cellular membranes. Several mechanisms might be involved in this process of which gp120 binding to the CD4 receptor of T cells is the best known and most important interaction as it facilitates viral entry into the CD4+ cells and their depletion, a hallmark of the HIV infection. Gp120 is shed from the viral membrane and accumulates in lymphoid tissues in significant amounts. Despite the overall genetic heterogeneity of the gp120 glycoprotein, the conserved CD4 binding site provides an attractive antiviral target. Interaction between gp120 and ITGA4/ITGB7 would allow the virus to enter GALT early in the infection, infecting and killing most of GALT's resting CD4+ Tcells. This T-cell depletion is believed to be the major insult to the host immune system leading to AIDS.

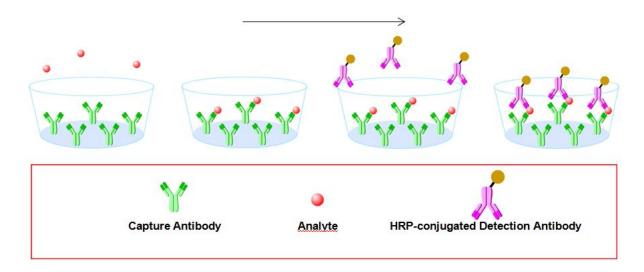
PRINCIPLE OF THE TEST

The Sino Biological ELISA Pair Set is a solid phase sandwich ELISA (Enzyme-Linked Immunosorbent Assay). It utilizes a monoclonal antibody specific for HIV-1 gp120 / Glycoprotein 120 coated on a 96-well plate. Standards and samples are added to the wells, and any HIV-1 gp120 / Glycoprotein 120 present binds to the immobilized antibody. The wells are washed and a horseradish peroxidase conjugated rabbit anti-HIV-1 gp120 / Glycoprotein 120 monoclonal antibody is then added, producing an antibody-antigenantibody "sandwich". The wells are again washed and TMB substrate solution is loaded, which produces color in proportion to the amount of HIV-1 gp120 / Glycoprotein 120 present in the sample. To end the enzyme reaction, the stop solution is added and absorbances of the microwell are read at 450 nm.

INTENDED USE

- ◆The HIV-1 gp120 / Glycoprotein 120 ELISA Pair Set is for the quantitative determination of HIV-1 gp120 / Glycoprotein 120.
- ◆This ELISA Pair Set contains the basic components required for the development of sandwich ELISAs.

ASSAY PROCEDURE SUMMARY



This Pair Set has been configured for research use only and is not to be used in diagnostic procedures.

MATERIALS PROVIDED

Bring all reagents to room temperature before use.

Capture Antibody – 1 mg/mL of rabbit anti-HIV-1 gp120 / Glycoprotein 120 monoclonal antibody (in PBS, pH 7.4). Dilute to a working concentration of 1 μ g/mL in PBS before coating. (Catalog: # 11233-R001)

Detection Antibody - 0.2 mg/mL of rabbit anti-HIV-1 gp120 / Glycoprotein 120 monoclonal antibody conjugated to horseradish-peroxidase (HRP) (in PBS, 50 % HRP-Protector, pH 7.4, store at 4°C). Dilute to working concentration of 0.5 µg/mL in detection antibody dilution buffer before use. (Catalog: # 11233-R011)

Standard – Each vial contains 168 ng of recombinant HIV-1 gp120 / Glycoprotein 120. Reconstitute with 1 mL detection antibody dilution buffer. After reconstitution, store at -20°C to -80°C in a manual defrost freezer. A seven-point standard curve using 2-fold serial dilutions in sample dilution buffer, and a high standard of 3000 pg/mL is recommended.

Standard reconstitution tips: Add dilution buffer, gently mix it up and down 3~5 times. Avoid violent and long-time shock.

SOLUTIONS REQUIRED

PBS - 136.9 mM NaCl, 10.1 mM Na₂HPO₄, 2.7 mM KCl, 1.8 mM KH₂PO₄, pH 7.4, 0.2 μ m filtered

Wash Buffer - 0.05% Tween20 in PBS, pH 7.2 - 7.4

Blocking Buffer - 2% BSA in Wash Buffer

Dilution Buffer - 0.1% BSA in wash buffer, pH 7.2 - 7.4, 0.2 µm filtered

Substrate Solution: To achieve best assay results, fresh substrate solution is recommended

Substrate stock solution - 10mg / ml TMB (Tetramethylbenzidine) in DMSO

Substrate dilution buffer - 0.05M Na₂HPO₄ and 0.025M citric acid; adjust pH to 5.5

Substrate working solution - For each plate dilute 250 μ l substrate stock solution in 25ml substrate dilution buffer and then add 80 μ l 0.75% H_2O_2 , mix it well

Stop Solution - 2 N H₂SO₄

PRECAUTION

The Stop Solution suggested for use with this Pair Set is an acid solution. Wear eye, hand, face, and clothing protection when using this material.

STORAGE

Capture Antibody: Aliquot and store at -20°C to -80°C for up to 6 months from date of receipt. Avoid repeated freeze-thaw cycles.

Detection Antibody: Store at 4°C and protect it from prolonged exposure to light for up to 6 months from date of receipt. **DO NOT FREEZE!**

Standard: Store lyophilized standard at -20°C to -80°C for up to 6 months from date of receipt. Aliquot and store the reconstituted standard at -80°C for up to 1 month. Avoid repeated freeze-thaw cycles.

GENERAL ELISA PROTOCOL

Plate Preparation

- 1.Dilute the capture antibody to the working concentration in PBS. Immediately coat a 96-well microplate with 100µL per well of the diluted capture antibody. Seal the plate and incubate overnight at 4°C.
- 2.Aspirate each well and wash with at least 300μ I wash buffer, repeating the process two times for a total of three washes. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining wash buffer by inverting the plate and blotting it against clean paper towels. 3.Block plates by adding $300~\mu$ L of blocking buffer to each well. Incubate at room temperature for a minimum of 1 hour.
- 4.Repeat the aspiration/wash as in step 2. The plates are now ready for sample addition.

Assay Procedure

- $1.Add\ 100\ \mu L$ of sample or standards in sample dilution buffer per well. Seal the plate and incubate 2 hours at room temperature.
- 2. Repeat the aspiration/wash as in step 2 of plate preparation.
- $3.Add\ 100\ \mu L$ of the detection antibody, diluted in antibody dilution buffer, to each well. Seal the plate and incubate 1 hour at room temperature.
- 4. Repeat the aspiration/wash as in step 2 of plate preparation.
- $5.Add\ 200\ \mu L$ of substrate solution to each well. Incubate for 20 minutes at room temperature (if substrate solution is not as requested, the incubation time should be optimized). Avoid placing the plate in direct light.
- 6.Add 50 µL of stop solution to each well. Gently tap the plate to ensure thorough mixing.
- 7.Determine the optical density of each well immediately, using a microplate reader set to 450 nm.

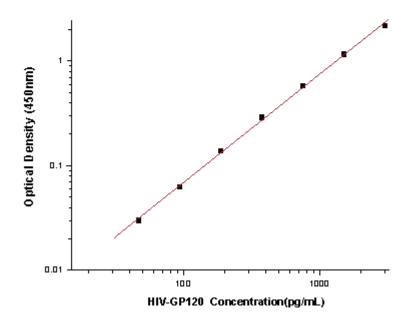
CALCULATION OF RESULTS

- •Calculate the mean absorbance for each set of duplicate standards, controls and samples. Subtract the mean zero standard absorbance from each.
- •Construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph.
- •To determine the concentration of the unknowns, find the unknowns' mean absorbance value on the y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the x-axis and read the concentration. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.
- •Alternatively, computer-based curve-fitting statistical software may also be employed to calculate the concentration of the sample.

TYPICAL DATA

This standard curve is only for demonstration purposes. A standard curve should be generated for each assay.

| Concentration (pg/mL) | Zero standard subtracted OD |
|------------------------|-----------------------------|
| 0 | 0 |
| 46.88 | 0.030 |
| 93.75 | 0.063 |
| 187.5 | 0.139 |
| 375 | 0.291 |
| 750 | 0.580 |
| 1500 | 1.167 |
| 3000 | 2.180 |



PERFORMANCE CHARACTERISTIC

SENSITIVITY

The minimum detectable dose of HIV-1 gp120 / Glycoprotein 120 was determined to be approximately **46.88 pg/ml**. This is defined as at least three times standard deviations above the mean optical density of 10 replicates of the zero standard.

TROUBLE SHOOTING

| Problems | Possible Sources | Solutions |
|------------------------|---|--|
| No signal | Incorrect or no Detection Antibody was added | Add appropriate Detection Antibody and continue |
| | Substrate solution was not added | Add substrate solution and continue |
| | Incorrect storage condition | Check if the kit is stored at recommended condition and used before expiration date |
| Poor Standard Curve | Standard was incompletely reconstituted or was inappropriately stored | Aliquot reconstituted standard and store at -80 °C |
| | Imprecise / inaccurate pipetting | Check / calibrate pipettes |
| | Incubations done at inappropriate temperature, timing or agitation | Follow the general ELISA protocol |
| | Background wells were contaminated | Avoid cross contamination by using the sealer appropriately |
| Poor detection value | The concentration of antigen in samples was too low | Enriching samples to increase the concentration of antigen |
| | Samples were ineffective | Check if the samples are stored at cold environment. Detect samples in timely manner |
| High Background | Insufficient washes | Use multichannel pipettes without touching the reagents on the plate |
| | | Increase cycles of washes and soaking time between washes |
| | TMB Substrate Solution was contaminated | TMB Substrate Solution should be clear and colorless prior to addition to wells |
| | Materials were contaminated. | Use clean plates, tubes and pipettes tips |
| Non-specificity | Samples were contaminated | Avoid cross contamination of samples |
| | The concentration of samples was too high | Try higher dilution rate of samples |

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Notes